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Editorial

The Diptera of Lesotho: a history of collecting in the Mountain Kingdom, summary of recent collecting sites and introduction to the topical collection in African Invertebrates

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Abstract

The Kingdom of Lesotho in southern Africa is entirely bordered by South Africa, with which it shares many geological and vegetation types, but the vast extent of alpine vegetation and high altitudes suggests that differences in insect diversity should be expected. Globally, the Diptera are one of the most diverse orders of insects, though biodiversity estimates for the order are lacking for many regions. Here, we present a summary of the limited historical Diptera collections from Lesotho, summarise our collecting expeditions in 2021, 2022 and 2023, and provide photographs and descriptions of the collecting sites. This paper is the first of a series of papers in a Topical Collection of the Diptera of Lesotho in African Invertebrates. We hope to stimulate more research on Afrotropical Diptera and to improve their conservation in Lesotho in particular.

Key words: Alpine entomology, biodiversity surveys, conservation, Insecta, southern Africa, species checklists, true flies

Introduction

The Kingdom of Lesotho is a small (30 355 km^2), landlocked country surrounded by the Republic of South Africa. While Lesotho shares most vegetation and habitat types with South Africa, the exception being Western Lesotho Basalt Shrubland (Gd9) (Mucina and Rutherford 2006; SANBI 2006), the country has an average altitude ~900 m higher than South Africa, suggesting that differences in the fauna and flora may exist. As can be expected due to its smaller size, Lesotho has only 13 different vegetation types, all within the grassland biome, while South Africa has 466 vegetation types across nine biomes (Mucina and Rutherford 2006; SANBI 2006). Despite the smaller size, Lesotho's higher altitude means that alpine vegetation types are much more extensive (e.g., Drakensberg Afroalpine Heathland) and the country offers habitats for alpine adapted species that are less

extensive in South Africa (Fig. 1). The eastern border of Lesotho is formed by the Drakensberg escarpment, which rapidly drops in altitude into the South African provinces of KwaZulu-Natal and, in the south west, the Eastern Cape. While small patches of alpine vegetation occur on the South African side of the border, these rapidly transition to mid-altitude vegetation. In the south, high altitude grasslands extend from Lesotho into the Eastern Cape, and in the west and north, highveld grassland and shrubland vegetation types occur on both sides of the border, in Lesotho and the Free State province (Mucina and Rutherford 2006; SANBI 2006).

Geologically, the country is similarly limited. The geology is solely comprised of Karoo Supergroup strata, predominantly Drakensberg formation basalts with some exposed sedimentary rocks of the Elliot formation in the river valleys and western lowlands. In the western lowlands, some exposed Tarkastad formation sedimentary rocks and Karoo Dolerite also occur. These four geological types are significantly fewer than the 119 lithographies found in South Africa. This pattern is also reflected in the geomorpholgy, with only the Lesotho Highlands and Eastern Escarpment Hinterland geomorphic provinces represented in the country, compared to 43 geomorphic provinces in South Africa (Partridge et al. 2010).

While there has been widespread collecting of insects, and Diptera in particular, in South Africa, collecting in Lesotho has been less common (Kirk-Spriggs 2017). The combination of regionally rare habitats and comparative under-sampling creates an opportunity for research in Lesotho. With this in mind, three field trips were undertaken: 3 to 14 December 2021, 21 November to 1 December 2022 and 21 to 30 January 2023 by the authors of this introductory article to enhance the Diptera collections from Lesotho.

To increase collection effort without identifying the material within the collections is short-sighted, and so this special collection of articles on the Diptera of Lesotho was conceived. This will turn these specimens into published knowledge of the Diptera of Lesotho and promote conservation in the country. We have chosen to initiate this as a collection of articles rather than a special issue, to allow family accounts to be published as they are completed and thus make the knowledge available more rapidly.

Historic collections

Specimens from Lesotho are in various South African and international collections, but large samples are less common. Expeditions to Lesotho were undertaken by the Lund Zoological Institute (Hanström 1955), Brian and Pamela Stuckenberg, KwaZulu-Natal Museum (NMSA, South Africa), A. Lionel Bevis, Durban Natural Science Museum (DMSA, South Africa), and Charles Jacot-Guillarmod, Albany Museum (AMGS, South Africa) (Kirk-Spriggs 2017).

One of the first major expeditions to include Lesotho was the "Lund University Swedish South African Expedition" of 1950–51 (Brink 1955; Brink and Rudebeck 1955; Hanström 1955; Kirk-Spriggs 2017). Historically, the term "South Africa" has been used to refer to both the Republic of South Africa and (less frequently) the entire Southern African region (e.g., Schultze 1908) and so while the majority of the expedition covered South Africa, collections were also made in Namibia (previously South West Africa) and Lesotho (previously Basutoland) and, to a lesser extent, Eswatini, Zimbabwe and Zambia (previously Swaziland, Southern Rhodesia or Rhodesia, and Northern Rhodesia) (Brink and Rudebeck 1955). The

expedition entered Lesotho twice, first collecting at Qacha's Nek before returning to South Africa and re-entering near Quthing, where they were based for ten days. From there, the expedition travelled by road to Nazareth. The expedition moved by air to Mokhotlong and collections were undertaken in the high mountains on horseback (Brink 1955; Brink and Rudebeck 1955). Twenty-five undescribed Diptera species were collected from Lesotho during this expedition, notably six Asilidae (Hull 1967), four Empididae (Smith 1967) and three Tipulidae (Alexander 1964), all published in the South African Animal Life series.

A.L. Bevis occupied various positions during his tenure at the Durban Museum and Art Gallery and travelled to Lesotho multiple times for collecting (see Clancey 1956; Stuckenberg 1956). These trips were multidisciplinary and not focussed on Diptera collecting, but still resulted in the discovery of undescribed species (e.g., Stuckenberg 1956). About 770 specimens were accumulated during these trips and are housed at the DMSA.

Brian and Pamela Stuckenberg collected at seven sites in Lesotho in 1963 and 1964. These collections generated 884 specimens. Most of the sites were in western Lesotho, though specimens were also collected on God Help Me Pass (Lekhalong La Molimo Nthuse) (on labels as Bushman's Pass) and Blue Mountain Pass (Lekhalong La Thaba Putsoa). These expeditions yielded undescribed species (e.g. Whittington 1992). Along with other specimens accumulated over time, these specimens form the bulk of the collection of approximately 1 100 specimens housed at the NMSA.

Charles Jacot-Guillarmod was Entomologist and later Director of the AMGS. His parents ran a trading post in Ha-Masupha (called Mamathes at the time, now Mamathe or Masupha are also used) and he collected many specimens while visiting them. The majority of his collections were of Hymenoptera, but approximately 1 500 Diptera specimens from this collection are housed at the AMGS.

Recent collecting expeditions

Three expeditions were undertaken to Lesotho (Fig. 1) at six sites in 2021, seventeen in 2022 and seven in 2023 (Figs 2–29). The primary focus of activity was in the eastern highland regions, though some lowland sites were also visited. Details of these sites are given in Table 1. We focused our collecting on the eastern highlands for two reasons. Firstly, the highlands in eastern Lesotho are more extensively covered in alpine vegetation elements (Fig. 1). Secondly, western Lesotho is more densely populated and agricultural pressure is more intense than in eastern Lesotho. This is not to say that environmental degradation is absent from eastern Lesotho. The greatest threats to the environment in all regions of Lesotho are overgrazing (Fig. 30) and agriculture in, or very close to, riparian zones (Figs 11, 12), which lead to soil erosion (Figs 12, 31). Significant threats from mining activities are also present. In general, these are much less extensive than agricultural activities, but pollution impacts may be carried further by streams and rivers. Conservation planning and management should be aware of these threats, though effective management advice is beyond the scope of this paper.

Most collecting was done using sweep netting and hand collecting (Figs 16, 24) but where conditions allowed, Malaise traps were also deployed. Depending on the sites, either Gressitt-and-Gressitt (Fig. 13) or Townes-style (Fig. 20) traps were used. In total, 7 144 specimens were collected in these three expeditions.

Table 1. Collecting sites and coordinates from 2021, 2022 and 2023 in Lesotho. Collecting methods are indicated by: 1— hand and sweep netting, 2— Malaise trapping, 3— light trapping, 4— pan traps. Vegetation types are according to Mucina and Rutherford (2006) and SANBI (2006).

District	Site Name	Expedition(s)	Coordinates	Vegetation type	Comments
Butha-Buthe	Afriski Lodge	2021[1,2], 2022[1,2,4]	28°49.37'S, 28°43.68'E	Drakensberg Afroalpine Heathland (Gd10)	The area around Afriski lodge ranged from degraded to relatively pristine (Fig. 2). There was some grazing evident in places and localised building rubble and litter.
	Afriski, Malibamatšo tributary	2022[1]	28°47.81'S, 28°41.26'E	Lesotho Highland Basalt Grassland (Gd8)	The valley in which this tributary lies was relatively pristine, though there was some evidence of grazing (Fig. 3).
	Moteng Pass	2022[1]	28°44.76'S, 28°36.01'E	Lesotho Highland Basalt Grassland (Gd8)	Moteng Pass is similar to the Malibamatšo tributary, but was drier. Some evidence of grazing was present (Fig. 4).
	Khukhune Roadside drainage gully	2022[1]	28°44.09'S, 28°25.8'E	Eastern Free State Sandy Grassland (Gm4)	The site was in the Khukhune village and badly degraded. Evidence of grazing, alien vegetation (*Acacia mearnsii*, *Eucalyptus* sp. and *Pyracantha koidzumii*) and other habitat modifications was observed (Fig. 5).
Leribe	Motebong Lodge	2021[1,2,3]	29°6.06'S, 28°30.09'E	Lesotho Highland Basalt Grassland (Gd8)	The area around the Motebong lodge was mostly modified gardens with a large impoundment nearby. Some indigenous elements remained in the area, though there was evidence of grazing (Fig. 6).
Mafeteng	Maletsunyane Falls area	2023[1]	29°52.63'S, 28°03.22'E	Lesotho Highland Basalt Grassland (Gd8)	The area around the view point was mostly agricultural land. Some indigenous elements remain on the periphery (Fig. 7). While the waterfall is in Maseru District, the collection site is in Mafeteng District.
Maseru	Between Roma and St Michaels	2022[1]	29°26.19'S, 27°41.04'E	Eastern Free State Sandy Grassland (Gm4)	The area was moderately grazed. Alien vegetation (*Pyracantha koidzumii*) was present (Fig. 8).
	Blue Mountain Pass, along A3	2023[1]	29°28.35'S, 28°01.21'E to 29°27.3'S, 27°58.88'E	Lesotho Highland Basalt Grassland (Gd8)	The area was moderately grazed. Alien vegetation was present (Fig. 9).
	God Help Me Pass, A3, nr Molimo Nthuse Lodge	2023[1]	29°25.39'S, 27°54.33'E	Western Lesotho Basalt Shrubland (Gd9)	Small patch of indigenous forest. Modified garden with mostly alien vegetation, with some indigenous trees (Fig. 10).
	Koro-Koro	2022[1]	29°29.58'S, 27°39.26'E	Basotho Montane Shrubland (Gm5)	Agriculture and grazing were evident. Alien vegetation was present in lower areas. On the steeper slopes the shrubland was more intact, but still grazed (Fig. 11).
	Matsaba Nkesi village	2022[1]	29°40.14'S, 27°47.84'E	Western Lesotho Basalt Shrubland (Gd9)	Fallow agricultural fields in wetland areas. Some alien plants on the steeper slopes (Fig. 12).
	Roma Trading Post Lodge	2022[1,2]	29°26.59'S, 27°42.22'E	Peri urban garden	A highly modified garden site (Fig. 13).
	Semonkong area, A5	2023[1]	29°53.30'S, 28°06.5'E	Lesotho Highland Basalt Grassland (Gd8)	A highly modified garden site (Fig. 14).
	Semonkong Lodge, gardens and vegetation	2023[1]	29°50.6'S, 28°2.6'E	Lesotho Highland Basalt Grassland (Gd8)	A highly modified garden site (Fig. 15).
	St Michaels village	2022[1]	29°25.72'S, 27°40.47'E	Eastern Free State Sandy Grassland (Gm4)	Grazing and alien vegetation (*Acacia mearnsii*, *Pyracantha koidzumii*) was present, though fields with indigenous vegetation were extensive. Human settlement in close proximity to the wetland and stream (Fig. 16).
Mokhotlong	Roadside flowers at stream	2021[1]	29°27.56'S, 29°8.77'E	Lesotho Highland Basalt Grassland (Gd8)	Modified habitat, a culvert under the road. Some grazing was evident (Fig. 17).
	Roadside flowers at swamp	2021[1]	29°26.37'S, 29°7.94'E	Lesotho Highland Basalt Grassland (Gd8)	Modified habitat, a culvert under the road. Some grazing was evident.
	Roadside flowers in Khatleli village	2021[1]	29°16.71'S, 28°56.55'E	Lesotho Highland Basalt Grassland (Gd8)	Roadside flowers next to large puddle. Modified village habitat (Fig. 18).

District	Site Name	Expedition(s)	Coordinates	Vegetation type	Comments
Qacha's Nek	Sehlabathebe National Park, new lodge	2023[1,2]	29°52.71'S, 29°4.45'E	Lesotho Highland Basalt Grassland (Gd8)	Pristine grassland, minimal impact of grazing or alien species (Figs 19, 20).
	Sehlabathebe National Park, old lodge	2023[1]	29°52.07'S, 29°7.13'E	Lesotho Highland Basalt Grassland (Gd8)	Pristine grassland, minimal impact of grazing or alien species (Fig. 21).
Quthing	Fallow lands near Letšeng-la-Letsie	2022[1]	30°18.54'S, 28°7.98'E	Lesotho Highland Basalt Grassland (Gd8)	Fallow lands with various annual flowers (Fig. 22).
	Letšeng-la-Letsie	2022[1]	30°18.77'S, 28°10.06'E	Lesotho Highland Basalt Grassland (Gd8)	Intensely grazed habitat. Very little impact from alien vegetation but degradation due to grazing evident (Fig. 23).
	Mphaki Cell towers	2022[1]	30°11.55'S, 28°8.23'E	Lesotho Highland Basalt Grassland (Gd8)	Hill top in Mphaki town. Some peri urban development and grazing (Fig. 24).
	Mphaki Farmers Training Centre	2022[1,2,4]	30°11.60'S, 28°07.83'E	Lesotho Highland Basalt Grassland (Gd8)	Modified garden with alien vegetation, vegetable gardens and fruit trees (Fig. 25).
	Mphaki Roadside	2022[1]	30°13.21'S, 28°08.32'E	Lesotho Highland Basalt Grassland (Gd8)	Relatively pristine site, some evidence of grazing and agriculture (Fig. 26).
	Roadside seep/stream	2022[1]	30°12.88'S, 28°08.31'E	Lesotho Highland Basalt Grassland (Gd8)	Relatively pristine site, some evidence of grazing and agriculture.
	Roadside swamp	2022[1]	30°13.69'S, 28°08.44'E	Lesotho Highland Basalt Grassland (Gd8)	Relatively pristine site, some evidence of grazing and agriculture (Fig. 27).
	Wetland N of Letšeng-la-Letsie	2022[1]	30°17.03'S, 28°08.14'E	Lesotho Highland Basalt Grassland (Gd8)	Relatively pristine site, some evidence of grazing and agriculture (Fig. 28).
Thaba-Tseka	Sani area	2021[1]	29°35.05'S, 29°17.12'E	Lesotho Highland Basalt Grassland (Gd8)	Moderate to extensive evidence of grazing. Degraded habitat (Fig. 29).

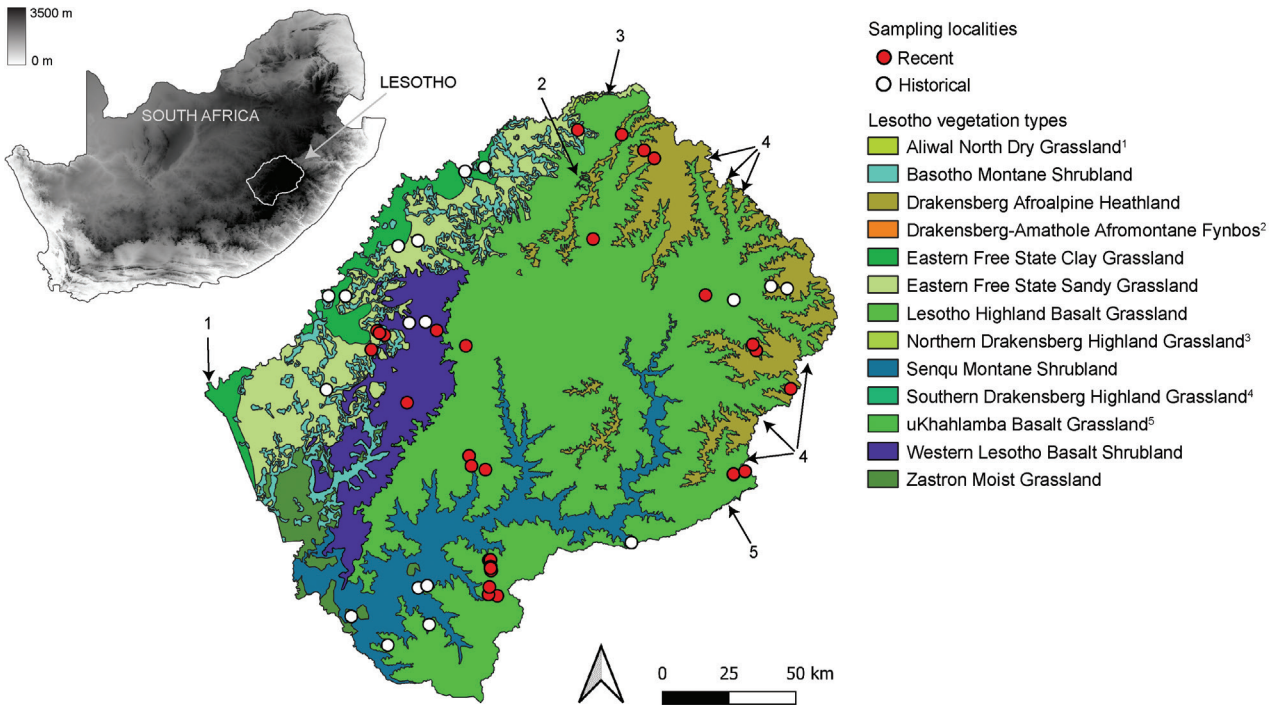
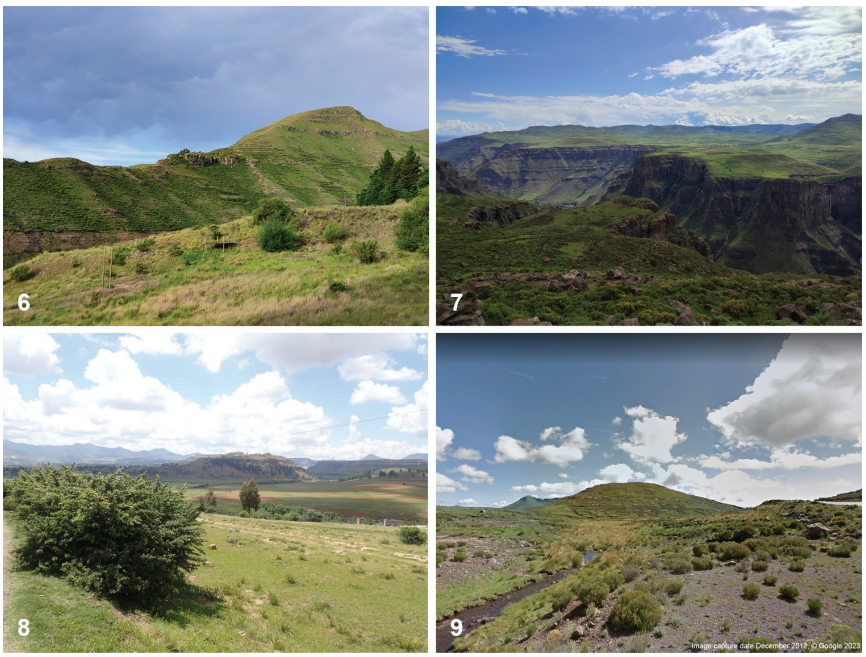


Figure 1. Map of major collecting sites in Lesotho with vegetation types. White indicates historical collecting sites and red indicates sites from recent expeditions (2021, 2022 and 2023). Vegetation data taken from SANBI (SANBI 2006). Arrows 1–5 indicate vegetation types with extents too small to be clearly visible on the map, as indicated on the legend.

Figures 2–5. Collecting sites during recent expeditions **2** Afriski Mountain Lodge area, looking southwest down the valley. Large areas of grazed but otherwise undisturbed landscape and disturbed piles of building rubble are visible **3** valley landscape around the tributary of the Malibamatšo River, looking southwest from the A1 road. While some grazing is evident, the landscape is otherwise undisturbed **4** view from Moteng Pass on the A1, looking southeast from near the top of the pass. The habitat is drier than surrounding sites **5** gulley in Khukhune village next to the A1. The area is disturbed, with evidence of grazing and multiple alien plant species (*Acacia mearnsii*, *Eucalyptus* sp. and *Pyracantha koidzumii* are visible in the photograph).

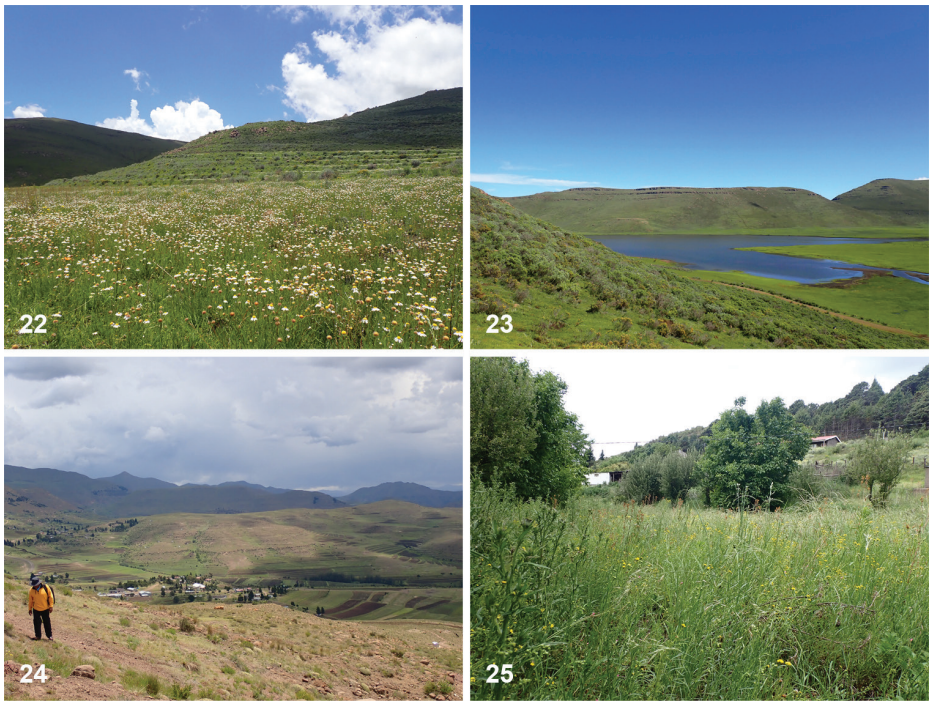


Figures 6–9. Collecting sites during recent expeditions **6** Motebong Lodge, looking east. Areas close to the lake are disturbed by fluctuating water levels and increased grazing pressure. Many non-native ornamentals were planted around the lodge **7** Maletsunyane Falls area, sampling area consisted of natural vegetation and is cordoned off with palisade fencing from the surrounding farmlands, protecting it somewhat from overgrazing **8** roadside *Pyracantha koidzumii* bushes on the A5 between Roma and St Michaels village, looking south. The bushes attract flies, disrupting indigenous pollination networks **9** Blue Mountain Pass, roadside rest stop on A3, moderately grazed, toilet paper and human faeces scattered near the road, less disturbance away from the road.

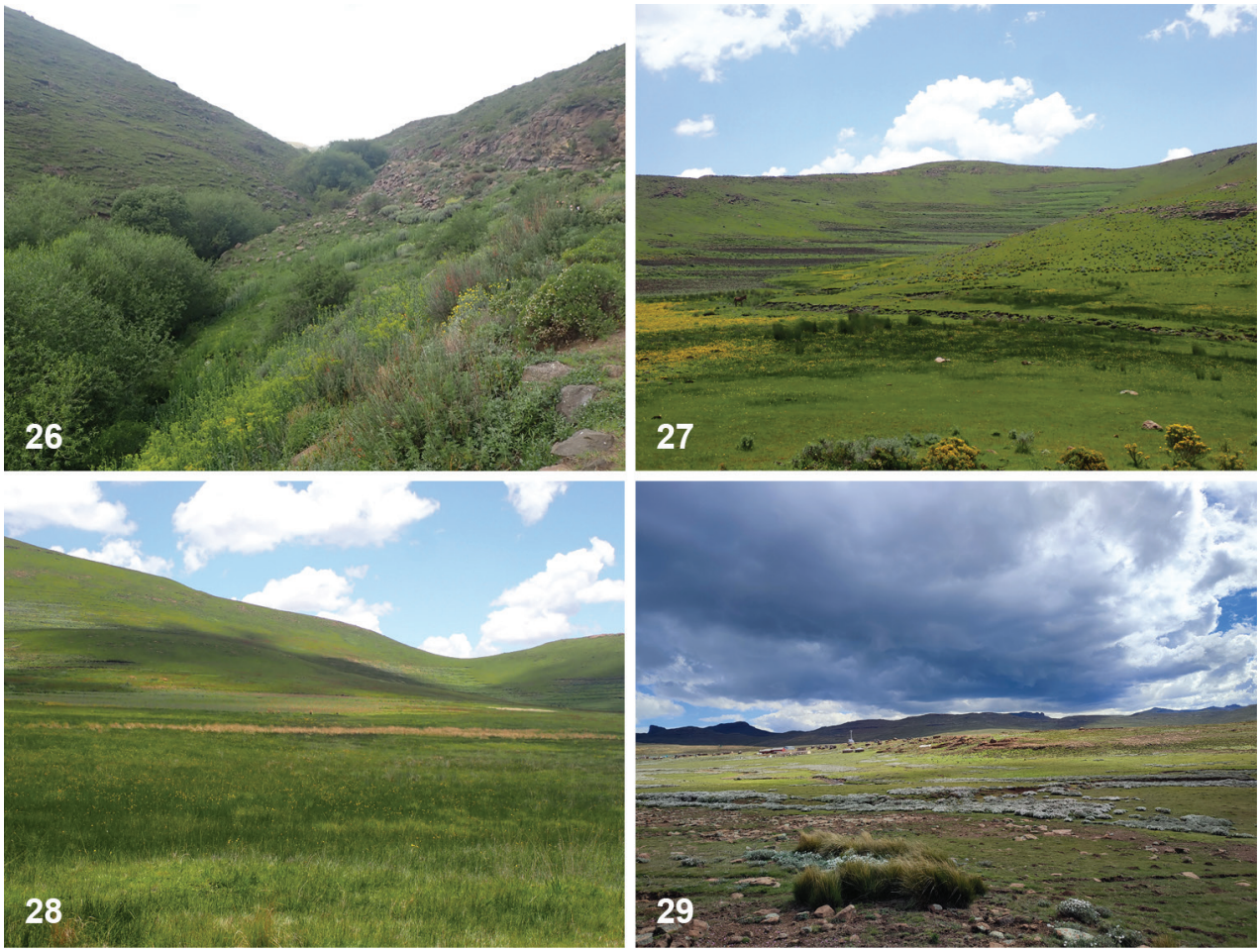
Figures 10–13. Collecting sites during recent expeditions **10** God Help Me Pass, Molimo Ntuse Lodge area on A3, indigenous and exotic trees, with grass elements **11** hillside and fields near Koro-Koro village, looking south. Flat lowland areas extensively cultivated and grazed, resulting in erosion as seen in the bottom left of the photograph. Steeper-sloped areas receive less grazing pressure, with small shrubs and other plants surviving **12** fallow and recently ploughed fields at Matsaba Nkesi, looking west from the A5 road. The dark area in the centre of the image is a seep that has been cultivated. The seep and surrounding areas included many *Ranunculus* sp. flowers, while the steeper sides have alien vegetation including *Pyracantha koidzumii* and *Rosa rubiginosa* **13** malaise trap on the grounds of Roma Trading Post Lodge. The site contained many exotic ornamental plants.

Figures 14–17. Collecting sites during recent expeditions **14** Semonkong roadside, brief sampling from shrubs and grasses on verge **15** Semonkong Lodge, mixed indigenous and exotic vegetation with some garden elements **16** seep and grassland at St Michaels village. Some *Ranunculus* sp. visible in foreground. Alien *Acacia mearnsii* visible in the background. Note the proximity of the latrine (top right) to the watercourse **17** stream next to the A14. Some grazing and disturbance due to road building.

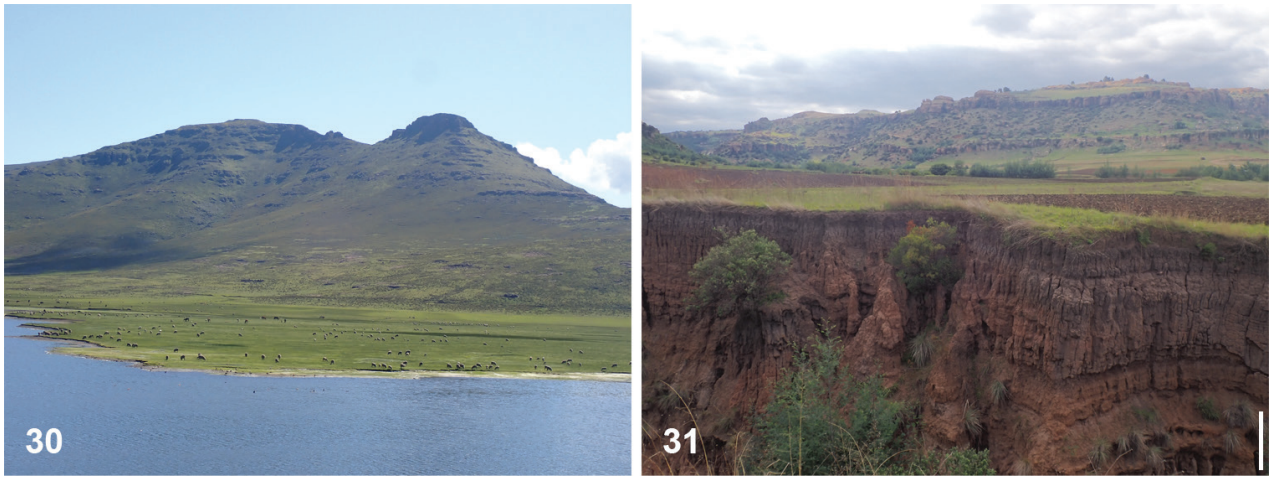
Figures 18–21. Collecting sites during recent expeditions **18** puddle and flowers in Khatleli village on the A1. Environment is heavily degraded, with agriculture and domestic disturbances evident **19** Sehlabathebe National Park, grassland vegetation near the new lodge and camping area **20** Sehlabathebe National Park, Malaise trap over small stream near new lodge area **21** Sehlabathebe National Park, semi-wetland area near old lodge, sampling from grass and shrubs.



Figures 22–25. Collecting sites during recent expeditions **22** fallow field near Letšeng la Letsie (Letsie's Lake). Diverse annual flowers growing in a highly modified habitat **23** Letšeng la Letsie view looking northwest. The site is the largest water source in the area, providing a suitable environment for livestock and therefore heavily influenced by grazing **24** hill above Mphaki, looking westwards. The site is on the edge of town and has some domestic buildings and other disturbances **25** Mphaki Farmers Training Centre garden. Vegetation is mostly exotic: alien (*Cirsium* sp. visible), ruderal or agricultural (*Juglans* sp. and *Prunus* sp. visible).



Figures 26–29. Collecting sites during recent expeditions **26** steep roadside cutting with exposed rocks and flowering *Scenecio* sp. Alien species are evident, including *Salix* sp. growing in riparian zone and *Rosa rubiginosa* on the steeper slopes **27** seep with *Ranunculus* sp. flowers. Grazing and other agriculture evident in the image **28** large high-altitude wetland with *Ranunculus* sp. flowers. Evidence of agriculture is visible in the background **29** Sani Pass area, overgrazing and other human disturbance visible.



Figures 30, 31. Evidence of land degradation during recent expeditions **30** high grazing density at Letšeng la Letsie. Cattle, sheep, goats, horses and donkeys were observed **31** deep erosion gullies at Koro-Koro. Poor agricultural practices are the primary cause, for example ploughing of fields in the riparian zone (see fields in the background and in figs 11, 12). Scale bar: 1 m (**31**).

Table 2. Summary of families in major South African Diptera collections, both historic (AMGS, BMSA, DMSA, NMSA) and from recent collection trips (BMSA, NMSA). Higher classification follows Wiegmann et al. (2011), family sequence follows Kirk-Spriggs and Sinclair (2017a, b, 2021, in prep).

Higher Classification	Family	Historic Collections	Recent collection	Total
Tipulomorpha	Tipulidae	146	1	147
Psychodomorpha	Blephariceridae	19	1	20
Bibionomorpha	Psychodidae	0	8	8
	Bibionidae	2	35	37
	Keroplatidae	0	1	1
	Mycetophilidae	0	474	474
	Sciaridae	0	14	14
	Cecidomyiidae	0	7	7
	Scatopsidae	0	2	2
Culicomorpha	Culicidae	17	35	52
	Simuliidae	5	143	148
	Ceratopogonidae	0	13	13
	Chironomidae	7	16	23
Tabanomorpha	Vermileonidae	7	0	7
	Rhagionidae	9	122	131
	Athericidae	3	0	3
	Tabanidae	89	56	145
Stratiomyomorpha	Stratiomyidae	75	146	221
Unplaced Brachycera	Acroceridae	3	3	6
Xylophagomorpha	Nemestrinidae	29	3	32
Asiloidea	Bombyliidae	165	59	224
	Mythicomyiidae	0	3	3
	Asilidae	528	46	574
	Therevidae	3	3	6
	Scenopinidae	2	0	2
Empidoidea	Empididae	110	488	598
	Hybotidae	0	98	98
	Dolichopodidae	69	345	414
	Platypezidae	1	2	3
	Lonchopteridae	1	24	25
Phoroidea	Phoridae	1	6	7
Unplaced Cyclorrhapha	Syrphidae	318	490	808
	Pipunculidae	7	12	19
	Agromyzidae	19	88	107
	Periscelididae	0	1	1
	Sphaeroceridae	10	329	339
Acalypterata	Diopsidae	2	0	2
	Sepsidae	36	105	141
	Clusiidae	0	1	1
	Conopidae	15	4	19
	Sciomyzidae	6	2	8
	Lonchaeidae	13	46	59
	Platystomatidae	9	4	13

Higher Classification	Family	Historic Collections	Recent collection	Total
Acalypterata	Tephritidae	264	150	414
	Pyrgotidae	1	1	2
	Lauxaniidae	131	499	630
	Chamaemyiidae	61	106	167
	Milichiidae	32	1	33
	Chloropidae	72	284	356
	Chyromyidae	0	2	2
	Heleomyzidae	3	47	50
	Ephydridae	82	285	367
	Curtonotidae	3	11	14
	Diastatidae	0	2	2
	Drosophilidae	10	66	76
	Cryptochaetidae	1	1	2
Calypterata	Scathophagidae	65	37	102
	Anthomyiidae	30	150	180
	Fanniidae	1	33	34
	Muscidae	394	1770	2164
	Calliphoridae *sensu strictu*	45	23	68
	Rhiniinae	17	62	79
	Sarcophagidae	266	36	302
	Rhinophoridae	1	14	15
	Tachinidae	228	328	556
	Oestridae	3	0	3
Totals		3436	7144	10580

Contributions towards this special collection

In all, historic South African collections and the recent expeditions have generated a collection of approximately 10 580 Diptera specimens from Lesotho (Table 2), excluding the material from the Lund Expedition. Despite this, there is little published information on the Diptera of Lesotho. That which is published is mostly in the South African Animal Life series, based on the Lund expedition (e.g., Alexander 1964; Hull 1964, 1967; Smith 1967). Other publications are fragmented and sporadic. The aim of initiating this Topical Collection in African Invertebrates is to bring this information together in one series, creating a resource for a diverse set of users. While we endeavoured to collect a diverse group of specimens, it is inevitable that some groups were sampled more thoroughly than others. This collection should be seen as a starting point for future research in Lesotho. Despite this, it is still of value for conservation practitioners in the region as it is the first attempt at a consolidated Diptera checklist for the country. The details of the collecting sites provided in Table 1 and the site photographs (Figs 2–29) are also intended to pre-empt repetition in future contributions to this Special collection but also allows future collectors to plan their trips with existing material in mind.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

BSM confirms ethical clearance, number NMB ECC 2022/02, forming part of project 542 of the National Museum, Bloemfontein, South Africa.

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Author contributions

Bellingan, Jordaens, Midgley and Muller conceptualized the project. Phoofolo, Jordaens, Midgley, Muller and Theron joined the fieldwork. Midgley and Muller wrote the first draft of the manuscript. Bellingan, Phoofolo, Jordaens and Theron commented on the draft.

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Data availability

All of the data that support the findings of this study are available in the main text.

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Checklist

Checklist of the spiders (Araneae) of South Africa

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Abstract

A checklist of 2265 spider species and subspecies, 495 genera and 71 families is provided. Data were extracted from the South African National Survey of Arachnida database and over 200 taxonomic revisions, ending December 2022. Global distributions, endemicity and conservation assessment using IUCN Criteria are provided for each species. A total of 1325 spp. are endemic to South Africa (58.5%), 126 spp. (5.6%) are of special concern and 693 spp. (30.6%) are Data Deficient (DD), while 15 species were described without exact locality data. Most species (1444 spp., 63.8%) are widely distributed with no known threats and are of Least Concern. A total of 1316 spp. (57.6%) are known from both sexes and 23 spp. (1.0%) were described from juveniles. Salticidae is the most species-rich family (354 spp.), followed by Gnaphosidae (195 spp.), Thomisidae (143 spp.) and Araneidae (100 spp.) and ten families are represented by a single species.

Key words: Conservation status, endemicity, faunistic surveys, global distribution, provincial, sample bias, SANSA, South African National Survey of Arachnida

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Introduction

The emerging field of conservation biogeography concerns species' distribution dynamics and how they relate to biodiversity conservation (Robertson et al. 2010); and its main currency is valid species-level determinations and distribution data. However, the knowledge and insights gained from these data depend on whether species are valid and systematically established, the existence of clear diagnoses to aid positive determinations, documented biology and reconstructed evolutionary histories. These activities depend on the primary data collected with specimens that provide vast information (Hamer 2012). Therefore, curating and disseminating this information in retrievable formats, such as a national list, is essential.

The South African National Survey of Arachnida (SANSA), which was initiated in 1997 (Dippenaar-Schoeman et al. 2015), had the main aim of documenting the arachnid fauna of South Africa by collating all the data into a relational database. The second phase of SANSA (2006–2010), funded by the Royal

Norwegian Ministry through the Threatened Species Programme of the South African National Biodiversity Institute (SANBI), was critical, as it surveyed large under-represented parts of South Africa (Dippenaar-Schoeman et al. 2015), culminating in the release of an unpublished report: the First Atlas of the Spiders of South Africa (FASSA) in 2010 (Dippenaar-Schoeman et al. 2010). Until now, this Atlas was the only known national list of South African spiders.

Only a few African countries have published spider species checklists: Botswana (Eagle 1985), Namibia (Griffin and Dippenaar-Schoeman 1991), Zimbabwe (FitzPatrick 2001), Sudan (Dunlop and Siyam 2014), Tanzania (Russell-Smith 2020) and Kenya (Kioko et al. 2021). Here, we provide the first annotated spider checklist for South Africa, based on published data and accessioned specimens in collections up to the end of December 2022. We also include references to unpublished resources that add value to the data and results summarised here.

Material and methods

Study area

The area covered is South Africa (Fig. 1A), excluding the enclave of Lesotho and Eswatini. It includes the following nine Provinces: Eastern Cape (EC), Free State (FS), Gauteng (G), KwaZulu-Natal (KZN), Limpopo (L), Mpumalanga (M), Northern Cape (NC), North West (NW) and Western Cape (WC).

Source of information

As part of SANSA, all available data were collated in a relational database maintained by the Agricultural Research Council (Dippenaar-Schoeman et al. 2012). This database is not publicly accessible, but the data retrieved from this database and used for this manuscript are available online in a tidy format (https://zenodo.org/record/8249595). These data were used to identify the gaps in the geographical coverage and guide the field surveys of phase II of SANSA, which were conducted using a standardised protocol (Haddad and Dippenaar-Schoeman 2015). All the specimens from postgraduate ecological surveys from 12 universities in South Africa were deposited in the National Collection of Arachnida (**NCA**) at the Agricultural Research Council in Pretoria as voucher specimens. Many specimens were also received from the public participating in SANSA surveys, including members of the Spider Club of Southern Africa and Reserve managers. A large number of specimens were collected during national and international projects included in unpublished studies, but included in SANSA newsletters. However, these newsletters went through an editorial process and most are available online (Suppl. material 1). Voucher specimens of sampled material were also deposited National Museum in Bloemfontein, Ditsong National Museum of Natural History in Pretoria and KwaZulu-Natal Museum in Pietermaritzburg.

Checklist structure

The checklist is based on data extracted from the SANSA database and references to the taxonomic and more than 200 published/unpublished faunistic surveys up to the end of December 2022 (Suppl. material 1). Only spiders

identified to species level are included. Families and species are arranged alphabetically. For each species, we provide:

Endemicity

The following terms are used: **SAE**: species endemic to South Africa; **STHE**: species endemic to southern Africa; **AE**: species endemic to the Afrotropical Region; **C**: species that also occur beyond the Afrotropical Region.

Distribution

The following terms are used: 6: species is known only from the type locality in South Africa; 5: species is known from several localities in the same province as the type locality; 4: species were sampled from two adjacent provinces; 3: species were sampled from ≥ three provinces in South Africa; 2: species occurs outside South Africa, but within southern Africa; 1: Afrotropical Region; 0: cosmopolitan.

Provincial data

Species listed per South African province obtained from primary label data of collected specimens, as reported in taxonomic and faunistic publications and the SANSA database.

Conservation assessment

As part of the Spider Red Listing Project (Foord et al. 2020), the preliminary conservation status of species as determined are listed with the following codes: DD (Data Deficient): species usually known from only one sex or based on old material without detailed locality data and where the species is difficult to identify (DDT: Data Deficient due to Taxonomic reasons); LC (Least Concern): species with a broad distribution (categories 0–2), without known threats; those of categories 3 and 4 are South African endemics (SAE) and many of them are also LC. Species of special concern (Rare, Critically Rare, Vulnerable, Threatened) usually belong to categories 5 or 6.

History

Almost a third of South African species were described from 1900–1920 (739 spp.) (Fig. 1B), focusing largely on the fauna of the coastal provinces, as most practising arachnologists were stationed there. From 1960 to 1980, there was a considerable decline in the description of new species due to lack of practising taxonomists. However, during the last three decades there has been a resurgence; more than 700 species have been described since 1980, mainly due to new appointments, modern taxonomic revisions, the training of several South African taxonomists and the efforts of taxonomists overseas. Ongoing SANSA surveys continued contributing accessions at the NCA and specimen identifications have also contributed to a constant increase of new species records (Fig. 1B). The increase in taxonomic output since 1997 can partly be attributed to SANSA (Dippenaar-Schoeman et al. 2015).

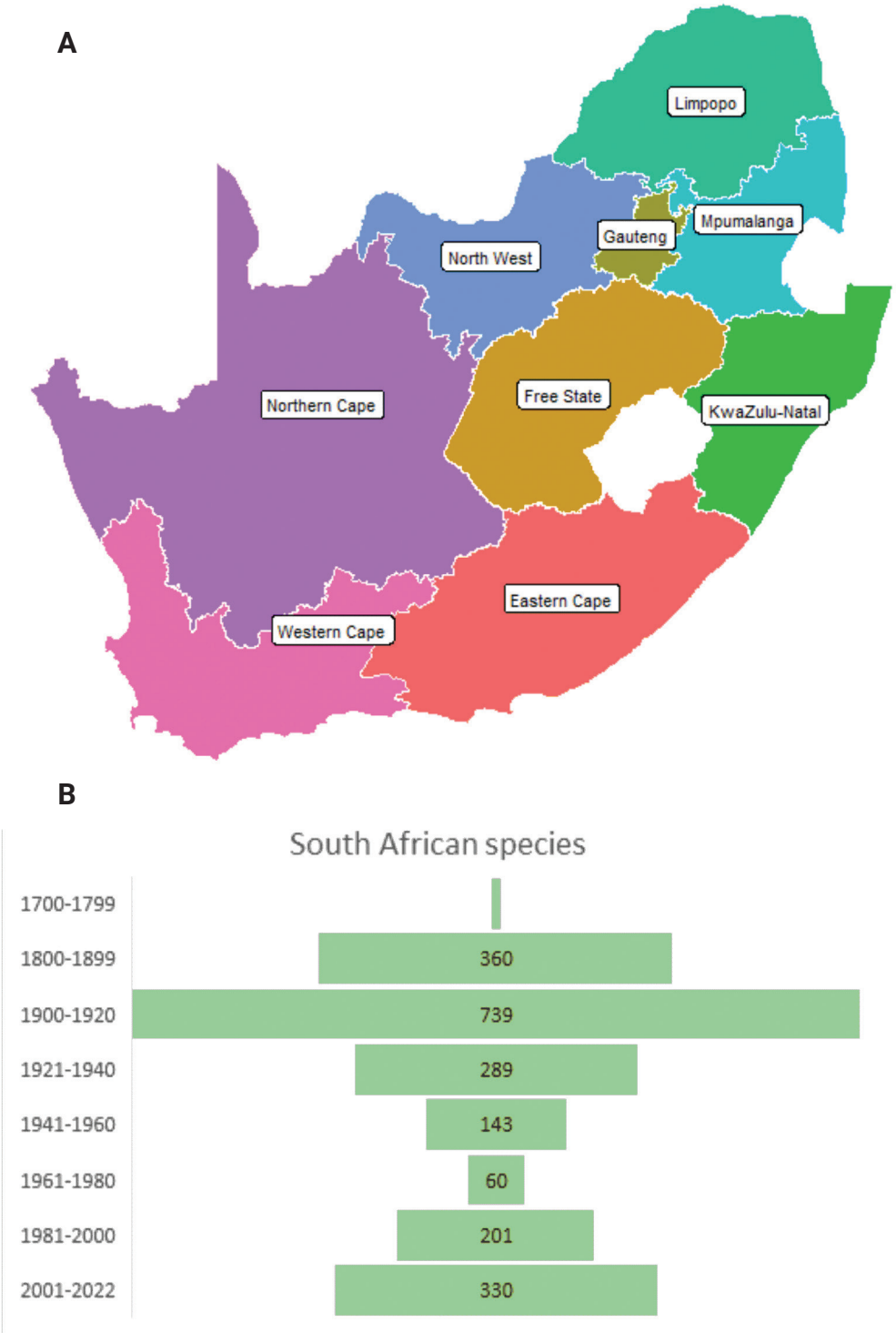

Figure 1. A Map of South Africa and its nine provinces **B** temporal trends in species described from South Africa (1700–2023).

Species identification

Unfortunately, the original descriptions of many species described from 1700–1950 are often rudimentary, only from one sex, lacking detailed morphological data and without drawings or detailed collecting data. Most type specimens are also housed in overseas museums or are lost. One of the most significant constraints for SANSA is the lack of taxonomic revisions for many of the larger

spider families in South Africa, with reference to Agelenidae, Araneidae, Dictynidae and Theridiidae. The number of South African species will inevitably increase, as many specimens housed in collections remain unidentified due to lack of taxonomic resolution.

With all the available data, SANSA has developed online photo identification guides for the 72 families to address the taxonomic constraints (Suppl. material 2). These guides list known information for all genera and species listed from South Africa. Species-level information includes a distribution map for South Africa, drawings (if available) and photographs of diagnostic characteristics, notes on their behaviour, a conservation assessment and possible threats. Complete guides can be downloaded from the World Spider Catalogue, as well as from Zenodo (https://zenodo.org/communities/sansa/) (for complete list see Suppl. material 2).

Results and discussion

A total of 2265 spider species and subspecies, belonging to 495 genera and 71 families, are presently known from South Africa (Table 1). The Salticidae (354 spp.), Gnaphosidae (195 spp.), Thomisidae (143 spp.) and Araneidae (100 spp.) are the most species-rich families. A total of nine families are only represented by a single species.

Table 1. Spider families from South Africa listing the number of genera and species per family, as well as the number endemic to South Africa (SAE), the number of Data Deficient (DD) species, the species of Least Concern (LC) and the number of species of Special Concern (SC).

FAMILIES	GEN	SPP	SAE	DD	LC	SC
Agelenidae	5	10	0	0	10	0
Amaurobiidae	5	16	16	11	3	2
Anapidae	3	4	3	2	1	1
Anyphaenidae	1	1	0	0	1	0
Araneidae	40	100	20	7	90	3
Archaeidae	1	14	14	6	2	6
Atypidae	1	2	2	0	0	2
Barychelidae	2	3	0	0	3	0
Bemmeridae	2	33	32	30	3	0
Caponiidae	3	16	10	6	8	2
Cheiracanthiidae	3	47	36	20	26	1
Cithaeronidae	1	1	1	1	0	0
Clubionidae	1	26	17	12	14	0
Corinnidae	15	44	22	11	28	5
Ctenidae	2	8	3	1	7	0
Cyatholipidae	6	16	16	4	7	5
Cyrtaucheniidae	1	29	28	17	12	0
Deinopidae	2	7	1	0	6	0
Desidae	2	2	0	0	2	0
Dictynidae	4	5	2	1	4	0
Drymusidae	1	5	5	1	0	4
Dysderidae	1	1	0	0	1	0

FAMILIES	GEN	SPP	SAE	DD	LC	SC
Entypesidae	5	30	30	17	12	1
Eresidae	5	30	16	11	19	0
Euagridae	2	5	4	1	4	0
Filistatidae	1	1	0	0	1	0
Gallieniellidae	2	29	29	13	13	3
Gnaphosidae	28	195	106	48	147	0
Hahniidae	1	8	5	2	6	0
Hersiliidae	3	12	4	1	10	1
Idiopidae	6	46	44	26	15	5
Ischnothelidae	1	1	0	0	1	0
Linyphiidae	23	32	16	7	23	2
Liocranidae	3	11	8	5	6	0
Lycosidae	24	113	61	40	72	1
Microstigmatidae	1	6	6	2	3	1
Migidae	2	22	20	11	6	5
Mimetidae	3	4	1	0	4	0
Miturgidae	2	2	2	1	1	0
Mysmenidae	1	1	1	1	0	0
Nephilidae	2	5	0	0	5	0
Nesticidae	1	1	0	0	1	0
Oecobiidae	4	7	3	1	6	0
Oonopidae	9	17	12	7	10	0
Orsolobidae	3	4	3	2	2	0
Oxyopidae	3	40	3	1	39	0
Palpimanidae	3	24	19	14	8	2
Philodromidae	6	34	6	4	30	0
Pholcidae	8	46	34	6	20	20
Phyxelididae	9	30	27	10	16	4
Pisauridae	14	38	8	4	34	0
Prodidomidae	6	29	17	12	17	0
Pycnothelidae	1	1	1	0	0	1
Salticidae	80	354	177	96	244	11
Scytodidae	1	30	24	11	17	2
Segestriidae	1	15	13	7	8	0
Selenopidae	2	71	52	23	47	1
Sicariidae	2	14	9	4	9	1
Sparassidae	9	50	35	12	35	3
Stasimopidae	1	44	44	37	3	4
Symphytognathidae	2	2	2	2	0	0
Telemidae	1	1	1	0	0	1
Tetragnathidae	7	31	10	6	24	1
Theraphosidae	8	40	34	18	19	3
Theridiidae	19	59	27	17	42	0
Thomisidae	38	143	36	10	130	3
Trachelidae	12	55	45	25	28	4
Trochanteriidae	1	11	6	5	6	0
Uloboridae	5	10	4	1	9	0
Zodariidae	21	97	67	29	58	10
Zoropsidae	2	25	25	13	7	5
	495	2265	1325	692	1445	126
		%	58.55	30.62	63.81	5.57

Provincial surveys

The number of families, genera and species for each of the nine provinces are listed in Table 2. KwaZulu-Natal has the highest number of known species (1024), followed by the Western Cape (908) and Limpopo Province (905), while North West had the least (344). Note: almost all the surveys without citations listed below, can be found in the grey literature, references and DOIs to these documents being included in Suppl. material 1 of the supplementary material.

Eastern Cape (Fig. 2A): The Eastern Cape becomes progressively wetter from west to east. The west is mostly semi-arid Karoo, except in the far south, where there is a temperate rainforest in the band between the Cape Fold Mountains and the Indian Ocean. The coast is generally rugged with interspersed beaches. Most of the Province is hilly to very mountainous to the west of East London and Queenstown and towards the KwaZulu-Natal border – a region known previously as Transkei – it is a lush grassland on rolling hills, punctuated by deep gorges with intermittent forest. A total of 837 species (137 endemic) have been recorded from the Province (Table 2).

Major surveys in this Province include: Addo Elephant National Park (Dippenaar-Schoeman et al. 2020); Amathole Mountains (Haddad et al. 2023); Asante Sana Private Game Reserve; Jeffreys Bay; Mkambati Nature Reserve (Dippenaar-Schoeman et al. 2011); Mountain Zebra National Park (Dippenaar-Schoeman 1988, 2006); Nduli and Luchaba Nature Reserves (Niba and Yekwayo 2016); Silaka Nature Reserve (Forbanka and Niba 2013); Thyspunt; Tsolwana Game Reserve.

Free State (Fig. 2B): The Free State is situated on the central escarpment and is comprised of a succession of flat grassy plains sprinkled with pastures and cultivated lands, resting on a general elevation of 1200 m, only broken by the occasional hill. The rich soil and pleasant climate allow for a thriving agricultural industry. It is known locally as the breadbasket of South Africa with more than 30,000 farms, which produce over 70% of the country's grain.

The Free State is the third largest province (10.6%) and a total of 517 species (25 endemic) have been recorded here (Table 2).

Major surveys in the Province include: Amanzi Private Game Reserve (Haddad and Butler 2018); Farm Bankfontein, Luckhof District (Haddad 2021); Farm Deelhoek, Bloemfontein District (Haddad and Dippenaar-Schoeman 2002, 2006a; Haddad 2005); Farm Lusthof, Edenville District (Dippenaar-Schoeman et al. 1978);

Table 2. Number of families, genera, species and endemics recorded for each of the nine provinces in South Africa, as well as the percentage of all the known South African species.

Provinces	Family	Genera	Species	Endemics	% spp. total SA
Eastern Cape	59	312	837	137	37
Free State	50	224	517	25	24
Gauteng	52	250	563	11	25
KwaZulu-Natal	60	362	1024	159	45
Limpopo	60	346	905	55	41
Mpumalanga	57	279	645	20	28
North West	41	188	344	6	15
Northern Cape	52	236	559	62	25
Western Cape	60	319	908	261	41

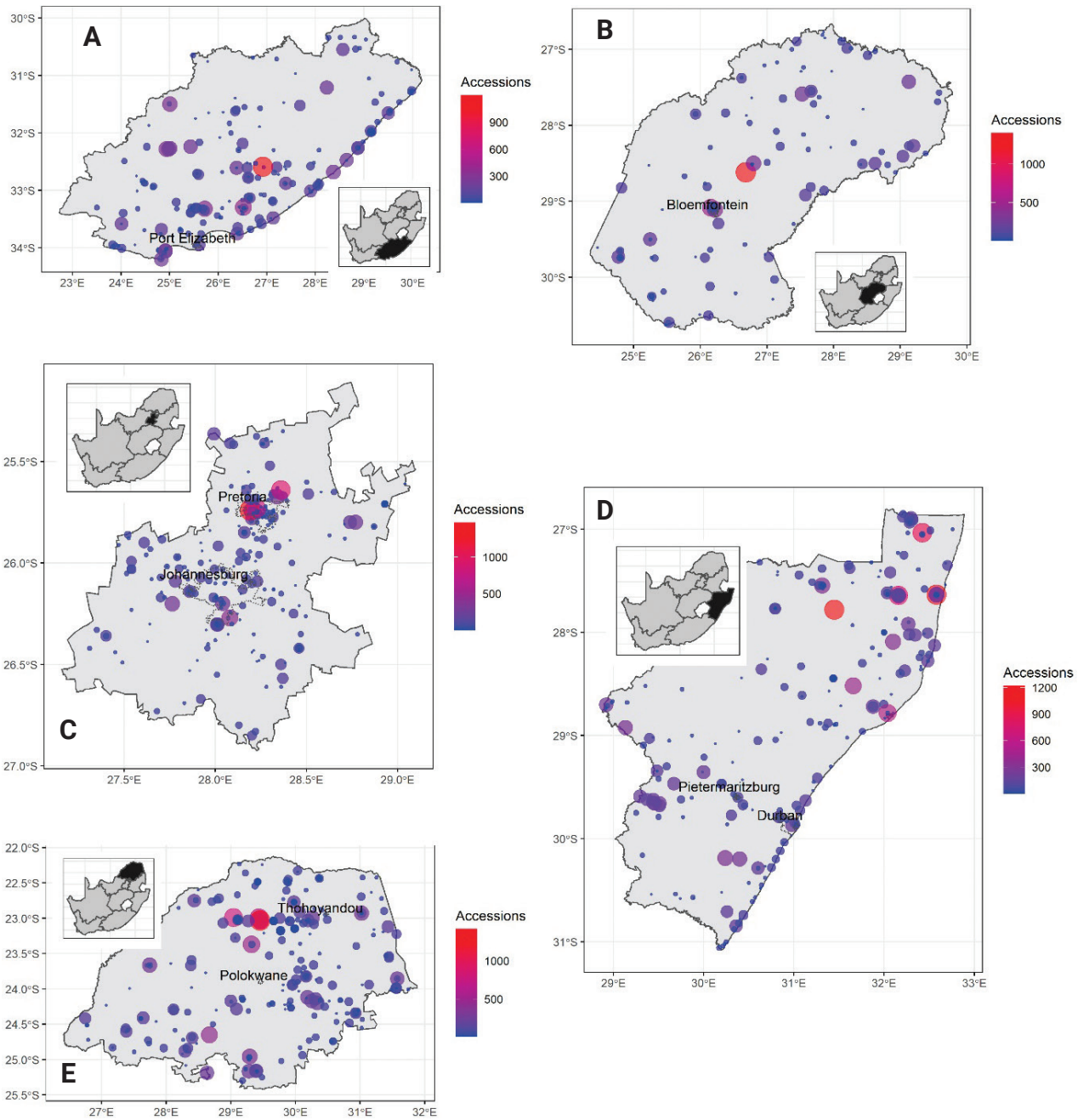

Figure 2. Spatial distribution and number of accessions for localities in: **A** Eastern Cape **B** Free State **C** Gauteng **D** KwaZulu-Natal **E** Limpopo.

Erfenisdam Nature Reserve (Fourie et al. 2013; Haddad et al. 2015); Florisbad Research Station (Lotz et al. 1991); Free State National Botanical Gardens (Butler and Haddad 2011; Neethling and Haddad 2013, 2019; Haddad et al. 2019, 2021; Luwes and Haddad 2020); Golden Gate Highlands National Park (van der Merwe et al. 2020); Kalkfontein Dam Nature Reserve, and Mpetsane Conservancy.

Gauteng (Fig. 2C): This is the smallest province, covering only 1.4% of South Africa (Fig. 2C) and falls within both the Savannah and highly threatened Grassland Biome with approximately 83% of the Province falling within the Highveld Grassland vegetation type, of which a mere 0.8% is currently conserved in South Africa. A total of 563 spp. (11 endemic) have been recorded from this Province (Table 2).

Major surveys in this Province include: Bakwena Cave in Irene (Durand et al. 2012); Ezemvelo Nature Reserve; Faerie Glen Nature Reserve in Pretoria; Gauteng caves (Dippenaar-Schoeman and Myburgh 2009); Groenkloof Nature Reserve; Irene survey; Kliprivierberg Nature Reserve; Rietondale Research Station (Van den Berg and Dippenaar-Schoeman 1991); Roodeplaat Research Station (Dippenaar-Schoeman 1976; Roodeplaatdam Nature Reserve (Dippenaar-Schoeman et al. 1989; Engelbrecht 2013); Pretoria National Botanical Gardens; Serene Valley; Suikerbosrand Nature Reserve; Tswaing Crater Nature Reserve; and Waterkloof Nature Reserve.

KwaZulu-Natal (Fig. 2D): KwaZulu-Natal covers 7.6% of South Africa and harbours many vegetation types. Grassland predominates in the western interior of the Province, with savannah towards the east. Alpine vegetation occurs on the high Drakensberg Escarpment on the border with Lesotho. The Coastal Belt vegetation is regularly interrupted by large river systems, where a thicket-type vegetation is often present in the valleys. A total of 1034 species have been recorded, which constitutes 45.3% of the spiders of South Africa (Table 2). Dr. Reginald Lawrence, one of the best-known South African arachnologists, was stationed at the Natal Museum from 1935 to 1986 and he extensively sampled and described species from the Province.

Major surveys in the Province include: Buffelsdraai Landfill Conservancy; Howick, Farm Wakefield; Drakensberg; False Bay Park (Lovell et al. 2007); Hluhluwe Game Reserve (Mgobozi et al. 2008); Farm Vergeval, Pongola district (Dippenaar-Schoeman et al. 1978); iSimangaliso Wetland Park, various localities; Ithala Game Reserve; Kosi Bay Nature Reserve; Maputaland (Lawrence et al. 1980); Maluti-Drakensberg Transfrontier Project (MDTP); Midlands, Good Hope and Maybole plantation estates (Yekwayo et al. 2016, 2017); Midlands, Ncandu Falls, Moorfield Mountain Farm and Normandien Farms (Booysen and Haddad 2021); Ngome State Forest (Van der Merwe et al. 1996; Dippenaar-Schoeman et al. 2006); Ndumo Game Reserve (Haddad et al. 2006; Haddad 2016, 2022); Ophathe Game Reserve (Haddad and Dippenaar-Schoeman 2015); Vernon Crookes Nature Reserve; Phinda Game Reserve (Lovell et al. 2010); Richards Bay rehabilitated coastal forest (Dippenaar-Schoeman and Wassenaar 2002, 2006); Sani Pass altitudinal gradient; Sodwana Bay National Park; Tembe Elephant Park (Haddad et al. 2010); and uMkhuze Game Reserve (Lovell et al. 2007).

Limpopo (Fig. 2E): This Province covers 10.6% of South Africa. The vegetation ranges from Afromontane forests, savannahs, shrubland, to semi-desert areas with small trees and bushes. The landscape also ranges from mountainous to flat plains. The first surveys in the Province were undertaken during a five-year harvester termite control project (Dippenaar-Schoeman et al. 1978). Another large dataset became available during surveys looking at the effect of chemical control of quelea finches on the Springbok Flats.

A total of 905 species (55 endemic) is known from the Province (Table 2).

Major surveys in the Province include: Atherstone Game Reserve; Blouberg Nature Reserve (Muelelwa et al. 2010; Foord et al. 2019); Farm Amsterdam, Dendron District (Dippenaar-Schoeman et al. 1978); Farm Zandrivier, Lephalale District, Bioblitz; Ka-Ndengeza Village (Joseph et al. 2017; Foord et al. 2018); Klein Kariba Resort; Lekgalameetse Nature Reserve (Foord et al. 2016); Lephalale; Limpopo Valley; Luvhondo Nature Reserve (Foord et al. 2008); Makelali Nature Reserve (Whitmore et al. 2001, 2002); Marakele National Park; Nylsvley

Nature Reserve (Heidger 1988; Dippenaar-Schoeman et al. 2009); Pietersburg Nature Reserve (Dippenaar et al. 2008); Rust de Winter cotton surveys (Dippenaar-Schoeman et al. 1999b); Soutpansberg long-term survey (Foord et al. 2002); Sovenga Hill (Modiba et al. 2005); Syferkuil research site; Venetia Limpopo Nature Reserve; Vhembe Biosphere Reserve; and Waterberg Biosphere.

Mpumalanga (Fig. 3A): Mpumalanga covers a total of 6.5% of South Africa's surface. The province contains several distinct physiographic regions: the Highveld, a plateau ranging in elevation from 1200 to 1800 metres in the west; the forested Drakensberg Mountains rising to more than 2300 metres in the east; and the Lowveld, a bush-clad plain that slopes gently upward towards the Lebombo Mountains on the Mozambique and Eswatini borders to the east. Much of Mpumalanga is drained by eastward-flowing tributaries of the Limpopo River.

A total of 645 species (20 endemic) have been recorded from the Province (Table 2).

Major surveys in the Province include: Barberton Nature Reserve (Mwabvu and Yekwayo 2019; Mavasa et al. 2023); Delmas (Bt maize); Kruger National Park (Dippenaar-Schoeman and Leroy 2003; Jonsson et al. 2010; Robertson et al. 2011), Lowveld Botanical Gardens, Mariepskop (Taylor et al. 2020); Marble Hall (Bt cotton) (Mellet et al. 2006); Mbombela (avocado, macadamia, citrus)

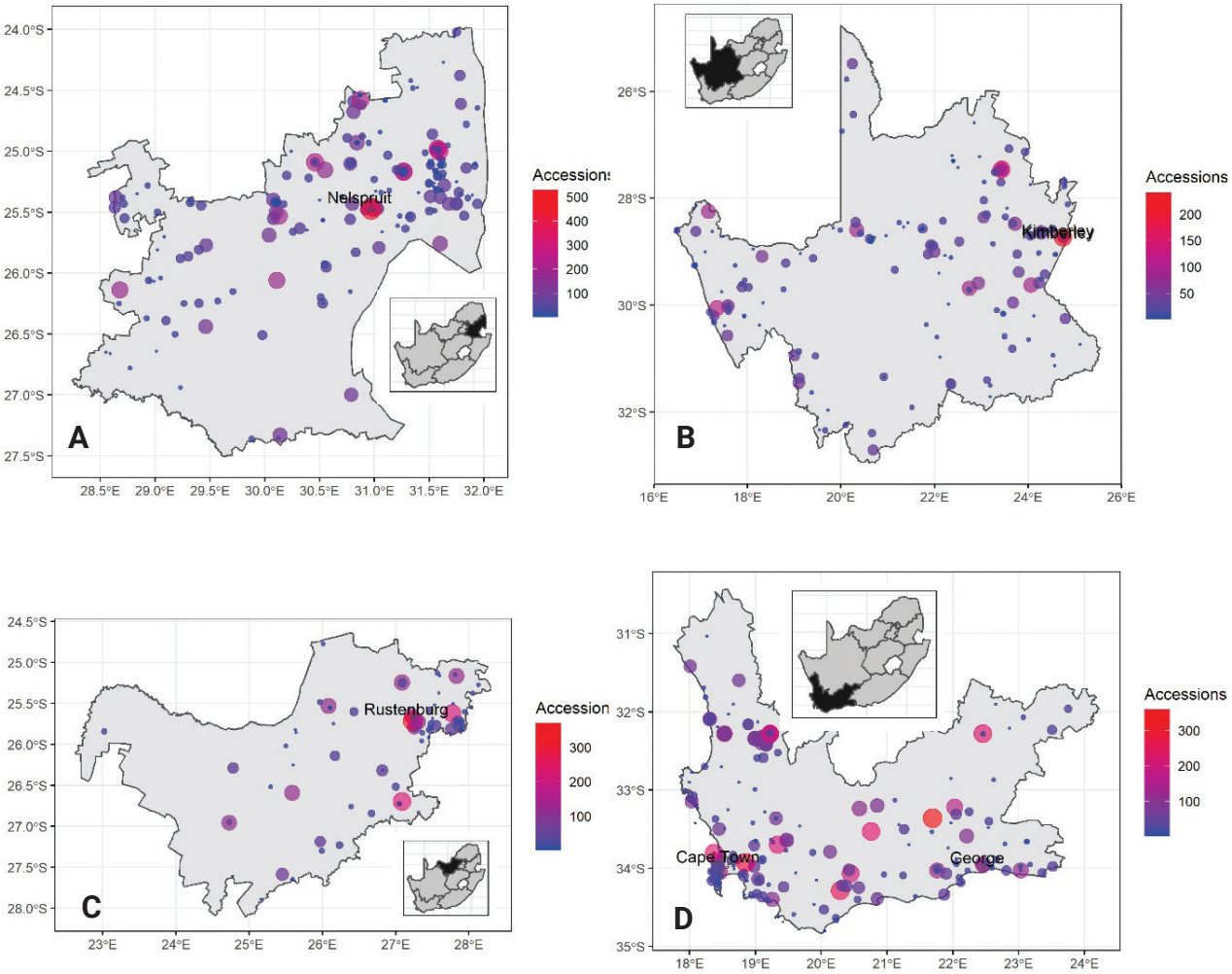

Figure 3. Spatial distribution and number of accessions for localities in: **A** Mpumalanga **B** Northern Cape **C** North West **D** Western Cape.

(Van den Berg et al. 1992; Dippenaar-Schoeman 1998; Dippenaar-Schoeman et al. 2001a, b; Dippenaar-Schoeman et al. 2005a); Steenkampsberg (Jansen et al. 2013); and Sterkspruit Nature Reserve (Mwabvu and Yekwayo 2019).

North West (Fig. 3C): This Province covers 9.5% of the surface of South Africa. Much of the Province consists of flat areas of scattered trees and grassland. The Magaliesberg mountain range in the north-east extends about 130 km from Pretoria to Rustenburg. The Vaal River flows along the southern border of the Province.

A total of 344 species (six endemic) are known from this Province (Table 2).

The only long-term survey (4 years) was undertaken in the Kgaswane Nature Reserve. Students at the North-West University and University of Johannesburg participated in agro-ecosystem surveys focusing on cotton near Brits and Bt maize near Potchefstroom (Botha et al. 2015). Other surveys were mainly funded by SANSA and the Spider Club: Barberspan Nature Reserve; Borakalala Nature Reserve; Pilanesberg National Park; and Zeerust.

Northern Cape (Fig. 3B): The Northern Cape is South Africa's largest Province (29.7%). The Karoo Basin dominates this Province and consists mainly of sedimentary rocks and some dolerite intrusions. The south and south-east of the Province are high-lying, 1200–1900 metres, in the Roggeveld and Nuweveld Districts. The west coast is dominated by the Namaqualand region, famous for its spring flowers and Succulent Karoo vegetation. This area is hilly to mountainous and consists of granites and metamorphic rocks. The central areas are generally flat with interspersed salt pans. Kimberlite intrusions punctuate the Karoo rocks, giving the Province its most precious natural resource, diamonds. The north is primarily Kalahari Desert, characterised by parallel red sand dunes and acacia tree dry savannah.

In spite of being the largest province, only 559 species (62 endemic) have so far been recorded in this Province (Table 2).

Very few sites have been intensively sampled: Akkerendam Nature Reserve; Augrabies Falls National Park; Brand-se-Baai; Diamond Route Reserves (Benfontein, Dronfield, Rooipoort and Tswalu Game Reserves; Koingnaas; Namaqua National Park; Namaqua strip mining; Prieska pistachio orchards (Haddad et al. 2004, 2005; Haddad and Dippenaar-Schoeman 2005, 2006b; Haddad et al. 2009); Nigramoep; Richterveld National Park; Tankwa Karoo National Park; and Witsand Nature Reserve.

Western Cape (Fig. 3D): This Province covers 9.7% of the surface of South Africa. Spiders were included in the Cape Nature Western Cape State of Biodiversity 2012 report (Veldtman 2012), a first for South Africa.

A total of 938 spp. are known from the Province representing 41.1% of South African species. Major surveys include: Aardvark Nature Reserve; Beaufort West; Borrelfontein; Bontebok National Park; Cape Peninsula (Sharratt et al. 2000); Cape Floristic Region (Pryke and Samways 2008, 2009, 2010, 2012; Theron et al. 2020a, 2020b); Cape Winelands (Gaigher and Samways 2010, 2014; Geldenhuys et al. 2021, 2022); Coast to Karoo transect: Spiders of the Cederberg (Foord and Dippenaar-Schoeman 2016); De Hoop Nature Reserve (Haddad and Dippenaar-Schoeman 2009); Fernkloof Nature Reserve; Jonkershoek (Swart et al. 2017); Karoo National Park (Dippenaar-Schoeman et al. 1999a); Kirstenbosch National Botanical Garden (Tucker 1920); Kogelberg Biosphere Reserve between Kogelberg and Protea survey (Coetzee et al. 1990; Visser et al. 1999; Yekwayo et al. 2018); Robben Island (Mukherjee et al. 2010; Roets and Pryke 2013; Cooper

et al. 2017); Southern Cape Forest Complex (Swart et al. 2018, 2020); Swartberg National Park (Dippenaar-Schoeman et al. 2005b); Table Mountain National Park; Tierberg Long-Term Ecological Research site (Dean 1988; Dean and Milton 1995; Henschel and Lubin 2018); and Tierhoek Organic Farm (Arvidsson et al. 2020).

Conservation

Many species recorded from South Africa (946 spp., 41.4%) are still only known from one sex. Only 1316 spp. (57.6%) are known from both sexes and 23 spp. were described from juveniles (Table 4), resulting in many species that are still Data Deficient.

Conservation status

The majority (1445 spp., 64%) of species have a wide distribution without known threats and are listed as Least Concern (Table 3). Sufficient data is still lacking for 691 spp. (31%), which could not be evaluated and are listed as Data Deficient; 504 of these species are data deficient for taxonomic reasons, 187 are lacking sufficient distribution data to make informed assessments and 288 (12.6%) are known only from the type locality. Only 129 spp. (5.6%) are of special conservation concern (Table 3).

Table 3. Conservation status and endemicity of the spider species sampled from South Africa.

CONSERVATION STATUS	NO SPP	%
Data Deficient (DD)	692	31
Not Evaluated (NE)	2	0.001
Least Concern (LC)	1445	64
Special concern	126	5.6
• Rare	43	2
• Critical rare (CR)	22	1
• Endangered (EN)	24	1
• Vulnerable (VU)	32	1.4
• Near threatened (NT)	5	0.2
Endemicity		
0 – Africa and wider (C)	86	3.8
1 – African endemics (AE)	460	20.4
2 – Southern African endemics (STHE)	394	17.4
3–5 South African endemics (SAE)	932	41.3
6– Known only from type locality	390	17.3

Endemicity

Almost the two-thirds of recorded species (1322 spp., 58.4%) are endemic to South Africa (Table 3; Suppl. material 2), while 3.8% have a distribution beyond the Afrotropical Region, including several cosmopolitan species, 460 spp. (20.4%) which are endemic to Africa (AE) and 394 spp. (16.9%) only known from southern Africa.

Table 4. Checklist of spiders (Araneae) of South Africa.

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY AGELENIDAE C.L. Koch, 1837														
F	*Agelena australis* Simon, 1896	1	LC	AE		1	1			1		1		1
F	*Agelena gaerdesi* Roewer, 1955	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Benoitia deserticola* (Simon, 1910)	2	LC	STHE		1				1		1		1
B	*Benoitia ocellata* (Pocock, 1900)	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Benoitia raymondeae* (Lessert, 1915)	1	LC	AE		1		1		1				1
B	*Mistaria zuluana* (Roewer, 1955)	2	LC	STHE		1				1		1	1	1
M	*Olorunia punctata* Lehtinen, 1967	1	LC	AE		1		1		1	1	1		1
B	*Tegenaria domestica* (Clerck, 1757)	0	LC	C			1	1		1				1
B	*Tegenaria pagana* C.L. Koch, 1840	0	LC	C						1				1
B	*Tegenaria parietina* (Fourcroy, 1785)	0	LC	C										1
FAMILY AMAUROBIIDAE Thorell, 1870														
F	*Chresiona convexa* Simon, 1903	5	DDT	SAE	WCE									1
F	*Chresiona invalida* (Simon, 1898)	3	LC	SAE				1	1		1			1
F	*Chresiona nigrosignata* Simon, 1903	5	DDT	SAE	WCE									1
F	*Chumma bicolor* Jocqué & Alderweireldt, 2017	6	DDT	SAE	WCE									1
B	*Chumma foliata* Jocqué & Alderweireldt, 2017	5	DD	SAE	ECE	1								
B	*Chumma gastroperforata* Jocqué, 2001	4	LC	SAE		1								1
B	*Chumma inquieta* Jocqué, 2001	4	EN	SAE		1								1
B	*Chumma interfluvialis* Jocqué & Alderweireldt, 2017	6	DD	SAE	FSE		1							
B	*Chumma striata* Jocqué & Alderweireldt, 2017	5	RARE	SAE	WCE									1
B	*Chumma subridens* Jocqué & Alderweireldt, 2017	4	DD	SAE		1								1
M	*Chumma tsitsikamma* Jocqué & Alderweireldt, 2017	6	DDT	SAE	ECE	1								
F	*Macrobunus caffer* (Simon, 1898)	5	DDT	SAE	WCE									1
F	*Obatala armata* Lehtinen, 1967	5	DDT	SAE	WCE									1
M	*Pseudauximus annulatus* Purcell, 1908	4	DDT	SAE						1		1		
B	*Pseudauximus pallidus* Purcell, 1903	4	LC	SAE								1		1
F	*Pseudauximus reticulatus* Simon, 1902	5	DDT	SAE	WCE									1
FAMILY ANAPIDAE Simon, 1895														
B	*Crozetulus rhodesiensis* Brignoli, 1981	2	LC	STHE					1	1				1
B	*Crozetulus scutatus* (Lawrence, 1964)	5	CR	SAE	WCE									1
M	*Dippenaaria luxurians* Wunderlich, 1995	6	DDT	SAE	ECE	1								
B	*Metanapis bimaculata* (Simon, 1895)	6	DD	SAE	WCE									1
FAMILY ANYPHAENIDAE Bertkau, 1878														
B	*Amaurobioides africana* Hewitt, 1917	2	LC	STHE		1						1		1
FAMILY ARANEIDAE Clerck, 1757														
B	*Acusilas africanus* Simon, 1895	1	LC	AE		1			1	1				
F	*Aethriscus olivaceus* Pocock, 1902	1	LC	AE		1		1	1	1	1			
F	*Afracantha camerunensis* (Thorell, 1899)	1	LC	AE		1			1	1	1			
F	*Arachnura scorpionoides* Vinson, 1863	1	LC	AE		1		1	1	1	1			1
F	*Araneus apricus* Karsch, 1884	1	LC	AE		1		1	1	1	1	1	1	1
F	*Araneus coccinella* Pocock, 1898	3	LC	SAE					1	1	1			1
F	*Araneus graemii* Pocock, 1900	6	DDT	SAE	ECE	1								
F	*Araneus haploscapella* (Strand, 1907)	3	DDT	SAE		1		1						
F	*Araneus nigroquadratus* Lawrence, 1937	2	LC	STHE		1		1	1	1	1			1
F	*Araneus strupifer* (Simon, 1886)	1	LC	AE		1			1	1				
F	*Araneus tatianae* Lessert, 1938	1	LC	AE				1						
M	*Araneus varus* (Kauri, 1950)	6	DDT	SAE	KZNE				1					
M	*Argiope anomalopalpis* Bjørn, 1997	1	LC	AE		1								

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Argiope aurocincta* Pocock, 1898	1	LC	AE		1	1		1	1	1			1
B	*Argiope australis* (Walckenaer, 1805)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Argiope flavipalpis* (Lucas, 1858)	1	LC	AE				1		1	1			
B	*Argiope levii* Bjørn, 1997	1	LC	AE					1	1	1			
B	*Argiope lobata* (Pallas, 1772)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Argiope trifasciata* (Forsskål, 1775)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Argiope tapinolobata* Bjørn, 1997	1	LC	AE				1						
B	*Bijoaraneus legonensis* (Grasshoff & Edmunds, 1979)	1	LC	AE		1			1	1	1			1
F	*Caerostris corticosa* Pocock, 1902	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Caerostris sexcuspidata* (Fabricius, 1793)	1	LC	AE		1		1	1	1	1	1	1	1
B	*Caerostris tinamaze* Gregorič, 2015	6	CR	SAE	LE					1				
B	*Caerostris vicina* (Blackwall, 1866)	1	LC	AE		1			1	1				
B	*Cladomelea akermani* Hewitt, 1923	5	EN	SAE	KZNE				1					
F	*Cladomelea debeeri* Roff & Dippenaar-Schoeman, 2004	5	EN	SAE	KZNE				1					
F	*Cladomelea longipes* (O.P.-Cambridge, 1877)	1	LC	AE						1				
B	*Clitaetra irenae* Kuntner, 2006	1	LC	AE					1					
B	*Cyclosa insulana* (Costa, 1834)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Cyclosa oculata* (Walckenaer, 1802)	0	LC	C		1		1	1	1	1			1
B	*Cyphalonotus larvatus* (Simon, 1881)	1	LC	AE		1	1	1	1	1	1	1	1	
B	*Cyrtarachne ixoides* (Simon, 1870)	0	LC	C		1	1		1					
B	*Cyrtophora citricola* (Forsskål, 1775)	0	LC	C		1	1	1	1	1	1	1	1	1
F	*Cyrtophora petersi* Karsch, 1878	2	LC	STHE					1		1			
B	*Eriovixia excelsa* (Simon, 1889)	0	LC	C				1	1		1		1	
F	*Gasteracantha falcicornis* Butler, 1873	1	LC	AE					1		1			
B	*Gasteracantha milvoides* Butler, 1873	1	LC	AE		1			1	1	1			
B	*Gasteracantha sanguinolenta* C.L. Koch, 1844	1	LC	AE		1	1	1	1	1	1		1	1
B	*Gasteracantha versicolor* (Walckenaer, 1842)	1	LC	AE		1		1	1	1	1			1
F	*Gastroxya benoiti* Emerit, 1973	4	LC	SAE					1					
B	*Gea infuscata* Tullgren, 1910	1	LC	AE		1			1	1	1			1
B	*Hypsacantha crucimaculata* (Dahl, 1914)	1	LC	AE		1			1	1	1	1		1
B	*Hypsosinga holzapfelae* (Lessert, 1936)	2	LC	STHE				1	1	1	1		1	
B	*Hypsosinga lithyphantoides* Caporiacco, 1947	1	LC	AE		1	1	1	1	1	1			
B	*Hypsosinga pygmaea* (Sundevall, 1831)	0	LC	C				1	1	1	1			
F	*Ideocaira transversa* Simon, 1903	3	LC	SAE		1			1	1				1
B	*Ideocaira triquetra* Simon, 1903	4	LC	SAE		1								
B	*Isoxya cicatricosa* (C.L. Koch, 1844)	1	LC	AE		1			1	1	1	1		1
B	*Isoxya mossamedensis* Benoit, 1962	2	LC	STHE				1		1				1
B	*Isoxya mucronata* Walckenaer, 1841	1	LC	AE		1			1	1				
F	*Isoxya stuhlmanni* Bosenberg & Lenz, 1895	1	LC	AE		1	1		1	1	1			
B	*Isoxya tabulata* (Thorell, 1859)	1	LC	AE		1			1	1	1		1	1
F	*Isoxya yatesi* Emerit, 1973	3	DDT	SAE					1	1				
B	*Kilima decens* (Blackwall, 1866)	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Larinia bifida* Tullgren, 1910	1	LC	AE		1	1	1					1	
B	*Larinia chloris* (Audouin, 1826)	0	LC	C		1				1	1	1		1
B	*Larinia natalensis* (Grasshoff, 1971)	3	LC	SAE		1		1	1	1		1	1	1
B	*Lipocrea longissima* (Simon, 1881)	1	LC	AE			1	1	1	1	1			1
B	*Mahembea hewitti* (Lessert, 1930)	1	LC	AE				1	1	1			1	
B	*Megaraneus gabonensis* (Lucas, 1858)	1	LC	AE					1					
M	*Nemoscolus cotti* Lessert, 1933	2	LC	STHE		1	1	1	1	1	1	1		
F	*Nemoscolus elongatus* Lawrence, 1947	3	LC	SAE		1	1		1	1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Nemoscolus obscurus* Simon, 1897	3	DDT	SAE				1					1	
F	*Nemoscolus tubicola* (Simon, 1887)	2	LC	STHE		1	1	1		1	1	1	1	1
F	*Nemoscolus vigintipunctatus* Simon, 1897	2	LC	STHE		1	1	1	1		1		1	1
F	*Nemospiza conspicillata* Simon, 1903	5	DDT	SAE	LE					1				
B	*Neoscona alberti* (Strand, 1913)	1	LC	AE		1			1					
B	*Neoscona angulatula* (Schenkel, 1937)	1	LC	AE					1					
B	*Neoscona blondeli* (Simon, 1886)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Neoscona chiarinii* (Pavesi, 1883)	1	LC	AE					1		1			
B	*Neoscona hirta* (C. L. Koch, 1844)	1	LC	AE		1		1	1	1	1			1
B	*Neoscona kivuensis* Grasshoff, 1986	1	LC	AE								1		1
B	*Neoscona moreli* (Vinson, 1863)	0	LC	C		1	1	1	1	1	1	1	1	1
F	*Neoscona novella* (Simon, 1907)	1	LC	AE		1				1	1	1		1
B	*Neoscona penicillipes* (Karsch, 1879)	1	LC	AE		1	1	1	1	1	1	1	1	
B	*Neoscona quadrigibbosa* Grasshoff, 1986	1	LC	AE						1				
B	*Neoscona quincasea* Roberts, 1983	1	LC	AE		1	1	1	1	1	1			1
B	*Neoscona rapta* (Thorell, 1899)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Neoscona rufipalpis* (Lucas, 1858)	1	LC	AE		1		1	1	1	1		1	1
B	*Neoscona subfusca* (C.L. Koch, 1837)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Neoscona theisi theisiella* (Tullgren, 1910)	1	LC	AE		1		1	1	1	1		1	1
B	*Neoscona triangula* (Keyserling, 1864)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Neoscona vigilans* (Blackwall, 1865)	0	LC	C		1		1	1		1			1
B	*Nephilingis cruentata* (Fabricius, 1775)	0	LC	C					1	1	1			
M	*Paralarinia bartelsi* (Lessert, 1933)	3	LC	SAE		1			1					1
F	*Paraplectana thorntoni* (Blackwall, 1865)	1	LC	AE		1		1	1	1	1			1
F	*Paraplectana walleri* (Blackwall, 1865)	2	LC	STHE		1			1	1	1			
B	*Pararaneus cyrtoscapus* (Pocock, 1898)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Pararaneus perforatus* (Thorell, 1899)	1	LC	AE		1								
B	*Pararaneus spectator* (Karsch, 1885)	0	LC	C		1	1	1	1	1	1		1	
F	*Pasilobus dippenaarae* Roff & Haddad, 2015	5	DDT	SAE	KZNE				1					
F	*Poltys furcifer* Simon, 1881	1	LC	AE		1		1	1	1	1		1	
B	*Prasonica albolimbata* Simon, 1895	1	LC	AE				1	1	1	1			
B	*Prasonica nigrotaeniata* (Simon, 1909)	1	LC	AE				1	1	1	1			
B	*Prasonica seriata* Simon, 1895	1	LC	AE		1		1	1	1	1		1	1
B	*Pycnacantha tribulus* (Fabricius, 1781)	2	LC	STHE		1		1	1	1	1	1	1	1
F	*Singa albodorsata* Kauri, 1950	3	LC	SAE		1			1	1	1			
F	*Singa lawrencei* (Lessert, 1930)	1	LC	AE				1	1	1			1	
F	*Singafrotypa mandela* Kuntner & Hormiga, 2002	3	LC	SAE					1	1				1
B	*Trichonephila fenestrata* (Thorell, 1859)	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Trichonephila inaurata madagascariensis* (Vinson, 1863)	1	LC	AE		1			1		1			
B	*Trichonephila komaci* Kuntner & Coddington, 2009	1	LC	AE					1					
B	*Trichonephila senegalensis annulata* (Thorell, 1859)	2	LC	STHE		1	1	1	1	1	1	1	1	1
F	*Ursa turbinata* Simon, 1895	3	LC	SAE				1		1	1			1
FAMILY ARCHAEIDAE C. L. Koch & Berendt, 1854														
B	*Afrarchaea ansiae* Lotz, 2015	6	DD	SAE	KZNE				1					
B	*Afrarchaea bergae* Lotz, 1996	4	LC	SAE						1	1			
B	*Afrarchaea cornutus* Lotz, 2003	6	VU	SAE	KZNE				1					
F	*Afrarchaea entabeniensis* Lotz, 2003	6	CR	SAE	LE					1				
M	*Afrarchaea fernkloofensis* Lotz, 1996	6	CR	SAE	ECE	1								
B	*Afrarchaea godfreyi* Hewitt, 1919	4	LC	SAE		1			1					
F	*Afrarchaea haddadi* Lotz, 2006	6	DDT	SAE	ECE		1							

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Afrarchaea harveyi* Lotz, 2003	6	DDT	SAE	KZNE				1					
F	*Afrarchaea kranskopensis* Lotz, 1996	6	DDT	SAE	KZNE				1					
M	*Afrarchaea lawrencei* Lotz, 1996	6	DDT	SAE	KZNE				1					
B	*Afrarchaea neethlingi* Lotz, 2017	6	Rare	SAE	FSE			1						
B	*Afrarchaea ngomensis* Lotz, 1996	4	VU	SAE					1		1			
F	*Afrarchaea royalensis* Lotz, 2006	6	DDT	SAE	KZNE				1					
B	*Afrarchaea woodae* Lotz, 2006	5	EN	SAE	ECE	1								
FAMILY ATYPIDAE Thorell, 1870														
M	*Calommata meridionalis* Fourie, Haddad & Jocqué, 2011	5	NT	SAE	FSE		1							
B	*Calommata transvaalica* Hewitt, 1916	4	VU	SAE				1		1				
FAMILY BARYCHELIDAE Simon, 1889														
F	*Pisenor arcturus* (Tucker, 1917)	2	LC	STHE						1	1		1	
B	*Pisenor notius* Simon, 1889	1	LC	AE						1				
F	*Sipalolasma humicola* (Benoit, 1965)	1	LC	AE						1		1	1	
FAMILY BEMMERIDAE Simon, 1903														
F	*Homostola abernethyi* (Purcell, 1903)	5	DDT	SAE	ECE	1								
F	*Homostola pardalina* (Hewitt, 1913)	3	LC	SAE			1			1	1			
F	*Homostola reticulata* (Purcell, 1902)	5	DDT	SAE	WCE									1
F	*Homostola vulpecula* Simon, 1892	3	LC	SAE				1	1	1	1			
B	*Homostola zebrina* Purcell, 1902	2	LC	STHE				1	1	1	1			
M	*Spiroctenus armatus* Hewitt, 1913	6	DDT	SAE	ECE	1								
F	*Spiroctenus broomi* Tucker, 1917	6	DDT	SAE	WCE									1
B	*Spiroctenus cambierae* (Purcell, 1902)	6	DD	SAE	WCE									1
M	*Spiroctenus coeruleus* Lawrence, 1952	6	DDT	SAE	KZNE				1					
B	*Spiroctenus collinus* (Pocock, 1900)	5	DD	SAE	WCE									1
B	*Spiroctenus curvipes* Hewitt, 1919	6	DD	SAE	KZNE				1					
M	*Spiroctenus exilis* Lawrence, 1938	6	DDT	SAE	KZNE				1					
F	*Spiroctenus flavopunctatus* (Purcell, 1903)	6	DDT	SAE	ECE		1							
F	*Spiroctenus fossorius* (Pocock, 1900)	6	DDT	SAE	ECE		1							
F	*Spiroctenus fuliginosus* (Pocock, 1902)	6	DDT	SAE	ECE		1							
M	*Spiroctenus gooldi* (Purcell, 1903)	5	DDT	SAE	WCE									1
M	*Spiroctenus inermis* (Purcell, 1903)	4	DDT	SAE								1		1
M	*Spiroctenus latus* Purcell, 1904	6	DDT	SAE	WCE									1
B	*Spiroctenus lightfooti* (Purcell, 1902)	4	DD	SAE								1		1
F	*Spiroctenus lignicola* Lawrence, 1937	6	DDT	SAE	KZNE				1					
B	*Spiroctenus londinensis* Hewitt, 1919	6	DD	SAE	ECE	1								
B	*Spiroctenus marleyi* Hewitt, 1919	6	DD	SAE	KZNE				1					
B	*Spiroctenus minor* (Hewitt, 1913)	6	DD	SAE	ECE	1								
M	*Spiroctenus pallidipes* Purcell, 1904	6	DDT	SAE	WCE									1
J	*Spiroctenus pardalina* (Simon, 1903)	6	DDT	SAE	?									
J	*Spiroctenus pectiniger* (Simon, 1903)	6	DDT	SAE	WCE									1
F	*Spiroctenus pilosus* Tucker, 1917	5	DDT	SAE	FSE		1							
F	*Spiroctenus punctatus* Hewitt, 1916	5	DDT	SAE	KZNE				1					
M	*Spiroctenus purcelli* Tucker, 1917	6	DDT	SAE	WCE									1
B	*Spiroctenus sagittarius* (Purcell, 1902)	6	DD	SAE	WCE									1
F	*Spiroctenus schreineri* (Purcell, 1903)	4	DDT	SAE								1		1
M	*Spiroctenus tricalcaratus* (Purcell, 1903)	5	DDT	SAE	WCE									1
B	*Spiroctenus validus* (Purcell, 1902)	5	DD	SAE	WCE									1
FAMILY CAPONIIDAE Simon, 1890														
M	*Caponia braunsi* Purcell, 1904	4	DDT	SAE		1								1
B	*Caponia capensis* Purcell, 1904	2	LC	STHE		1						1		1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
M	*Caponia chelifera* Lessert, 1936	2	LC	STHE				1	1	1	1			
M	*Caponia forficifera* Purcell, 1904	5	DDT	SAE	WCE									1
B	*Caponia hastifera* Purcell, 1904	4	LC	SAE		1	1							
B	*Caponia karrooica* Purcell, 1904	4	DDT	SAE		1								1
B	*Caponia natalensis* (O.P.-Cambridge, 1874)	1	LC	AE					1	1	1			
F	*Caponia secunda* Pocock, 1900	6	DDT	SAE	ECE	1								
B	*Caponia simoni* Purcell, 1904	6	DD	SAE	WCE									1
B	*Caponia spiralifera* Purcell, 1904	3	LC	SAE		1		1			1	1	1	
B	*Diplogena arida* Haddad, 2015	2	LC	STHE								1		
B	*Diploglena capensis* Purcell, 1904	5	DD	SAE	WCE									1
B	*Diploglena dippenaarae* Haddad, 2015	5	EN	SAE	WCE									1
B	*Diploglena karooica* Haddad, 2015	2	LC	STHE								1		1
B	*Diploglena major* Lawrence, 1928	2	LC	STHE						1				
B	*Diploglena proxila* Haddad, 2015	6	CR	SAE	WCE									1
FAMILY CHEIRACANTHIIDAE Wagner, 1887														
B	*Cheiracanthium aculeatum* Simon, 1884	1	LC	AE		1		1	1	1	1			
B	*Cheiracanthium africanum* Lessert, 1921	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Cheiracanthium angolensis* Lotz, 2007	2	LC	STHE						1	1			
F	*Cheiracanthium dippenaarae* Lotz, 2007	3	DD	SAE				1	1	1				
F	*Cheiracanthium foordi* Lotz, 2015	6	DDT	SAE	LE					1				
B	*Cheiracanthium furculatum* Karsch, 1879	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Cheiracanthium minshullae* Lotz, 2007	2	LC	STHE			1		1					
B	*Cheiracanthium schenkeli* Caporiacco, 1949	1	LC	AE					1	1				
F	*Cheiracanthium shiluvanensis* Lotz, 2007	4	DDT	SAE					1	1				
B	*Cheiracanthium vansoni* Lawrence, 1936	1	LC	AE		1	1	1	1	1	1	1	1	
M	*Cheiramiona akermani* (Lawrence, 1942)	5	DDT	SAE	KZNE				1					
B	*Cheiramiona amarifontis* Lotz, 2002	4	LC	SAE		1			1					
B	*Cheiramiona ansiae* Lotz, 2002	4	LC	SAE		1			1					1
F	*Cheiramiona baviaan* Lotz, 2015	6	DDT	SAE	ECE	1								
F	*Cheiramiona boschrandensis* Lotz, 2015	6	DDT	SAE	FSE		1							
B	*Cheiramiona clavigera* (Simon, 1897)	3	LC	SAE		1			1	1	1			1
B	*Cheiramiona collinita* (Lawrence, 1938)	4	LC	SAE		1			1					
M	*Cheiramiona debeeri* Lotz, 2015	6	DDT	SAE	ME						1			
B	*Cheiramiona ferrumfontis* Lotz, 2002	4	LC	SAE								1		1
B	*Cheiramiona filipes* (Simon, 1898)	2	LC	STHE		1			1	1				
B	*Cheiramiona florisbadensis* Lotz, 2002	3	LC	SAE		1		1	1		1			1
M	*Cheiramiona fontanus* Lotz, 2002	6	DDT	SAE	FSE		1							
M	*Cheiramiona haddadi* Lotz, 2015	6	DDT	SAE	KZNE				1					
M	*Cheiramiona hlathikulu* Lotz, 2015	5	DDT	SAE	KZNE				1					
B	*Cheiramiona hogsbackensis* Lotz, 2015	5	CR	SAE	ECE	1								
M	*Cheiramiona jakobsbaaiensis* Lotz, 2015	6	DDT	SAE	WCE									1
B	*Cheiramiona kentaniensis* Lotz, 2003	5	LC	SAE	ECE	1								
B	*Cheiramiona kirkspriggsi* Lotz, 2015	5	LC	SAE	KZNE				1					
B	*Cheiramiona krugerensis* Lotz, 2003	3	LC	SAE		1			1	1	1			1
B	*Cheiramiona lajuma* Lotz, 2003	3	LC	SAE		1			1	1	1			1
M	*Cheiramiona langi* Lotz, 2003	2	DDT	STHE						1				
M	*Cheiramiona lindae* Lotz, 2015	6	DDT	SAE	ECE	1								
B	*Cheiramiona mkhambathi* Lotz, 2015	6	DDT	SAE	ECE	1								
B	*Cheiramiona mlawula* Lotz, 2003	2	LC	STHE						1	1			
B	*Cheiramiona paradisus* Lotz, 2003	2	LC	STHE		1	1		1	1	1			1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Cheiramiona plaatbosensis* Lotz, 2015	4	LC	SAE		1								1
M	*Cheiramiona qachasneki* Lotz, 2015	5	DDT	SAE	ECE	1								
B	*Cheiramiona regis* Lotz, 2003	4	LC	SAE			1		1					
M	*Cheiramiona robinae* Lotz, 2015	5	DDT	SAE	ECE	1								
B	*Cheiramiona saniensis* Lotz, 2015	5	LC	SAE	KZNE				1					
B	*Cheiramiona silvicola* (Lawrence, 1938)	3	LC	SAE		1			1					1
B	*Cheiramiona simplicitarsis* (Simon, 1910)	3	LC	SAE					1		1	1		1
M	*Cheiramiona stellenboschiensis* Lotz, 2003	5	DD	SAE	WCE									1
M	*Cheiramiona tembensis* Lotz, 2015	6	DDT	SAE	KZNE				1					
F	*Cheiramiona upperbyensis* Lotz, 2015	5	DDT	SAE								1		
M	*Lessertina capensis* Haddad, 2014	5	DDT	SAE	WCE									1
B	*Lessertina mutica* Lawrence, 1942	3	LC	SAE		1			1	1	1			
FAMILY CITHAERONIDAE Simon, 1893														
M	*Cithaeron contentum* Jocqué & Russel-Smith, 2011	4	DD	SAE						1	1			
FAMILY CLUBIONIDAE Wagner, 1887														
B	*Clubiona abbajensis* Strand, 1906	1	LC	AE		1			1	1	1			1
B	*Clubiona africana* Lessert, 1921	1	LC	AE		1	1	1	1	1	1		1	1
B	*Clubiona annuligera* Lessert, 1929	1	LC	AE					1	1				
M	*Clubiona aspidiphora* Simon, 1910	2	DDT	STHE								1		
M	*Clubiona bevisi* Lessert, 1923	3	LC	SAE			1		1	1			1	
M	*Clubiona biaculeata* Simon, 1897	6	DDT	SAE	ECE	1								
F	*Clubiona capensis* Simon, 1897	6	DDT	SAE	ECE	1								
M	*Clubiona citricolor* Lawrence, 1952	4	DDT	SAE			1		1					
F	*Clubiona durbana* Roewer, 1951	3	LC	SAE		1	1		1	1	1			
M	*Clubiona godfreyi* Lessert, 1921	1	LC	AE		1				1				
F	*Clubiona helva* Simon, 1897	6	DDT	SAE	KZNE				1					
M	*Clubiona kiboschensis* Lessert, 1921	1	LC	AE		1								
F	*Clubiona lawrencei* Roewer, 1951	2	LC	STHE		1			1	1				1
B	*Clubiona limpida* Simon, 1897	6	DDT	SAE	KZNE				1					
M	*Clubiona natalica* Simon, 1897	6	DDT	SAE	KZNE				1					
F	*Clubiona nollothensis* Simon, 1897	5	DDT	SAE	NCE							1		
B	*Clubiona pongolensis* Lawrence, 1952	3	LC	SAE		1		1	1	1				
F	*Clubiona pupillaris* Lawrence, 1938	3	LC	SAE		1		1	1	1	1			
B	*Clubiona revillioidi* Lessert, 1936	2	LC	STHE		1	1	1		1	1			
M	*Clubiona rumpiana* Lawrence, 1952	5	DDT	SAE	KZNE				1					
M	*Clubiona sigillata* Lawrence, 1952	3	LC	SAE		1		1	1					
J	*Clubiona sparassella* Strand, 1909	6	DDT	SAE	?									
B	*Clubiona subtrivialis* Strand, 1906	1	LC	AE					1	1				
M	*Clubiona umbilensis* Lessert, 1923	3	LC	SAE		1								1
F	*Clubiona vachoni* Lawrence, 1952	5	DDT	SAE	KZNE				1					
B	*Clubiona valens* Simon, 1897	5	DD	SAE	KZNE				1					
FAMILY CORINNIDAE Karsch, 1880														
B	*Apochinomma decepta* Haddad, 2013	2	LC	STHE						1	1			
M	*Apochinomma elongata* Haddad, 2013	1	LC	AE				1	1					
B	*Apochinomma formicaeforme* Pavesi, 1881	1	LC	AE					1	1	1		1	
B	*Austrophaea zebra* Lawrence, 1952	4	LC	SAE		1								1
B	*Cambalida dippenaarae* Haddad, 2012	1	LC	AE		1	1	1	1	1	1	1	1	
B	*Cambalida fulvipes* Simon, 1896	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Coenoptychus mutillica* (Haddad, 2004)	1	LC	AE			1		1	1	1	1		
B	*Coenoptychus tropicalis* (Haddad, 2004)	1	LC	AE		1	1		1	1				1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Copa flavoplumosa* Simon, 1885	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Copa kei* Haddad, 2013	4	LC	SAE		1			1					
B	*Copuetta erecta* Haddad, 2013	2	LC	STHE			1		1					
B	*Copuetta lacustris* (Strand, 1916)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Copuetta lotzi* Haddad, 2013	3	LC	SAE			1	1			1	1		1
B	*Copuetta magna* Haddad, 2013	1	LC	AE					1		1			
B	*Copuetta maputa* Haddad, 2013	2	LC	STHE					1					
B	*Corinnomma lawrencei* Haddad, 2006	1	LC	AE					1	1				
B	*Corinnomma semiglabrum* (Simon, 1896)	1	LC	AE				1	1	1	1		1	
B	*Echinax natalensis* Haddad, 2012	4	LC	SAE		1			1					
B	*Echinax similis* Haddad, 2012	6	DD	SAE	KZNE				1					
B	*Graptartia granulosa* Simon, 1896	1	LC	AE						1				
F	*Hortipes aelurisiepae* Bosselaers & Jocqué, 2000	2	LC	STHE					1					
F	*Hortipes atalante* Bosselaers & Jocqué, 2000	5	VU	SAE	KZNE				1					
B	*Hortipes coccinatus* Bosselaers & Jocqué, 2000	5	VU	SAE	LE					1				
B	*Hortipes contubernalis* Bosselaers & Jocqué, 2000	5	Rare	SAE	LE					1				
M	*Hortipes griswoldi* Bosselaers & Jocqué, 2000	4	DDT	SAE					1		1			
M	*Hortipes hyakutake* Bosselaers & Jocqué, 2000	6	DDT	SAE	ECE		1							
F	*Hortipes irimus* Bosselaers & Jocqué, 2000	6	DDT	SAE	KZNE				1					
F	*Hortipes licnophorus* Bosselaers & Jocqué, 2000	6	DDT	SAE	ME						1			
B	*Hortipes luytenae* Bosselaers & Ledoux, 1998	6	DD	SAE	KZNE				1					
B	*Hortipes merwei* Bosselaers & Jocqué, 2000	5	VU	SAE	KZNE				1					
F	*Hortipes mesembrinus* Bosselaers & Jocqué, 2000	5	DDT	SAE	ECE		1							
M	*Hortipes rothorum* Bosselaers & Jocqué, 2000	6	DDT	SAE	KZNE				1					
B	*Hortipes schoemanae* Bosselaers & Jocqué, 2000	2	LC	STHE				1	1		1			
B	*Hortipes wimmertensi* Bosselaers & Jocqué, 2000	5	CR	SAE	KZNE				1					
B	*Medmassa semiaurantiaca* Simon, 1910	1	LC	AE					1					
B	*Merenius alberti* Lessert, 1923	2	LC	STHE		1		1	1	1	1		1	1
B	*Merenius simoni* Lessert, 1921	1	LC	AE						1				1
B	*Messapus martini* Simon, 1898	1	LC	AE				1	1		1			
F	*Messapus meridionalis* Haddad & Mbo, 2015	6	DDT	SAE	KZNE				1					
B	*Messapus natalis* (Pocock, 1898)	2	LC	STHE					1		1			
B	*Pronophaea natalica* Simon, 1897	3	LC	SAE		1			1	1	1			1
B	*Pronophaea proxima* (Lessert, 1923)	3	LC	SAE		1			1	1				1
F	*Pronophaea vidua* (Lessert, 1923)	5	DDT	SAE	KZNE				1					
B	*Vendaphaea lajuma* Haddad, 2009	5	DD	SAE	LE					1				
FAMILY CTENIDAE Keyserling, 1877														
M	*Africactenus tridentatus* Hyatt, 1954	2	LC	STHE		1					1			
B	*Ctenus caligineus* des Arts, 1912	1	LC	AE						1				
M	*Ctenus corniger* F.O.P.-Cambridge, 1898	6	DDT	SAE	KZNE				1					
F	*Ctenus gulosus* Des Arts, 1912	2	LC	STHE		1			1	1	1			
F	*Ctenus parvoculatus* Benoit, 1979	3	LC	SAE		1			1	1	1			1
B	*Ctenus pulchriventris* (Simon, 1896)	2	LC	STHE		1			1	1	1			
B	*Ctenus spectabilis* Lessert, 1921	1	LC	AE								1		
M	*Ctenus transvaalensis* Benoit, 1981	3	LC	SAE				1	1	1	1	1		
FAMYLY CYATHOLIPIDAE Simon, 1894														
B	*Cyatholipus avus* Griswold, 1987	4	LC	SAE		1								1
F	*Cyatholipus hirsutissimus* Simon, 1894	4	LC	SAE			1							1
F	*Cyatholipus icubatus* Griswold, 1987	5	DD	SAE	KZNE				1					
B	*Cyatholipus isolatus* Griswold, 1987	4	NT	SAE						1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Cyatholipus quadrimaculatus* Simon, 1894	4	LC	SAE		1								1
M	*Cyatholipus tortilis* Griswold, 1987	6	Rare	SAE	KZNE				1					
B	*Ilisoa conjugalis* Griswold, 1987	5	Rare	SAE	WCE									1
F	*Ilisoa hawequas* Griswold, 1987	6	DD	SAE	WCE									1
M	*Ilisoa knysna* Griswold, 1987	5	VU	SAE	WCE									1
B	*Isicabu reavelli* Griswold, 1987	6	DD	SAE	KZNE				1					
M	*Isicabu zuluensis* Griswold, 1987	5	VU	SAE	KZNE				1					
B	*Pokennips dentipes* Simon, 1894	5	DD	SAE	WCE									1
B	*Ubacisi capensis* (Griswold, 1987)	4	LC	SAE		1								1
B	*Ulwembua denticulata* Griswold, 1987	3	LC	SAE		1			1	1	1			1
B	*Ulwembua outeniqua* Griswold, 1987	4	LC	SAE		1								1
B	*Ulwembua pulchra* Griswold, 1987	4	LC	SAE		1			1					
FAMILY CYRTAUCHENIIDAE Simon, 1889														
B	*Ancylotrypa barbertoni* (Hewitt, 1913)	4	DD	SAE						1	1			
B	*Ancylotrypa brevicornis* (Hewitt, 1919)	3	LC	SAE				1		1			1	
B	*Ancylotrypa brevipalpis* (Hewitt, 1916)	3	LC	SAE				1	1	1	1	1	1	
B	*Ancylotrypa breyeri* (Hewitt, 1919)	6	DD	SAE	KZNE				1					
B	*Ancylotrypa bulcocki* (Hewitt, 1916)	6	DD	SAE	ECE	1								
F	*Ancylotrypa coloniae* (Pocock, 1902)	6	DDT	SAE	ECE	1								
B	*Ancylotrypa cornuta* Purcell, 1904	5	DD	SAE	ECE	1								
F	*Ancylotrypa dentata* (Purcell, 1903)	6	DDT	SAE	NCE							1		
B	*Ancylotrypa dreyeri* (Hewitt, 1915)	5	DDT	SAE	FSE		1							
B	*Ancylotrypa elongata* Purcell, 1908	2	LC	STHE						1	1	1		
F	*Ancylotrypa flavidofusula* (Hewitt, 1915)	6	DDT	SAE	ECE	1								
F	*Ancylotrypa lateralis* (Purcell, 1902)	5	DDT	SAE	ECE	1								
F	*Ancylotrypa magnisigillata* (Hewitt, 1914)	5	DDT	SAE	KZNE				1					
M	*Ancylotrypa namaquensis* (Purcell, 1908)	5	LC	SAE	NCE							1		
M	*Ancylotrypa nigriceps* (Purcell, 1902)	3	LC	SAE			1	1	1					
B	*Ancylotrypa nuda* (Hewitt, 1916)	3	LC	SAE				1	1	1	1		1	
M	*Ancylotrypa nudipes* (Hewitt, 1923)	6	DDT	SAE	ECE	1								
F	*Ancylotrypa oneili* (Purcell, 1902)	6	DDT	SAE	ECE	1								
M	*Ancylotrypa pallidipes* (Purcell, 1904)	5	DDT	SAE	WCE									1
M	*Ancylotrypa parva* (Hewitt, 1916)	6	DDT	SAE	ECE	1								
B	*Ancylotrypa pretoriae* (Hewitt, 1913)	3	LC	SAE			1	1		1	1	1		
B	*Ancylotrypa pusilla* (Purcell, 1903)	5	LC	SAE	NCE							1		
B	*Ancylotrypa rufescens* (Hewitt, 1916)	3	LC	SAE				1		1	1		1	
B	*Ancylotrypa sororum* (Hewitt, 1916)	3	LC	SAE		1	1					1		1
B	*Ancylotrypa spinosa* Simon, 1889	5	DD	SAE	ECE	1								
M	*Ancylotrypa tookei* (Hewitt, 1919)	5	DDT	SAE	ECE	1								
F	*Ancylotrypa vryheidensis* (Hewitt, 1915)	5	LC	SAE	KZNE				1					
F	*Ancylotrypa zebra* (Simon, 1892)	3	LC	SAE		1			1	1	1			
M	*Ancylotrypa zuluensis* (Lawrence, 1937)	5	DDT	SAE	KZNE				1					
FAMILY DEINOPIDAE C.L. Koch, 1850														
B	*Deinopis anchietae* Brito Capello, 1867	1	LC	AE					1	1	1			
F	*Deinopis aspectans* Pocock, 1900	1	LC	AE					1	1	1			
B	*Deinopis cornigera* Gerstäcker, 1873	1	LC	AE					1		1		1	
B	*Deinopis cylindrica* Pocock, 1898	2	LC	STHE				1	1	1				1
B	*Menneus camelus* Pocock, 1902	3	LC	SAE		1		1	1	1	1		1	1
B	*Menneus capensis* (Purcell, 1904)	5	LC	SAE	WCE									1
B	*Menneus dromedarius* Purcell, 1904	1	LC	AE		1			1					1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY DESIDAE Pocock, 1895														
B	*Badumna longinqua* (L. Koch, 1867)	0	LC	C		1	1							
B	*Desis formidabilis* (O.P.Cambridge, 1890)	2	LC	STHE		1						1		1
FAMILY DICTYNIDAE O. Pickard-Cambridge, 1871														
B	*Archaeodictyna condocta* (O.P.-Cambridge, 1876)	0	LC	C				1	1	1		1		1
B	*Archaeodictyna ulova* Griswold & Meikle-Griswold, 1987	3	LC	SAE		1			1	1				
B	*Brigittea civica* (Lucas, 1850)	0	LC	C				1						
F	*Mashimo leleupi* Lehtinen, 1967	1	LC	AE		1			1	1	1			1
F	*Shango capicola* (Strand, 1909)	6	DDT	SAE	WCE									1
FAMILY DRYMUSIDAE Simon, 1893														
B	*Izithunzi capense* (Simon, 1893)	5	Rare	SAE	WCE									1
B	*Izithunzi lina* Labarque, Pérez-González & Griswold, 2017	5	Rare	SAE	WCE									1
B	*Izithunzi productum* (Purcell, 1904)	5	Rare	SAE	WCE									1
B	*Izithunzi silvicola* (Purcell, 1904)	4	Rare	SAE		1								1
F	*Izithunzi zondii* Labarque, Pérez-González & Griswold, 2017	5	DDT	SAE	KZNE				1					
FAMILY DYSDERIDAE C.L. Koch, 1837														
B	*Dysdera crocata* C.L. Koch, 1838	0	LC	C			1	1						1
FAMILY ENTYPESIDAE Bond, Opatova & Hedin, 2020														
B	*Afropesa gauteng* Zonstein & Ríos-Tamayo, 2021	6	DD	SAE	GE			1						
B	*Afropesa schoutedeni* (Benoit, 1965)	6	LC	SAE	LE					1				
B	*Afropesa schwendingeri* Zonstein & Ríos-Tamayo, 2021	6	DD	SAE	LE					1				
B	*Brachytheliscus bicolor* Pocock, 1897	5	VU	SAE	KZNE				1					
B	*Hermacha brevicauda* Purcell, 1903	5	DDT	SAE	WCE									1
B	*Hermacha curvipes* Purcell, 1902	3	LC	SAE					1					1
B	*Hermacha evanescens* Purcell, 1903	3	DDT	SAE	NCE	1						1		1
M	*Hermacha fulva* Tucker, 1917	6	DDT	SAE	WCE									1
B	*Hermacha lanata* Purcell, 1902	5	DDT	SAE	WCE									1
M	*Hermacha maraisae* Ríos-Tamayo, Engelbrecht & Golobof, 2021	6	DDT	SAE	NCE							1		
B	*Hermacha montana* Ríos-Tamayo, Engelbrecht & Golobof, 2021	5	LC	SAE	WCE									1
B	*Hermacha nigrispinosa* Tucker, 1917	5	LC	SAE	WCE									1
J	*Hermacha purcelli* (Simon, 1903)	6	DDT	SAE	WCE									1
B	*Hermacha septemtrionalis* Ríos-Tamayo, Engelbrecht & Goloboff, 2021	3	LC	SAE				1		1	1		1	
B	*Hermacha sericea* Purcell, 1902	4	LC	SAE								1		1
B	*Hermacha tuckeri* Raven, 1985	5	DD	SAE	WCE									1
B	*Hermachola capensis* Ríos-Tamayo, Engelbrecht & Goloboff, 2021	6	DD	SAE	WCE									1
B	*Hermachola crudeni* (Hewitt, 1913)	5	DDT	SAE	ECE	1								
B	*Hermachola lyleae* Ríos-Tamayo, Engelbrecht & Goloboff, 2021	5	LC	SAE	ECE	1								
B	*Lepthercus confusus* Ríos-Tamayo & Lyle, 2020	5	LC	SAE	KZNE				1					
B	*Lepthercus dippenaarae* Ríos-Tamayo & Lyle, 2020	5	DD	SAE	ECE	1								
B	*Lepthercus dregei* Purcell, 1902	5	LC	SAE	ECE	1								
B	*Lepthercus engelbrechti* Ríos-Tamayo & Lyle, 2020	4	DD	SAE		1								1
F	*Lepthercus filmeri* Ríos-Tamayo & Lyle, 2020	5	DD	SAE	ME						1			
B	*Lepthercus haddadi* Ríos-Tamayo & Lyle, 2020	4	DD	SAE		1								1
B	*Lepthercus kwazuluensis* Ríos-Tamayo & Lyle, 2020	3	LC	SAE		1			1					
F	*Lepthercus lawrencei* Ríos-Tamayo & Lyle, 2020	5	DD	SAE	ECE	1								
B	*Lepthercus mandelai* Ríos-Tamayo & Lyle, 2020	6	DD	SAE	ECE	1								
B	*Lepthercus rattrayi* Hewitt, 1917	5	LC	SAE	ECE	1								
M	*Lepthercus sofiae* Ríos-Tamayo & Lyle, 2020	5	LC	SAE	WCE									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY ERESIDAE C.L. Koch, 1845														
B	*Dresserus angusticeps* Purcell, 1904	5	DDT	SAE	WCE									1
F	*Dresserus collinus* Pocock, 1900	5	DDT	SAE	WCE									1
F	*Dresserus colsoni* Tucker, 1920	3	LC	SAE			1	1	1	1	1			
B	*Dresserus kannemeyeri* Tucker, 1920	4	LC	SAE			1	1						
F	*Dresserus laticeps* Purcell, 1904	5	DDT	SAE	NCE							1		
F	*Dresserus namaquensis* Purcell, 1908	5	LC	SAE	NCE							1		
F	*Dresserus nigellus* Tucker, 1920	5	DDT	SAE	WCE									1
F	*Dresserus obscurus* Pocock, 1898	5	DDT	SAE	KZNE				1					
F	*Dresserus olivaceus* Pocock, 1900	6	DDT	SAE	ECE	1								
B	*Dresserus schreineri* Tucker, 1920	4	DD	SAE								1		1
F	*Dresserus tripartitus* Lawrence, 1938	6	DDT	SAE	KZNE				1					
B	*Gandanameno fumosa* (C.L. Koch, 1837)	3	LC	SAE		1	1			1		1	1	1
F	*Gandanameno purcelli* (Tucker, 1920)	3	LC	SAE		1		1	1	1		1		1
B	*Gandanameno spenceri* (Pocock, 1900)	2	LC	STHE		1	1		1			1	1	1
M	*Paradonea parva* (Tucker, 1920)	2	LC	STHE			1			1	1	1	1	
M	*Paradonea presleyi* Miller, Griswold, Scharff, Rezac, Szuts & Marhabaie, 2012	2	LC	STHE						1	1			
M	*Paradonea splendens* (Lawrence, 1936)	3	DDT	SAE			1	1				1		
M	*Paradonea striatipes* Lawrence, 1968	2	LC	STHE								1		
B	*Paradonea variegata* (Purcell, 1904)	2	LC	STHE								1		1
B	*Seothyra fasciata* Purcell, 1904	2	LC	STHE			1			1		1		
B	*Seothyra longipedata* Dippenaar-Schoeman, 1991	2	LC	STHE								1		1
M	*Seothyra perelegans* Simon, 1906	4	DDT	SAE		1	1							
B	*Seothyra schreineri* Purcell, 1903	2	LC	STHE			1					1		1
M	*Seothyra semicoccinea* Simon, 1906	6	DDT	SAE	ECE	1								
B	*Stegodyphus africanus* (Blackwall, 1866)	1	LC	AE					1	1	1	1	1	
B	*Stegodyphus bicolor* (O.P. Cambridge, 1869)	2	LC	STHE								1		
B	*Stegodyphus dumicola* Pocock, 1898	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Stegodyphus mimosarum* Pavesi, 1883	1	LC	AE		1	1	1	1	1	1	1		1
F	*Stegodyphus sabulosus* Tullgren, 1910	1	LC	AE						1	1			
B	*Stegodyphus tentoriicola* Purcell, 1904	2	LC	STHE		1	1	1		1		1		1
FAMILY EUAGRIDAE Raven, 1979														
B	*Allothele australis* (Purcell, 1903)	4	LC	SAE		1								1
B	*Allothele caffer* (Pocock, 1902)	4	LC	SAE		1			1					
B	*Allothele malawi* Coyle, 1984	1	LC	AE				1		1	1			
B	*Allothele teretis* Tucker, 1920	3	LC	SAE	WCE						1			1
F	*Euagrus atropurpureus* Purcell, 1903	6	DD	SAE										1
FAMILY FILISTATIDAE Simon, 1864														
B	*Andoharano ansieae* Zonstein & Marusik, 2015	2	LC	STHE						1				
FAMILY GALLIENIELLIDAE Millot, 1947														
B	*Austrachelas bergi* Haddad, Lyle, Bosselaers & Ramirez, 2009	4	LC	SAE						1	1			
F	*Austrachelas entabeni* Haddad & Mbo, 2017	6	DDT	SAE	LE					1				
B	*Austrachelas incertus* Lawrence, 1938	5	VU	SAE	KZNE				1					
M	*Austrachelas kalaharinus* Haddad, Lyle, Bosselaers & Ramirez, 2009	4	DDT	SAE		1	1					1		
F	*Austrachelas merwei* Haddad, Lyle, Bosselaers & Ramirez, 2009	6	DD	SAE	KZNE				1					
B	*Austrachelas natalensis* Lawrence, 1942	5	LC	SAE	KZNE				1					
B	*Austrachelas pondoensis* Haddad, Lyle, Bosselaers & Ramirez, 2009	6	DD	SAE	ECE	1								
B	*Austrachelas reavelli* Haddad, Lyle, Bosselaers & Ramirez, 2009	5	DD	SAE	KZNE				1					

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
M	*Austrachelas sexoculata* Haddad, Lyle, Bosselaers & Ramirez, 2009	6	DDT	SAE	ECE	1								
B	*Austrachelas wassenaari* Haddad, Lyle, Bosselaers & Ramirez, 2009	5	Rare	SAE	KZNE				1					
B	*Drassodella amatola* Mbo & Haddad, 2019	5	LC	SAE	ECE	1								
B	*Drassodella aurostriata* Mbo & Haddad, 2019	5	LC	SAE	WCE									1
M	*Drassodella baviaans* Mbo & Haddad, 2019	6	DDT	SAE	ECE	1								
B	*Drassodella flava* Mbo & Haddad, 2019	3	LC	SAE					1	1	1			
B	*Drassodella guttata* Mbo & Haddad, 2019	6	DD	SAE	FSE		1							
F	*Drassodella lotzi* Mbo & Haddad, 2019	6	DDT	SAE	KZNE				1					
F	*Drassodella maculata* Mbo & Haddad, 2019	5	LC	SAE	ECE	1								
B	*Drassodella melana* Tucker, 1923	3	LC	SAE					1	1	1			
B	*Drassodella montana* Mbo & Haddad, 2019	5	LC	SAE	KZNE				1					
F	*Drassodella purcelli* Tucker, 1923	5	DDT	SAE	WCE									1
B	*Drassodella quinquelabecula* Tucker, 1923	5	LC	SAE	WCE									1
B	*Drassodella salisburyi* Hewitt, 1916	5	LC	SAE	ECE		1							
B	*Drassodella septemmaculata* (Strand, 1909)	4	LC	SAE			1							1
B	*Drassodella tenebrosa* Lawrence, 1938	5	EN	SAE	KZNE				1					
B	*Drassodella tolkieni* Mbo & Haddad, 2019	5	DD	SAE	ECE	1								
B	*Drassodella transversa* Mbo & Haddad, 2019	5	DD	SAE	ME						1			
F	*Drassodella trilineata* Mbo & Haddad, 2019	6	DDT	SAE	ECE	1								
B	*Drassodella vasivulva* Tucker, 1923	5	LC	SAE	WCE									1
B	*Drassodella venda* Mbo & Haddad, 2019	5	LC	SAE	LE					1				
FAMYLY GNAPHOSIDAE Banks, 1892														
B	*Ammoxenus amphalodes* Dippenaar & Meyer, 1980	3	LC	SAE			1	1	1	1	1	1		
B	*Ammoxenus coccineus* Simon, 1893	2	LC	STHE								1	1	1
B	*Ammoxenus daedalus* Dippenaar & Meyer, 1980	5	DD	SAE	LE					1				
F	*Ammoxenus kalaharicus* Benoit, 1972	2	LC	STHE								1		1
B	*Ammoxenus pentheri* Simon, 1896	2	LC	STHE		1	1	1				1		1
B	*Ammoxenus psammodromus* Simon, 1910	2	LC	STHE			1			1			1	1
B	*Amusia cataracta* Tucker, 1923	2	LC	STHE			1		1					1
F	*Aneplasa balnearia* Tucker, 1923	5	DDT	SAE	WCE									1
F	*Aneplasa facies* Tucker, 1923	4	LC	SAE		1								1
F	*Aneplasa interrogationis* Tucker, 1923	3	LC	SAE						1				1
F	*Aneplasa nigra* Tucker, 1923	4	LC	SAE								1		1
M	*Aneplasa primaris* Tucker, 1923	5	DDT	SAE	WCE									1
F	*Aneplasa sculpturata* Tucker, 1923	5	DDT	SAE	WCE									1
M	*Aphantaulax australis* Simon, 1893	6	DDT	SAE	ECE	1								
M	*Aphantaulax inornata* Tucker, 1923	2	LC	STHE				1	1	1	1	1		1
F	*Aphantaulax signicollis* Tucker, 1923	2	LC	STHE		1			1	1	1	1		1
B	*Aphantaulax stationis* Tucker, 1923	2	LC	STHE		1		1						1
B	*Asemesthes albovittatus* Purcell, 1908	2	LC	STHE		1	1					1		1
F	*Asemesthes ales* Tucker, 1923	6	DDT	SAE	ECE	1								
B	*Asemesthes ceresicola* Tucker, 1923	3	LC	SAE		1	1	1	1	1	1	1	1	1
F	*Asemesthes decoratus* Purcell, 1908	2	LC	STHE				1	1	1	1	1		1
F	*Asemesthes flavipes* Purcell, 1908	2	LC	STHE						1				
M	*Asemesthes fodina* Tucker, 1923	2	LC	STHE						1				
F	*Asemesthes lamberti* Tucker, 1923	5	DDT	SAE	WCE									1
F	*Asemesthes lineatus* Purcell, 1908	1	LC	AE			1			1		1		1
M	*Asemesthes modestus* Dalmas, 1921	6	DDT	SAE	LE					1				
B	*Asemesthes montanus* Tucker, 1923	3	LC	SAE			1					1		1
F	*Asemesthes nigristernus* Dalmas, 1921	6	DDT	SAE	?									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Asemesthes numisma* Tucker, 1923	2	LC	STHE				1		1	1		1	
B	*Asemesthes oconnori* Tucker, 1923	3	LC	SAE			1			1		1		1
F	*Asemesthes pallidus* Purcell, 1908	3	LC	SAE						1		1	1	
F	*Asemesthes paynteri* Tucker, 1923	3	LC	SAE				1	1	1	1	1		1
F	*Asemesthes purcelli* Tucker, 1923	2	LC	STHE			1	1		1	1	1		1
B	*Asemesthes reflexus* Tucker, 1923	3	LC	SAE		1	1			1	1			1
B	*Asemesthes subnubilus* Simon, 1887	2	LC	STHE								1		1
B	*Camillina aldabrae* (Strand, 1907)	1	LC	AE		1	1		1					1
B	*Camillina biplagia* Tucker, 1923	3	LC	SAE		1			1			1	1	1
B	*Camillina capensis* Platnick & Murphy, 1987	3	LC	SAE		1		1						1
B	*Camillina cordifera* (Tullgren, 1910)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Camillina maun* Platnick & Murphy, 1987	1	LC	AE			1		1	1	1	1	1	1
B	*Camillina pavesii* (Simon, 1897)	1	LC	AE						1	1			1
B	*Camillina procurva* (Purcell, 1908)	1	LC	STHE		1				1	1	1		1
B	*Camillina setosa* Tucker, 1923	3	LC	SAE			1		1		1			1
F	*Diaphractus leipoldti* Purcell, 1907	3	LC	SAE			1					1		1
F	*Drassodes bechuanicus* Tucker, 1923	2	LC	STHE			1	1	1	1				
M	*Drassodes caffrerianus* Purcell, 1907	6	DDT	SAE	ECE	1								
F	*Drassodes calceatus* Purcell, 1907	6	DDT	SAE	WCE									1
F	*Drassodes dregei* Purcell, 1907	6	DDT	SAE	ECE	1								
B	*Drassodes ereptor* Purcell, 1907	3	LC	SAE			1	1	1			1	1	1
F	*Drassodes gooldi* Purcell, 1907	6	DDT	SAE	WCE									1
M	*Drassodes helenae* Purcell, 1907	3	LC	SAE				1		1		1		1
B	*Drassodes lophognathus* Purcell, 1907	3	LC	SAE		1	1	1	1	1		1		1
F	*Drassodes lyratus* Purcell, 1907	6	DDT	SAE	WCE									1
M	*Drassodes masculus* Tucker, 1923	2	LC	STHE				1		1				
B	*Drassodes sesquidentatus* Purcell, 1908	3	LC	SAE								1		
F	*Drassodes solitarius* Purcell, 1907	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Drassodes splendens* Tucker, 1923	2	LC	STHE		1	1	1	1	1	1	1	1	1
M	*Drassodes stationis* Tucker, 1923	3	LC	SAE		1	1	1	1	1	1	1		1
B	*Drassodes tesselatus* Purcell, 1907	3	LC	SAE		1	1					1		
F	*Drassodes tortuosus* Tucker, 1923	6	DDT	SAE	KZNE				1					
B	*Echemus erutus* Tucker, 1923	2	LC	STHE					1	1		1		1
F	*Eilica fusca* Platnick, 1975	6	DDT	SAE	ECE	1								
M	*Eilica lotzi* FitzPatrick, 2002	5	DDT	SAE	FSE		1							
B	*Ibala arcus* (Tucker, 1923)	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Ibala bilinearis* (Tucker, 1923)	2	LC	STHE		1				1	1	1		1
B	*Ibala bulawayensis* (Tucker, 1923)	2	LC	STHE						1		1		
B	*Ibala lapidaria* (Lawrence, 1928)	2	LC	STHE			1	1		1				
F	*Ibala okorosave* FitzPatrick, 2009	2	LC	STHE								1		
F	*Leptodrassus bergensis* Tucker, 1923	6	DDT	SAE	WCE									1
F	*Leptodrassus licentiosus* Dalmas, 1919	6	DDT	SAE	WCE									1
B	*Marinarozelotes jaxartensis* (Kroneberg, 1875)	0	LC	C		1	1	1	1	1	1	1	1	1
M	*Megamyrmaekion schreineri* Tucker, 1923	2	LC	STHE			1				1	1	1	1
F	*Megamyrmaekion transvaalense* Tucker, 1923	3	LC	SAE		1	1	1		1	1	1	1	1
F	*Megamyrmaekion velox* Simon, 1887	2	LC	STHE								1		
B	*Micaria basaliducta* Booysen & Haddad, 2021	5	LC	SAE	WCE									1
B	*Micaria beaufortia* Tucker, 1923	1	LC	AE		1	1	1	1	1	1	1		1
B	*Micaria bispicula* Booysen & Haddad, 2021	2	LC	STHE								1		1
B	*Micaria chrysis* (Simon, 1910)	1	LC	AE		1	1	1	1	1	1	1		1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Micaria durbana* Booysen & Haddad, 2021	1	LC	AE					1					
B	*Micaria felix* Booysen & Haddad, 2021	1	LC	AE		1	1	1	1	1	1			1
M	*Micaria koingnaas* Booysen & Haddad, 2021	4	DD	SAE								1		1
F	*Micaria lata* Booysen & Haddad, 2021	2	LC	STHE								1		
M	*Micaria laxa* Booysen & Haddad, 2021	6	DD	SAE	ECE	1								
M	*Micaria mediospina* Booysen & Haddad, 2021	6	DD	SAE	ECE	1								
B	*Micaria quinquemaculosa* Booysen & Haddad, 2021	2	LC	STHE			1					1		
M	*Micaria sanipass* Booysen & Haddad, 2021	6	DD	SAE	KZNE				1					
M	*Micaria scutellata* Booysen & Haddad, 2021	5	DD	SAE	KZNE				1					
M	*Micaria tersissima* Simon, 1910	6	DDT	SAE	NCE							1		
M	*Nomisia australis* Dalmas, 1921	6	DDT	SAE	WCE									1
F	*Nomisia frenata* (Purcell, 1908)	6	DDT	SAE	NCE							1		
F	*Nomisia notia* Dalmas, 1921	5	DDT	SAE	NCE							1		
F	*Nomisia transvaalica* Dalmas, 1921	4	LC	SAE				1		1	1			
M	*Nomisia tubula* (Tucker, 1923)	2	LC	STHE			1		1	1			1	
B	*Nomisia varia* (Tucker, 1923)	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Odontodrassus aphanes* (Thorell, 1897)	0	LC	C						1				
F	*Poecilochroa anomala* (Hewitt, 1915)	2	LC	STHE		1			1					1
F	*Poecilochroa capensis* Strand, 1909	3	LC	SAE					1			1		1
F	*Poecilochroa involuta* Tucker, 1923	3	LC	SAE			1		1					1
B	*Pterotricha auris* (Tucker, 1923)	3	LC	SAE		1	1	1		1		1		1
B	*Rastellus deserticola* Haddad, 2003	2	LC	STHE								1		1
B	*Rastellus florisbad* Platnick & Griffin, 1990	3	DD	SAE			1		1					
B	*Rastellus kariba* Platnick & Griffin, 1990	2	LC	STHE						1				
F	*Scotophaeus marleyi* Tucker, 1923	3	LC	SAE			1		1	1			1	
F	*Scotophaeus natalensis* Lawrence, 1938	2	LC	STHE			1		1					
F	*Scotophaeus purcelli* Tucker, 1923	3	LC	SAE				1	1					
M	*Scotophaeus relegatus* Purcell, 1907	2	LC	STHE			1	1	1					1
B	*Setaphis browni* (Tucker, 1923)	0	LC	C			1	1	1	1	1	1	1	1
B	*Setaphis makalali* Fitzpatrick, 2005	6	DD	SAE	LE					1				
B	*Setaphis sexmaculata* Simon, 1893	4	DDT	SAE								1	1	
B	*Setaphis subtilis* (Simon, 1897)	0	LC	C		1	1	1	1	1	1	1	1	**1**
F	*Smionia capensis* Dalmas, 1920	6	DDT	SAE	?									
B	*Smionia lineatipes* (Purcell, 1908)	2	LC	STHE		1	1	1				1	1	1
F	*Trephopoda aplanita* (Tucker, 1923)	4	LC	SAE			1							1
F	*Trephopoda biamenta* (Tucker, 1923)	5	DDT	SAE	WCE									1
F	*Trephopoda hanoveria* Tucker, 1923	5	DDT	SAE	NCE							1		
B	*Trephopoda kannemeyeri* (Tucker, 1923)	3	LC	SAE		1	1							
M	*Trephopoda parvipalpa* (Tucker, 1923)	2	LC	STHE					1	1	1		1	
F	*Trichothyse africana* (Tucker, 1923)	2	LC	STHE		1	1	1	1		1	1	1	1
F	*Trichothyse hortensis* Tucker, 1923	2	LC	STHE			1		1					
B	*Urozelotes rusticus* (L. Koch, 1872)	0	LC	C			1	1	1	1	1	1	1	1
F	*Xerophaeus ahenus* Purcell, 1908	4	LC	SAE								1		1
F	*Xerophaeus anthropoides* Hewitt, 1916	3	LC	SAE				1			1	1	1	
B	*Xerophaeus appendiculatus* Purcell, 1907	3	LC	SAE		1	1	1	1	1		1		1
B	*Xerophaeus aridus* Purcell, 1907	2	LC	STHE			1					1		
B	*Xerophaeus aurariarum* Purcell, 1907	2	LC	STHE		1	1	1		1	1	1	1	1
B	*Xerophaeus bicavus* Tucker, 1923	3	LC	SAE		1		1	1	1	1	1	1	1
F	*Xerophaeus biplagiatus* Tullgren, 1910	1	LC	AE				1						
B	*Xerophaeus capensis* Purcell, 1907	4	LC	SAE								1		1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Xerophaeus communis* Purcell, 1907	3	LC	SAE		1			1			1		1
B	*Xerophaeus coruscus kibonotensis* Tullgren, 1910	1	LC	AE		1								
B	*Xerophaeus crusculus* Tucker, 1923	3	LC	SAE		1			1			1		1
M	*Xerophaeus crustosus* Purcell, 1907	5	DDT	SAE	ECE	1								
B	*Xerophaeus exiguus* Purcell, 1907	4	DD	SAE		1								1
F	*Xerophaeus flammeus* Tucker, 1923	5	DDT	SAE	WCE									1
M	*Xerophaeus flavescens* Purcell, 1907	4	DDT	SAE								1		1
F	*Xerophaeus hottentottus* Purcell, 1908	3	LC	SAE		1						1	1	
B	*Xerophaeus lightfooti* Purcell, 1907	3	LC	SAE		1								1
B	*Xerophaeus longispinus* Purcell, 1908	3	DD	SAE					1			1		
B	*Xerophaeus lunulifer* Purcell, 1907	3	LC	SAE		1			1	1				1
F	*Xerophaeus maritimus* Lawrence, 1938	5	DDT	SAE	KZNE				1					
F	*Xerophaeus matroosbergensis* Tucker, 1923	4	LC	SAE		1								1
M	*Xerophaeus pallidus* Tucker, 1923	5	DDT	SAE	KZNE				1					
M	*Xerophaeus patricki* Purcell, 1907	2	LC	STHE				1		1				
F	*Xerophaeus phaseolus* Tucker, 1923	5	DDT	SAE	WCE									1
B	*Xerophaeus rostratus* Purcell, 1907	3	LC	SAE		1	1	1						
F	*Xerophaeus rubeus* Tucker, 1923	2	LC	STHE					1					1
F	*Xerophaeus silvaticus* Tucker, 1923	3	LC	SAE		1								
B	*Xerophaeus spiralifer* Purcell, 1907	4	LC	SAE			1					1		1
B	*Xerophaeus spoliator* Purcell, 1907	2	LC	STHE						1		1	1	1
F	*Xerophaeus tenebrosus* Tucker, 1923	3	LC	SAE						1		1		1
B	*Xerophaeus vickermani* Tucker, 1923	3	LC	SAE			1	1		1		1	1	
F	*Xerophaeus zuluensis* Lawrence, 1938	3	LC	SAE					1					1
B	*Zelotes aestus* (Tucker, 1923)	2	LC	STHE				1		1				
F	*Zelotes albanicus* (Hewitt, 1915)	3	LC	SAE		1						1		1
B	*Zelotes aridus* (Purcell, 1907)	1	LC	AE						1				1
B	*Zelotes bastardi* (Simon, 1896)	1	LC	AE				1		1				
B	*Zelotes broomi* (Purcell, 1907)	5	LC	SAE	WCE									1
B	*Zelotes caldarius* (Purcell, 1907)	2	LC	STHE						1				1
B	*Zelotes capensis* FitzPatrick, 2007	3	LC	SAE		1	1							1
B	*Zelotes capsula* Tucker, 1923	3	LC	SAE		1		1						1
B	*Zelotes chinguli* Fitzpatrick, 2007	2	LC	STHE						1				
B	*Zelotes corrugatus* (Purcell, 1907)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Zelotes doddieburni* FitzPatrick, 2007	2	LC	STHE						1				
F	*Zelotes flavitarsis* (Purcell, 1908)	5	DDT	SAE	NCE							1		
M	*Zelotes florisbad* FitzPatrick, 2007	3	LC	SAE			1					1		1
B	*Zelotes frenchi* Tucker, 1923	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Zelotes fuligineus* (Purcell, 1907)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Zelotes gooldi* (Purcell, 1907)	2	LC	STHE		1	1					1		1
B	*Zelotes haplodrassoides* (Denis, 1955)	1	LC	AE				1		1				1
B	*Zelotes humilis* (Purcell, 1907)	2	LC	STHE		1	1	1	1	1		1	1	1
F	*Zelotes invidus* (Purcell, 1907)	2	LC	STHE		1	1							1
F	*Zelotes kuncinyanus* FitzPatrick, 2007	6	DDT	SAE	WCE									1
B	*Zelotes lavus* Tucker, 1923	2	LC	STHE			1	1	1	1		1	1	1
F	*Zelotes lightfooti* (Purcell, 1907)	3	LC	SAE		1				1	1			1
B	*Zelotes lotzi* FitzPatrick, 2007	3	LC	SAE			1			1	1			
F	*Zelotes mashonus* FitzPatrick, 2007	1	LC	AE					1		1			
M	*Zelotes muizenbergensis* FitzPatrick, 2007	6	DDT	SAE	WCE									1
M	*Zelotes namaquus* FitzPatrick, 2007	3	LC	SAE						1		1		

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Zelotes namibensis* FitzPatrick, 2007	2	LC	STHE					1					
B	*Zelotes natalensis* Tucker, 1923	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Zelotes ngomensis* FitzPatrick, 2007	5	LC	SAE	KZNE				1					
B	*Zelotes otavi* Fitzpatrick, 2007	2	LC	STHE						1				
F	*Zelotes ovambensis* Lawrence, 1927	2	LC	STHE								1		
M	*Zelotes pallidipes* Tucker, 1923	2	LC	STHE			1		1	1	1			
B	*Zelotes pulchripes* (Purcell, 1908)	6	DD	SAE	NCE							1		
F	*Zelotes qwabergensis* FitzPatrick, 2007	5	DDT	SAE			1		1					
B	*Zelotes radiatus* Lawrence, 1928	2	LC	STHE						1				
B	*Zelotes reduncus* (Purcell, 1907)	2	LC	STHE		1	1	1	1	1	1	1		1
B	*Zelotes resolution* FitzPatrick, 2007	5	LC	SAE	ECE	1								
B	*Zelotes sclateri* Tucker, 1923	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Zelotes scrutatus* (O.P.-Cambridge, 1872)	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Zelotes songus* FitzPatrick, 2007	5	DDT	SAE	LE					1				
B	*Zelotes tuckeri* Roewer, 1951	1	LC	AE		1	1	1	1	1	1			
B	*Zelotes uquathus* FitzPatrick, 2007	3	LC	SAE			1	1	1	1	1	1		1
B	*Zelotes zonognathus* (Purcell, 1907)	1	LC	AE			1		1	1		1	1	
FAMILY HAHNIIDAE Bertkau, 1878														
B	*Hahnia abrahami* (Hewitt, 1915)	5	DD	SAE	WCE									1
F	*Hahnia clathrata* Simon, 1898	2	LC	STHE		1			1		1			1
B	*Hahnia larseni* Marusik, 2017	6	DD	SAE	WCE									1
F	*Hahnia laticeps* Simon, 1898	3	LC	SAE		1			1					1
M	*Hahnia lobata* Bosmans, 1981	3	LC	SAE			1		1					1
B	*Hahnia schubotzi* Strand, 1913	1	LC	AE							1			1
B	*Hahnia tabulicola* Simon, 1898	1	LC	AE		1	1	1	1	1	1			1
F	*Hahnia zodarioides* (Simon, 1898)	3	LC	SAE										1
FAMILY HERSILIIDAE Thorell, 1869														
B	*Hersilia arborea* Lawrence, 1928	2	LC	STHE				1	1	1		1		
B	*Hersilia sagitta* Foord & Dippenaar-Schoeman, 2006	1	LC	AE					1	1				
B	*Hersilia sericea* Pocock, 1898	1	LC	AE		1		1	1	1	1	1	1	1
B	*Hersilia setifrons* Lawrence, 1928	2	LC	STHE				1	1	1	1	1	1	1
B	*Neotama corticola* (Lawrence, 1937)	3	LC	SAE		1			1					1
F	*Tyrotama abyssus* Foord & Dippenaar-Schoeman, 2005	2	LC	STHE			1					1		1
B	*Tyrotama arida* (Smithers, 1945)	3	LC	SAE						1		1		1
B	*Tyrotama australis* (Simon, 1893)	2	LC	STHE			1	1	1	1		1	1	1
B	*Tyrotama bicava* (Smithers, 1945)	2	LC	STHE						1				
F	*Tyrotama incerta* (Tucker, 1920)	2	LC	STHE								1		1
B	*Tyrotama soutpansbergensis* Foord & Dippenaar-Schoeman, 2005	5	VU	SAE	LE					1				
F	*Tyrotama taris* Foord & Dippenaar-Schoeman, 2005	6	DDT	SAE	NCE							1		
FAMILY IDIOPIDAE Simon, 1889														
M	*Ctenolophus cregoei* (Purcell, 1902)	3	LC	SAE				1	1					
F	*Ctenolophus fenoulheti* Hewitt, 1913	3	LC	SAE				1		1	1			
F	*Ctenolophus kolbei* (Purcell, 1902)	4	LC	SAE		1								1
F	*Ctenolophus oomi* Hewitt, 1913	3	LC	SAE					1	1	1			
M	*Ctenolophus pectinipalpis* (Purcell, 1903)	4	DDT	SAE			1		1					
B	*Ctenolophus spiricola* (Purcell, 1903)	4	LC	SAE		1			1					
F	*Galeosoma coronatum* Hewitt, 1915	4	DDT	SAE			1						1	
F	*Galeosoma crinitum* Hewitt, 1919	6	DDT	SAE	NWE								1	
F	*Galeosoma hirsutum* Hewitt, 1916	5	EN	SAE	GE			1						
B	*Galeosoma pallidum* Hewitt, 1916	5	EN	SAE	GE			1						

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Galeosoma planiscutatum* Hewitt, 1919	3	LC	SAE				1		1			1	
F	*Galeosoma pluripunctatum* Hewitt, 1919	6	DDT	SAE	NWE								1	
F	*Galeosoma robertsi* Hewitt, 1916	4	VU	SAE				1					1	
F	*Galeosoma schreineri* Hewitt, 1913	6	DDT	SAE	NCE							1		
F	*Galeosoma scutatum* Purcell, 1903	3	EN	SAE			1	1					1	
B	*Galeosoma vandami* Hewitt, 1913	5	LC	SAE	LE					1				
F	*Gorgyrella inermis* Hewitt, 1913	4	DDT	SAE		1								1
F	*Gorgyrella namaquensis* Purcell, 1902	6	DDT	SAE	NCE							1		
F	*Gorgyrella schreineri* Purcell, 1903	3	LC	SAE		1	1	1				1		1
F	*Heligmomerus caffer* Purcell, 1903	5	DDT	SAE	LE					1				
F	*Idiops castaneus* Hewitt, 1913	4	LC	SAE						1	1			
B	*Idiops crudeni* (Hewitt, 1914)	5	DD	SAE	ECE	1								
B	*Idiops flaveolus* (Pocock, 1901)	5	DD	SAE	ECE	1								
F	*Idiops fryi* (Purcell, 1903)	3	LC	SAE			1	1					1	
F	*Idiops gerhardti* Hewitt, 1913	6	DDT	SAE	ME						1			
B	*Idiops gracilipes* (Hewitt, 1919)	6	DD	SAE	ECE	1								
F	*Idiops grandis* (Hewitt, 1915)	5	DDT	SAE	KZNE				1					
F	*Idiops gunningi* Hewitt, 1913	4	DDT	SAE				1	1	1				
B	*Idiops hamiltoni* (Pocock, 1902)	6	DD	SAE	FSE		1							
B	*Idiops hepburni* (Hewitt, 1919)	2	DDT	STHE			1							
B	*Idiops hirsutus* (Hewitt, 1919)	6	DD	SAE	ECE	1								
B	*Idiops kentanicus* (Purcell, 1903)	6	DD	SAE	ECE	1								
F	*Idiops microps* (Hewitt, 1913)	6	DDT	SAE	ECE	1								
B	*Idiops monticola* Hewitt, 1916	3	LC	SAE		1	1	1	1	1	1		1	
B	*Idiops mossambicus* (Hewitt, 1919)	2	LC	STHE					1		1			
B	*Idiops nigropilosus* (Hewitt, 1919)	4	DD	SAE				1			1			
M	*Idiops ochreolus* (Pocock, 1902)	3	DDT	SAE		1						1		
F	*Idiops parvus* Hewitt, 1915	6	DDT	SAE	FSE		1							
B	*Idiops pretoriae* (Pocock, 1898)	5	VU	SAE	GE			1						
M	*Idiops pullus* Tucker, 1917	4	LC	SAE								1	1	
M	*Idiops sylvestris* (Hewitt, 1925)	6	DDT	SAE	LE?					1				
M	*Idiops thorelli* O.P.-Cambridge, 1870	6	DDT	SAE										
B	*Idiops vandami* (Hewitt, 1925)	6	DD	SAE	MPE						1			
B	*Segregara abrahami* (Hewitt, 1913)	5	DD	SAE	ECE	1								
F	*Segregara paucispinulosus* (Hewitt, 1915)	5	LC	SAE	LE					1				
F	*Segregara transvaalensis* (Hewitt, 1913)	3	LC	SAE			1	1	1	1	1			
FAMILY ISCHNOTHELIDAE F. O. Pickard-Cambridge, 1897														
B	*Thelechoris striatipes* (Simon, 1889)	2	LC	AE					1	1				
FAMILY LINYPHIIDAE Blackwall, 1859														
M	*Afribactrus stylifrons* Wunderlich, 1995	6	DDT	SAE	WCE									1
B	*Agyneta gracilipes* (Holm, 1968)	1	LC	AE										1
B	*Agyneta habra* (Locket, 1968)	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Agyneta natalensis* (Jocqué, 1984)	3	LC	SAE				1	1	1			1	1
B	*Agyneta prosectes* (Locket, 1968)	1	LC	AE				1					1	1
B	*Agyneta prosectoides* (Locket & Russell-Smith, 1980)	1	LC	AE			1	1	1	1	1		1	
M	*Callitrichia minuta* (Jocqué, 1984)	6	DDT	SAE	GE			1						
F	*Ceratinopsis dippenaari* Jocqué, 1984	3	LC	SAE					1			1		1
B	*Ceratinopsis idanrensis* Locket & Russell-Smith, 1980	1	LC	AE			1			1				
B	*Erigone irrita* Jocqué, 1984	3	LC	SAE				1	1				1	
B	*Erigonops littoralis* (Hewitt, 1915)	5	VU	SAE	WCE									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Frontinellina locketi* van Helsdingen, 1970	3	LC	SAE		1			1					1
B	*Lepthyphantes rimicola* Lawrence, 1964	5	CR	SAE	WCE									1
B	*Limoneta sirimoni* (Bosmans, 1979)	2	LC	AE		1	1	1	1	1	1			1
F	*Lucrinus putus* O.P.-Cambridge, 1904	6	DDT	SAE	WCE									1
M	*Mecynidis dentipalpis* Simon, 1894	2	LC	STHE		1				1				1
B	*Mermessus fradeorum* (Berland, 1932)	0	LC	C			1	1		1	1	1	1	1
B	*Metaleptyphantes familiaris* Jocqué, 1984	3	LC	SAE		1	1					1	1	1
B	*Metaleptyphantes perexiguus* (Simon & Fage, 1922)	1	LC	AE			1	1	1	1	1		1	
B	*Microlinyphia sterilis* (Pavesi, 1883)	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Neriene flammea* van Helsdingen, 1969	4	DDT	SAE		1			1					
B	*Neriene natalensis* van Helsdingen, 1969	3	LC	SAE		1			1	1				
F	*Notioscopus australis* Simon, 1894	6	DDT	SAE	?									
B	*Ostearius melanopygius* (O.P.-Cambridge, 1879)	0	LC	C		1	1	1	1	1	1	1	1	1
M	*Pelecopsis intricata* Jocqué, 1984	4	LC	SAE		1								1
B	*Pelecopsis janus* Jocqué, 1984	2	LC	STHE		1	1	1	1	1	1	1	1	1
M	*Pelecopsis medusoides* Jocqué, 1984	6	DDT	SAE	WCE									1
B	*Prinerigone vagans* (Audioun, 1826)	0	LC	C		1	1		1					1
B	*Proelauna humicola* (Miller, 1970)	1	LC	AE			1		1			1		
B	*Pseudomicrocentria minutissima* Miller, 1970	1	LC	AE					1					
M	*Toschia minuta* Jocqué, 1984	6	DDT	SAE	GE			1						
B	*Tybaertiella krugeri* (Simon, 1894)	1	LC	AE			1	1	1	1	1		1	
FAMILY LIOCRANIDAE Simon, 1897														
B	*Andromma raffrayi* Simon, 1899	4	DD	SAE		1			1					
M	*Coryssiphus cinerascens* Simon, 1903	5	DDT	SAE	WCE									1
M	*Coryssiphus praeustus* Simon, 1903	5	DDT	SAE	WCE									1
F	*Coryssiphus unicolor* Simon, 1903	6	DDT	SAE	WCE									1
F	*Rhaeboctesis denotatus* Lawrence, 1928	2	LC	STHE			1					1		
F	*Rhaeboctesis equestris* Simon, 1897	6	DDT	SAE	?									
M	*Rhaeboctesis exilis* Tucker, 1920	3	LC	SAE					1	1	1			
F	*Rhaeboctesis matroosbergensis* Tucker, 1920	5	LC	SAE	WCE									1
B	*Rhaeboctesis secundus* Tucker, 1920	2	LC	STHE		1	1					1	1	1
B	*Rhaeboctesis transvaalensis* Tucker, 1920	3	LC	SAE			1	1		1	1			
F	*Rhaeboctesis trinotatus* Tucker, 1920	2	LC	STHE			1	1	1	1	1	1	1	
FAMILY LYCOSIDAE Sundevall, 1833														
B	*Allocosa algoensis* (Pocock, 1900)	6	DD	SAE	ECE	1								
F	*Allocosa aurata* (Purcell, 1903)	3	LC	SAE			1		1	1	1			
F	*Allocosa aurichelis* Roewer, 1959	4	DDT	SAE		1						1		
B	*Allocosa exserta* Roewer, 1959	2	LC	STHE				1		1				
B	*Allocosa faberrima* (Simon, 1910)	2	LC	STHE										1
B	*Allocosa gracilitarsis* (Purcell, 1903)	2	LC	STHE		1		1		1		1		
F	*Allocosa kalaharihensis* (Simon, 1910)	2	LC	STHE								1		
B	*Allocosa lawrencei* (Roewer, 1951)	2	LC	STHE				1	1	1	1	1		
B	*Allocosa montana* Roewer, 1959	1	LC	AE				1	1	1	1			
B	*Allocosa nebulosa* (Roewer, 1960)	1	LC	AE		1	1	1						
F	*Allocosa schoenlandi* (Pocock, 1900)	4	DDT	SAE		1	1							
F	*Allocosa testacea* Roewer, 1959	3	DDT	SAE				1		1			1	
B	*Allocosa tuberculipalpa* (Caporiacco, 1940)	1	LC	AE		1	1	1	1	1	1			
B	*Allocosa umtalica* (Purcell, 1903)	1	LC	AE				1		1				
B	*Amblyothele albocincta* Simon, 1910	2	LC	STHE			1					1		
F	*Amblyothele ecologica* Russell-Smith, Jocqué & Alderweireldt, 2009	3	LC	SAE				1		1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Amblyothele latedissipata* Russell-Smith, Jocqué & Alderweireldt, 2009	1	LC	AE		1			1	1				1
F	*Arctosa albida* (Simon, 1898)	6	DDT	SAE	WCE?									1
B	*Arctosa brevispina* (Lessert, 1915)	1	LC	AE					1	1				
B	*Arctosa capensis* Roewer, 1960	6	DD	SAE	?									
F	*Arctosa lawrencei* (Roewer, 1960)	6	DDT	SAE	?									
B	*Arctosa lightfooti* (Purcell, 1903)	5	DD	SAE	WCE									1
B	*Arctosa nivosa* (Purcell, 1903)	3	LC	SAE		1						1		1
B	*Arctosa oneili* (Purcell, 1903)	3	DD	SAE		1						1		1
F	*Arctosa promontorii* (Purcell, 1900)	3	LC	SAE		1	1						1	1
F	*Arctosa tenuissima* (Purcell, 1903)	6	DDT	SAE	ECE	1								
M	*Arctosa transvaalana* Roewer, 1960	5	DDT	SAE	LE?					1				
J	*Artoria lycosimorpha* Strand, 1909	6	DDT	SAE	ECE	1								
F	*Artoriellula bicolor* (Simon, 1898)	5	DDT	SAE	WCE									1
B	*Evippomma plumipes* (Lessert, 1936)	1	LC	AE			1	1	1					
B	*Evippomma squamulatum* (Simon, 1898)	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Foveosa adunca* Russell-Smith, Alderweireldt & Jocqué, 2007	3	LC	SAE			1	1			1	1	1	
B	*Foveosa foveolata* (Purcell, 1903)	1	LC	AE		1	1	1	1	1	1	1		1
F	*Geolycosa hectoria* (Pocock, 1900)	5	DDT	SAE	WCE									1
B	*Geolycosa natalensis* Roewer, 1960	6	DD	SAE	KZNE				1					
B	*Geolycosa nolotthensis* (Simon, 1910)	2	DD	STHE								1		
B	*Geolycosa subvittata* (Pocock, 1900)	4	LC	SAE		1								1
B	*Hippasa australis* Lawrence, 1927	1	LC	AE		1	1	1	1	1	1		1	
F	*Hippasa elienae* Alderweireldt & Jocqué, 2005	1	LC	AE						1				
B	*Hippasa funerea* Lessert, 1925	2	LC	STHE		1	1	1	1	1	1			1
B	*Hippasosa dewinterae* (Alderweireldt, 1996)	1	LC	AE						1				
B	*Hippasosa guttata* (Karsch, 1878)	1	LC	AE				1	1	1	1			
F	*Hogna adjacens* Roewer, 1959	3	DDT	SAE						1		1		
F	*Hogna bimaculata* (Purcell, 1903)	2	LC	STHE		1	1	1		1				1
F	*Hogna denisi* Roewer, 1959	6	DDT	SAE	WCE									1
M	*Hogna deweti* Roewer, 1959	6	DDT	SAE	?									
M	*Hogna idonea* Roewer, 1959	6	DDT	SAE	ECE	1								
B	*Hogna infulata* Roewer, 1959	6	DD	SAE	ECE	1								
F	*Hogna lawrencei* (Roewer, 1960)	3	LC	SAE				1	1			1		
B	*Hogna schreineri* (Purcell, 1903)	2	LC	STHE						1		1		
B	*Hogna simoni* Roewer, 1959	1	LC	AE					1					
B	*Hogna spenceri* (Pocock, 1898)	1	LC	AE			1	1	1	1	1	1	1	
B	*Hogna transvaalica* (Simon, 1898)	3	LC	SAE			1	1		1	1	1		
B	*Hogna unicolor* Roewer, 1959	2	LC	STHE										1
F	*Hogna zuluana* Roewer, 1959	3	LC	SAE				1	1	1		1		
B	*Lycosa capensis* Simon, 1898	6	DD	SAE	WCE?									1
F	*Lycosa connexa* Roewer, 1960	6	DDT	SAE	FSE		1							
F	*Lycosa gigantea* (Roewer, 1960)	6	DDT	SAE	NWE								1	
F	*Lycosa inviolata* Roewer, 1960	6	DDT	SAE	NCE							1		
F	*Lycosa pachana* Pocock, 1898	1	LC	AE				1	1		1		1	
F	*Lycosa palliata* Roewer, 1960	6	DDT	SAE	?									
M	*Lycosa perspicua* Roewer, 1960	6	DDT	SAE	WCE									1
B	*Lycosa rimicola* Purcell, 1903	4	DD	SAE								1		1
B	*Minicosa neptuna* Alderweireldt & Jocqué, 2006	3	LC	STHE					1	1	1	1		1
B	*Pardosa clavipalpis* Purcell, 1903	1	LC	AE			1			1	1	1	1	1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Pardosa crassipalpis* Purcell, 1903	2	LC	STHE		1	1	1	1	1	1	1	1	1
F	*Pardosa enucleata* Roewer, 1959	6	DDT	SAE	MPE?						1			
B	*Pardosa injucunda* (O.P.-Cambridge, 1876)	1	LC	AE					1	1		1	1	
B	*Pardosa leipoldti* Purcell, 1903	2	LC	STHE				1	1	1				1
F	*Pardosa lycosina* Purcell, 1903	4	DDT	SAE		1								1
B	*Pardosa manubriata* Simon, 1898	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Pardosa nostrorum* Alderweireldt & Jocqué, 1992	2	LC	STHE		1			1					
B	*Pardosa schreineri* Purcell, 1903	2	LC	STHE		1			1			1		
B	*Pardosa umtalica* Purcell, 1903	1	LC	AE						1	1			
F	*Passiena auberti* (Simon, 1898)	6	DDT	SAE	LE					1				
F	*Pirata africana* (Roewer, 1960)	2	LC	STHE					1	1	1			
B	*Proevippa albiventris* (Simon, 1898)	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Proevippa biampliata* (Purcell, 1903)	2	LC	STHE		1	1		1	1	1	1		1
B	*Proevippa bruneipes* (Purcell, 1903)	2	LC	STHE		1			1	1	1	1		1
B	*Proevippa dregei* (Purcell, 1903)	5	LC	SAE	ECE	1								
B	*Proevippa fascicularis* (Purcell, 1903)	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Proevippa hirsuta* (Russell-Smith, 1981)	2	LC	STHE					1	1	1			
B	*Proevippa lightfooti* Purcell, 1903	5	LC	SAE	WCE									1
B	*Proevippa schreineri* (Purcell, 1903)	2	LC	STHE		1	1	1	1			1	1	1
M	*Proevippa wanlessi* (Russell-Smith, 1981)	3	LC	SAE				1	1	1				
B	*Pterartoria arbuscula* (Purcell, 1903)	3	LC	SAE		1					1	1		
B	*Pterartoria caldaria* Purcell, 1903	4	LC	SAE		1								1
B	*Pterartoria cederbergensis* Russell-Smith & Roberts, 2017	5	CR	SAE	WCE									1
B	*Pterartoria confusa* Russell-Smith & Roberts, 2017	4	LC	SAE								1		1
F	*Pterartoria fissivittata* Purcell, 1903	6	DDT	SAE	ECE	1								
B	*Pterartoria flavolimbata* Purcell, 1903	5	DD	SAE	WCE									1
B	*Pterartoria lativittata* Purcell, 1903	4	LC	SAE				1				1		1
B	*Pterartoria polysticta* Purcell, 1903	5	DD	SAE	WCE									1
F	*Pterartoria sagae* (Purcell, 1903)	6	DDT	SAE	WCE									1
B	*Pterartoria subcrucifera* (Purcell, 1903)	4	LC	SAE								1		1
B	*Schizocosa darlingi* (Pocock, 1898)	2	LC	STHE					1	1				
B	*Schizocosa subpersonata* (Simon, 1910)	2	LC	STHE								1		
B	*Trabea heteroculata* Strand, 1913	1	LC	AE						1				
M	*Trabea natalensis* Russell-Smith, 1982	2	LC	STHE					1	1				
B	*Trabea nigriceps* Purcell, 1903	5	LC	SAE	ECE	1								
B	*Trabea ornatipalpis* Russell-Smith, 1982	3	LC	SAE		1		1	1	1		1		1
B	*Trabea purcelli* Roewer, 1951	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Trabea rubriceps* Lawrence, 1952	2	LC	STHE		1	1		1					1
M	*Trabea unicolor* Purcell, 1903	5	DDT	SAE	WCE									1
B	*Trabea varia* Purcell, 1903	5	LC	SAE	WCE									1
B	*Tricassa deserticola* Simon, 1910	2	LC	STHE								1		1
F	*Trochosa albipilosa* (Roewer, 1960)	6	DDT	SAE	ECE	1								
F	*Trochosa eugeni* (Roewer 1951)	6	DDT	SAE	NCE							1		
F	*Trochosa modesta* Roewer, 1960	6	DDT	SAE	MPE						1			
F	*Trochosa nigerrima* Roewer, 1960	6	DDT	SAE	MPE						1			
B	*Wadicosa oncka* Lawrence, 1927	1	LC	AE		1		1	1	1	1			1
B	*Zenonina albocaudata* Lawrence, 1952	3	LC	SAE			1	1	1	1				1
F	*Zenonina mystacina* Simon, 1898	2	LC	STHE			1		1	1	1	1		1
FAMILY MICROSTIGMATIDAE Roewer, 1942														
B	*Microstigmata amatola* Griswold, 1985	5	Rare	SAE	ECE	1								

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Microstigmata geophila* (Hewitt, 1916)	5	DD	SAE	ECE	1								
F	*Microstigmata lawrencei* Griswold, 1985	4	DDT	SAE		1			1					
B	*Microstigmata longipes* (Lawrence, 1938)	5	LC	SAE	KZNE				1					
B	*Microstigmata ukhahlamba* Griswold, 1985	4	LC	SAE		1			1					
B	*Microstigmata zuluensis* (Lawrence, 1938)	4	LC	SAE		1			1					
FAMILY MIGIDAE Simon, 1889														
F	*Moggridgea albimaculata* Hewitt, 1925	6	DDT	SAE	LE					1				
F	*Moggridgea ampullata* Griswold, 1987	6	DDT	SAE	WCE									1
F	*Moggridgea breyeri* Hewitt, 1915	5	DDT	SAE	LE					1				
F	*Moggridgea crudeni* Hewitt, 1913	5	DDT	SAE	ECE	1								
F	*Moggridgea dyeri* O.P.-Cambridge, 1875	4	LC	SAE		1			1					
B	*Moggridgea intermedia* Hewitt, 1913	5	Rare	SAE	WCE									1
F	*Moggridgea leipoldti* Purcell, 1903	6	DDT	SAE	WCE									1
F	*Moggridgea loistata* Griswold, 1987	5	EN	SAE	WCE									1
F	*Moggridgea microps* Hewitt, 1915	3	LC	SAE		1			1		1			
F	*Moggridgea mordax* Purcell, 1903	5	DDT	SAE	WCE									1
F	*Moggridgea paucispina* Hewitt, 1916	3	LC	SAE				1		1	1		1	
B	*Moggridgea peringueyi* Simon, 1903	3	LC	SAE		1	1					1		1
B	*Moggridgea pseudocrudeni* Hewitt, 1919	4	DDT	SAE		1								1
F	*Moggridgea pymi* Hewitt, 1914	2	LC	STHE						1				
B	*Moggridgea quercina* Simon, 1903	5	EN	SAE	WCE									1
B	*Moggridgea rupicola* Hewitt, 1913	4	DD	SAE		1								1
F	*Moggridgea rupicoloides* Hewitt, 1914	5	DDT	SAE	ECE	1								
B	*Moggridgea teresae* Griswold, 1987	5	Rare	SAE	WCE									1
F	*Moggridgea terrestris* Hewitt, 1914	6	DDT	SAE	ECE	1								
B	*Moggridgea terricola* Simon, 1903	5	VU	SAE	WCE									1
B	*Poecilomigas abrahami* (O.P.-Cambridge, 1889)	2	LC	STHE		1			1			1		
M	*Poecilomigas elegans* Griswold, 1987	6	DDT	SAE	KZNE				1					
FAMILY MIMETIDAE Simon, 1881														
B	*Anansi natalensis* (Lawrence, 1938)	3	LC	SAE		1	1	1	1	1	1	1		1
F	*Ero capensis* Simon, 1895	2	LC	STHE		1					1	1		1
B	*Ero lawrencei* Unzicker, 1966	2	LC	STHE		1			1	1				
F	*Mimetus cornutus* Lawrence, 1947	2	LC	STHE					1	1				
FAMILY MITURGIDAE Simon, 1886														
M	*Parapostenus hewitti* Lessert, 1923	3	LC	SAE		1			1	1	1			1
F	*Voraptus affinis* Lessert, 1925	3	DDT	SAE		1			1					1
FAMILY MYSMENIDAE Petrunkevitch, 1928														
B	*Isela okuncana* Griswold, 1985	4	DD	SAE		1			1					
FAMILY NESTICIDAE Simon, 1894														
B	*Nesticella benoiti* (Hubert, 1970	2	LC	STHE				1			1	1		
FAMILY OECOBIIDAE Blackwall, 1862														
B	*Oecobius navus* Blackwall, 1859	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Oecobius putus* O. P.-Cambridge, 1876	0	LC	C			1					1		
B	*Paroecobius nicolaii* Wunderlich, 1995	4	DD	SAE						1			1	
F	*Uroctea quinquenotata* Simon, 1910	4	LC	SAE								1		1
F	*Uroctea schinzi* Simon, 1887	2	LC	STHE								1		
F	*Uroctea septemnotata* Tucker, 1920	2	LC	STHE								1		
B	*Uroecobius ecribellatus* Kullmann & Zimmermann, 1976	3	LC	SAE			1			1			1	

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY OONOPIDAE Simon, 1890														
B	*Australoonops granulatus* Hewitt, 1915	3	LC	SAE		1	1							1
B	*Australoonops haddadi* Platnick & Dupérré, 2010	2	LC	STHE					1					
B	*Australoonops skaife* Platnick & Dupérré, 2011	4	LC	SAE		1								
F	*Dalmasula dodebai* Szűts & Ubig, 2012	6	DDT	SAE	NCE							1		
B	*Dalmasula griswoldi* Szűts & Ubig, 2012	6	DD	SAE	WCE									1
B	*Dysderina capensis* Simon, 1907	6	DD	SAE	WCE?									1
B	*Dysderina speculifera* Simon, 1907	2	LC	STHE			1	1	1		1	1	1	
B	*Gamasomorpha australis* Hewitt, 1915	3	LC	SAE		1		1		1	1			
F	*Gamasomorpha humicola* Lawrence, 1947	3	LC	SAE			1	1	1	1		1		1
M	*Gamasomorpha longisetosa* Lawrence, 1952	5	DDT	SAE	KZNE				1					
M	*Oonops caecus* Benoit, 1975	2	LC	STHE										1
B	*Opopaea mattica* Simon, 1893	4	LC	SAE			1							1
B	*Opopaea speciosa* (Lawrence, 1952)	1	LC	AE		1		1	1	1			1	1
F	*Orchestina cincta* Simon, 1893	6	DDT	SAE	?									
B	*Orchestina fannesi* Henrad & Jocqué, 2012	2	LC	STHE						1				
F	*Pseudoscaphiella parasita* Simon, 1907	6	DDT	SAE	WCE?									1
F	*Telchius transvaalicus* Simon, 1907	6	DDT	SAE	LE					1				
FAMILY ORSOLOBIDAE Cooke, 1965														
F	*Afrilobus australis* Griswold & Platnick, 1987	5	DDT	SAE	WCE									1
M	*Afrilobus capensis* Griswold & Platnick, 1987	4	LC	SAE								1		1
B	*Azanialobus lawrencei* Griswold & Platnick, 1987	2	LC	STHE			1		1	1	1			
F	*Calculus bicolor* Purcell, 1910	6	DD	SAE	WCE									1
FAMILY OXYOPIDAE Thorell, 1869														
F	*Hamataliwa fronticornis* (Lessert, 1927)	1	LC	AE		1			1	1	1			
B	*Hamataliwa kulczynskii* (Lessert, 1915)	1	LC	AE				1	1	1	1	1		1
F	*Hamataliwa rostrifrons* (Lawrence, 1928)	2	LC	STHE		1			1	1	1		1	
F	*Hamataliwa rufocaligata* Simon, 1898	1	LC	AE						1				
F	*Hamataliwa strandi* (Lessert, 1923)	2	LC	STHE		1				1				
B	*Oxyopes affinis* Lessert, 1915	1	LC	AE		1	1	1	1	1			1	1
M	*Oxyopes angulitarsus* Lessert, 1915	1	LC	AE					1	1	1			
F	*Oxyopes bedoti* Lessert, 1915	1	LC	AE					1	1				
M	*Oxyopes bonneti* Lessert, 1933	2	LC	STHE						1				
F	*Oxyopes bothai* Lessert, 1915	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Oxyopes castaneus* Lawrence, 1927	2	LC	STHE					1					
F	*Oxyopes chapini* Lessert, 1927	1	LC	AE					1					
M	*Oxyopes cornifrons avakubensis* Lessert, 1927	1	LC	AE		1								
B	*Oxyopes dumonti* (Vinson, 1863)	1	LC	AE		1				1				
B	*Oxyopes falconeri* Lessert, 1915	1	LC	AE						1	1			
B	*Oxyopes flavipalpis* (Lucas, 1858)	1	LC	AE		1	1	1	1	1	1		1	1
F	*Oxyopes galla* Caporiacco, 1941	1	LC	AE							1			
B	*Oxyopes hoggi* Lessert, 1915	1	LC	AE			1	1	1	1	1	1	1	1
B	*Oxyopes jacksoni* Lessert, 1915	1	LC	AE			1	1	1	1	1	1	1	1
B	*Oxyopes longispinosus* Lawrence, 1938	3	LC	SAE		1	1	1	1	1	1		1	1
B	*Oxyopes pallidecoloratus* Strand, 1906	1	LC	AE		1	1	1	1	1	1		1	1
F	*Oxyopes personatus* Simon, 1896	6	DD	SAE	WCE									1
F	*Oxyopes russoi* Caporiacco, 1940	1	LC	AE		1	1	1	1	1				1
B	*Oxyopes schenkeli* Lessert, 1917	1	LC	AE				1	1	1	1	1	1	
B	*Oxyopes singularis* Lessert, 1927	1	LC	AE					1	1				
M	*Oxyopes sjostedti* Lessert, 1915	1	LC	AE					1	1				

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Oxyopes subabebae* Caporiacco 1941	1	LC	AE					1					
B	*Oxyopes tuberculatus* Lessert, 1915	1	LC	AE					1	1	1			
B	*Oxyopes uncinatus* Lessert, 1915	1	LC	AE					1	1				
M	*Oxyopes vanderysti* Lessert, 1946	1	LC	AE				1		1				
B	*Oxyopes vogelsangeri* Lessert, 1946	1	LC	AE		1	1	1	1	1	1			1
B	*Peucetia crucifera* Lawrence, 1927	2	LC	STHE						1	1	1		1
B	*Peucetia lucasi* (Vinson, 1863)	1	LC	AE						1				
B	*Peucetia maculifera* Pocock, 1900	2	LC	STHE		1			1			1		1
B	*Peucetia madalenae* Van Niekerk & Dippenaar-Schoeman, 1994	2	LC	STHE					1	1				
B	*Peucetia nicolae* Van Niekerk & Dippenaar-Schoeman, 1994	3	LC	SAE		1						1		1
B	*Peucetia pulchra* (Blackwall, 1865)	1	LC	AE					1		1			
B	*Peucetia striata* Karsch, 1878	1	LC	AE			1	1	1	1	1	1	1	1
B	*Peucetia transvaalica* Simon, 1896	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Peucetia viridis* (Blackwall, 1858)	1	LC	AE		1	1	1	1	1	1	1	1	1
FAMILY PALPIMANIDAE Thorell, 1870														
B	*Diaphorocellus biplagiatus* Simon, 1893	2	LC	STHE			1			1	1	1		1
B	*Palpimanus armatus* Pocock, 1898	3	LC	SAE					1	1	1			
J	*Palpimanus aureus* Lawrence, 1927	2	DDT	STHE						1				
B	*Palpimanus capensis* Simon, 1893	3	LC	SAE		1						1		1
M	*Palpimanus crudeni* Lessert, 1936	4	DDT	SAE		1								1
M	*Palpimanus giltrayi* Lessert, 1936	2	LC	STHE								1		
M	*Palpimanus globulifer* Simon, 1893	3	DDT	SAE		1								
F	*Palpimanus leppanae* Pocock, 1902	6	DDT	SAE	ECE	1								
F	*Palpimanus namaquensis* Simon, 1910	2	LC	STHE			1					1		
F	*Palpimanus paroculus* Simon, 1910	6	DDT	SAE	NCE							1		
F	*Palpimanus potteri* Lawrence, 1937	3	LC	SAE					1	1				
B	*Palpimanus pseudarmatus* Lawrence, 1952	3	LC	SAE					1	1				
M	*Palpimanus subarmatus* Lawrence, 1947	6	DDT	SAE	KZNE				1					
F	*Palpimanus transvaalicus* Simon, 1893	3	LC	SAE		1	1	1	1	1	1		1	
M	*Palpimanus tuberculatus* Lawrence, 1952	6	DDT	SAE	KZNE				1					
FAMILY PENESTOMIDAE Simon, 1903														
M	*Penestomus armata* (Lehtinen, 1967)	4	DDT	SAE		1								1
F	*Penestomus croeseri* Dippenaar-Schoeman, 1989	6	DDT	SAE	ECE	1								
B	*Penestomus egazini* Miller, Griswold & Haddad, 2010	5	Rare	SAE	ECE	1								
F	*Penestomus kruger* Miller, Griswold & Haddad, 2010	5	DDT	SAE	WCE									1
B	*Penestomus montanus* Miller, Griswold & Haddad, 2010	2	Rare	STHE		1			1					
F	*Penestomus planus* Simon, 1902	4	DDT	SAE		1								1
F	*Penestomus prendinii* Miller, Griswold & Haddad, 2010	4	DDT	SAE		1								1
F	*Penestomus stilleri* (Dippenaar-Schoeman, 1989)	5	DDT	SAE	WCE									1
F	*Penestomus zulu* Miller, Griswold & Haddad, 2010	6	DDT	SAE	KZNE				1					
FAMILY PHILODROMIDAE														
B	*Gephyrota glauca* (Jézéquel, 1966)	1	LC	AE		1	1	1	1	1	1	1		1
B	*Hirriusa arenacea* (Lawrence, 1927)	2	LC	STHE		1	1		1	1		1	1	1
B	*Hirriusa bidentata* (Lawrence, 1927)	2	LC	STHE			1			1			1	1
F	*Hirriusa variegata* (Simon, 1895)	3	LC	SAE		1	1	1		1	1	1	1	1
F	*Philodromus bigibbus australis* Lawrence, 1928	3	LC	SAE		1	1	1		1		1	1	
F	*Philodromus brachycephalus* Lawrence, 1952	1	LC	AE		1	1	1	1	1	1	1		
F	*Philodromus browningi* Lawrence, 1952	2	LC	STHE		1	1	1	1	1	1	1		1
F	*Philodromus epigynatus* Strand, 1909	6	DDT	SAE	WCE									1
M	*Philodromus grosi* Lessert, 1943	1	LC	AE			1	1		1	1			1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Philodromus guineensis* Millot, 1941	1	LC	AE			1	1	1	1	1			1
F	*Philodromus partitus* Lessert, 1919	1	LC	AE		1		1	1	1				
J	*Philodromus thanatellus* Strand, 1909	6	DDT	SAE	WCE									1
F	*Philodromus vulpio* Simon, 1910	6	DDT	SAE	NCE							1		
B	*Suemus punctatus* Lawrence, 1938	2	LC	STHE				1	1	1	1	1		1
F	*Thanatus africanus* Karsch, 1878	1	LC	AE						1				
B	*Thanatus atlanticus* Berland, 1936	1	LC	AE			1	1		1		1		
B	*Thanatus dorsilineatus* Jézéquel, 1964	1	LC	AE			1	1	1	1	1		1	
B	*Thanatus fabricii* (Audouin, 1826)	0	LC	C					1	1	1			
B	*Thanatus lamottei* Jézéquel, 1964	1	LC	AE			1			1				1
F	*Thanatus namaquensis* Simon, 1910	5	DDT	SAE	NCE							1		
B	*Thanatus vulgaris* Simon, 1870	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Tibellus armatus* Lessert, 1928	1	LC	AE			1		1					
F	*Tibellus australis* (Simon, 1910)	2	LC	STHE						1				
F	*Tibellus bruneitarsis* Lawrence, 1952	2	LC	STHE					1	1				
M	*Tibellus cobusi* Van den Berg & Dippenaar-Schoeman, 1994	1	LC	AE						1				
B	*Tibellus demangei* Jézéquel, 1964	1	LC	AE					1					
B	*Tibellus flavipes* Caporiacco, 1939	1	LC	AE					1					
B	*Tibellus gerhardi* Van den Berg & Dippenaar-Schoeman, 1994	1	LC	AE				1	1	1				
B	*Tibellus hollidayi* Lawrence, 1952	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Tibellus kibonotensis* Lessert, 1919	1	LC	AE				1	1	1	1			
B	*Tibellus minor* Lessert, 1919	1	LC	AE		1	1	1	1	1	1		1	1
B	*Tibellus seriepunctatus* Simon, 1907	1	LC	AE		1					1			
B	*Tibellus sunetae* Van den Berg & Dippenaar-Schoeman, 1994	2	LC	STHE					1	1				
B	*Tibellus vossioni* Simon, 1884	1	LC	AE				1						1
FAMILY PHOLCIDAE C. L. Koch, 1850														
B	*Artema atlanta* Walckenaer, 1837	0	LC	C								1		
B	*Crossopriza lyoni* (Blackwall, 1867)	0	LC	C				1		1				
B	*Leptopholcus gracilis* Berland, 1920	1	LC	AE		1			1	1				
B	*Pehrforsskalia conopyga* Deeleman-Reinhold & van Harten, 2001	0	LC	C					1					
B	*Quamtana bonamanzi* Huber, 2003	3	LC	SAE		1			1	1	1			
B	*Quamtana ciliata* (Lawrence, 1938)	3	Rare	SAE					1			1		
B	*Quamtana embuleni* Huber, 2003	3	Rare	SAE						1			1	
B	*Quamtana entabeni* Huber, 2003	5	Rare	SAE	LE					1				
B	*Quamtana filmeri* Huber, 2003	4	LC	SAE			1		1					
B	*Quamtana hectori* Huber, 2003	3	LC	SAE				1	1	1			1	
B	*Quamtana knysna* Huber, 2003	5	EN	SAE	WCE									1
M	*Quamtana lajuma* Huber, 2003	5	DDT	SAE	LE					1				
M	*Quamtana leleupi* Huber, 2003	5	DDT	SAE	KZNE				1					
B	*Quamtana leptopholcica* (Strand, 1909)	5	CR	SAE	WCE									1
B	*Quamtana lotzi* Huber, 2003	5	DD	SAE	FSE		1							
B	*Quamtana mabusai* Huber, 2003	3	LC	STHE			1		1	1	1			
B	*Quamtana mbaba* Huber, 2003	6	CR	SAE	KZNE				1					
B	*Quamtana merwei* Huber, 2003	6	CR	SAE	KZNE				1					
B	*Quamtana meyeri* Huber, 2003	6	CR	SAE	KZNE				1					
B	*Quamtana nandi* Huber, 2003	6	DD/CR	SAE	KZNE				1					
B	*Quamtana nylsvley* Huber, 2003	6	CR	SAE	LE					1				
B	*Quamtana tsui* Huber, 2003	4	DD	SAE		1			1					
B	*Quamtana umzinto* Huber, 2003	6	CR	SAE	KZNE				1					
B	*Quamtana vidal* Huber, 2003	3	LC	SAE		1			1					1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Pholcus phalangioides* (Fuesslin, 1775)	0	LC	C								1		
B	*Smeringopus atomarius* Simon, 1910	2	LC	STHE						1		1		
B	*Smeringopus badplaas* Huber, 2012	4	LC	SAE						1	1			
B	*Smeringopus blyde* Huber, 2012	6	CR	SAE	ME						1			
B	*Smeringopus dehoop* Huber, 2012	5	EN	SAE	WCE									1
B	*Smeringopus florisbad* Huber, 2012	4	LC	SAE			1	1						
B	*Smeringopus hanglip* Huber, 2012	5	Rare	SAE	LE					1				
B	*Smeringopus hypocrita* Simon, 1910	2	LC	STHE								1	1	1
B	*Smeringopus koppies* Huber, 2012	2	LC	STHE			1					1	1	
B	*Smeringopus lesnei* Lessert, 1936	2	LC	STHE					1					
B	*Smeringopus lotzi* Hubert, 2012	3	LC	SAE			1					1	1	
B	*Smeringopus lydenberg* Hubert, 2012	6	CR	SAE	ME						1			
B	*Smeringopus mlilwane* Huber, 2012	2	Rare	STHE							1			
B	*Smeringopus natalensis* Lawrence, 1947	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Smeringopus ndumo* Huber, 2012	5	Rare	SAE	KZNE				1					
B	*Smeringopus sederberg* Huber, 2012	4	LC	SAE			1							1
B	*Smeringopus ubicki* Huber, 2012	4	LC	SAE		1								1
B	*Spermophora gordimerae* Huber, 2003	5	Rare	SAE	WCE									1
M	*Spermophora pembai* Huber, 2003	5	DDT	SAE	ECE	1								
B	*Spermophora peninsulae* Lawrence, 1964	5	Rare	SAE	WCE									1
B	*Spermophora schoemanae* Huber, 2003	6	CR	SAE	WCE									1
M	*Spermophora suurbraak* Huber, 2003	6	DDT	SAE	WCE									1
FAMILY PHYXELIDIDAE Lehtinen, 1967														
M	*Lamaika distincta* Griswold, 1990	6	DDT	SAE	WCE									1
B	*Malaika delicatula* Griswold, 1990	5	DD	SAE	WCE									1
B	*Malaika longipes* (Purcell, 1904)	5	Rare	SAE	WCE									1
B	*Matundua silvatica* (Purcell, 1904)	5	DD	SAE	WCE									1
F	*Namaquarachne angulata* Griswold, 1990	5	DD	SAE	NCE							1		
F	*Namaquarachne hottentotta* (Pocock, 1900)	6	DD	SAE	NCE							1		
B	*Namaquarachne khoikhoiana* Griswold, 1990	6	DD	SAE	NCE							1		
B	*Namaquarachne thaumatula* Griswold, 1990	5	DD	SAE	NCE							1		
B	*Namaquarachne tropata* Griswold, 1990	5	LC	SAE	WCE									1
B	*Phyxelida makapanensis* Simon, 1894	3	LC	SAE				1		1	1			
F	*Pongolania chrysionaria* Griswold, 1990	4	LC	SAE				1			1			
F	*Pongolania pongola* Griswold, 1990	6	DDT	SAE	ME						1			
B	*Themacrys cavernicola* (Lawrence, 1939)	2	LC	STHE					1		1			
B	*Themacrys irrorata* Simon, 1906	3	LC	SAE		1			1		1			
B	*Themacrys monticola* (Lawrence, 1939)	5	LC	SAE	KZNE				1					
B	*Themacrys silvicola* (Lawrence, 1938)	4	VU	SAE		1			1					
F	*Themacrys ukhahlamba* Griswold, 1990	5	DDT	SAE	KZNE				1					
B	*Vidole capensis* (Pocock, 1900)	3	LC	SAE		1						1		1
B	*Vidole helicigyna* Griswold, 1990	5	VU	SAE	KZNE				1					
F	*Vidole lyra* Griswold, 1990	3	LC	SAE		1	1	1						
B	*Vidole schreineri* (Purcell, 1904)	3	LC	SAE		1						1		
B	*Vidole sothoana* Griswold, 1990	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Xevioso amica* Griswold, 1990	5	LC	SAE	KZNE				1					
B	*Xevioso aululata* Griswold, 1990	5	LC	SAE	ME						1			
M	*Xevioso colobata* Griswold, 1990	3	LC	SAE					1	1	1			
B	*Xevioso kulufa* Griswold, 1990	3	LC	SAE					1	1	1			
B	*Xevioso lichmadina* Griswold, 1990	5	VU	SAE	LE					1				

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Xevioso orthomeles* Griswold, 1990	2	LC	STHE				1		1	1			
B	*Xevioso tuberculata* (Lawrence, 1939)	4	LC	SAE					1		1			
B	*Xevioso zuluana* (Lawrence, 1939)	5	DD	SAE	KZNE				1					
FAMILY PISAURIDAE Simon, 1890														
B	*Afropisaura ducis* (Strand, 1913)	1	LC	AE		1		1	1	1				
B	*Afropisaura rothiformis* (Strand, 1908)	1	LC	AE		1		1	1	1	1		1	
B	*Charminus aethiopicus* (Caporiacco, 1939)	1	LC	AE						1	1			
B	*Charminus ambiguus* (Lessert, 1925)	1	LC	AE					1	1				
B	*Charminus atomarius* (Lawrence, 1942)	1	LC	AE					1					
F	*Charminus natalensis* (Lawrence, 1947)	5	LC	SAE	KZNE				1					
F	*Chiasmopes hystrix* (Berland, 1922)	1	LC	AE							1			
B	*Chiasmopes lineatus* (Pocock, 1898)	1	LC	AE		1	1	1	1	1	1		1	1
F	*Chiasmopes namaquensis* (Roewer, 1955)	2	LC	STHE			1			1	1			
B	*Chiasmopes signatus* (Pocock, 1902)	6	DD	SAE	ECE	1								
F	*Cispius kimbius* Blandin, 1978	2	LC	STHE			1		1	1		1		
M	*Cispius problematicus* Blandin, 1978	1	LC	AE						1	1			
F	*Cispius variegatus* Simon, 1898	1	LC	AE		1			1	1				
F	*Dendrolycosa yukan* Jäger 2011	6	DD	WCE?										
B	*Euprosthenops australis* Simon, 1898	1	LC	AE		1	1	1	1	1	1	1	1	
B	*Euprosthenops bayaonianus* (Brito Capello, 1867)	1	LC	AE		1	1	1		1	1			
B	*Euprosthenops proximus* Lessert, 1916	1	LC	AE				1	1					
B	*Euprosthenopsis armata* (Strand, 1913)	1	LC	AE			1	1		1				
F	*Euprosthenopsis lamorali* Blandin, 1977	3	LC	SAE		1			1					1
B	*Euprosthenopsis pulchella* (Pocock, 1902)	2	LC	STHE		1	1	1		1	1	1		1
B	*Euprosthenopsis vuattouxi* Blandin, 1977	1	LC	AE		1	1	1	1	1	1		1	1
B	*Hygropoda tangana* (Roewer, 1955)	1	LC	AE		1				1				
B	*Maypacius bilineatus* (Pavesi, 1895)	1	LC	AE		1			1	1	1			
F	*Maypacius christophei* Blandin, 1975	1	LC	AE				1						
M	*Maypacius roeweri* Blandin, 1975	1	LC	AE				1		1				
F	*Maypacius stuhlmanni* (Bösenberg & Lenz, 1895)	1	LC	AE						1				
B	*Nilus curtus* O.P.-Cambridge, 1876	1	LC	AE		1	1	1	1	1	1			1
B	*Nilus margaritatus* (Pocock, 1898)	1	LC	AE				1	1	1	1	1		
B	*Nilus massajae* (Pavesi, 1883)	1	LC	AE		1			1	1	1			1
B	*Nilus radiatolineatus* Strand, 1906	1	LC	AE			1		1	1	1		1	1
B	*Nilus rossi* Pocock, 1902	1	LC	AE					1	1	1		1	
B	*Perenethis simoni* (Lessert, 1916)	1	LC	AE				1	1	1	1			
B	*Perenethis symmetrica* (Lawrence, 1927)	1	LC	AE					1	1	1			1
B	*Rothus aethiopicus* (Pavesi, 1883)	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Rothus auratus* Pocock, 1900	3	LC	SAE		1						1		1
F	*Rothus vittatus* Simon, 1898	3	LC	SAE		1	1					1		1
J	*Tapinothelella laboriosa* Strand, 1909	6	DDT	SAE	WCE									1
B	*Walrencea globosa* Blandin, 1979	4	DD	SAE		1			1					
FAMILY PRODIDOMIDAE Simon, 1884														
F	*Austrodomus scaber* (Purcell, 1904)	2	LC	STHE			1	1		1		1		1
F	*Austrodomus zuluensis* Lawrence, 1947	2	LC	STHE			1		1	1		1		
M	*Eleleis crinita* Simon, 1893	6	DDT	SAE	WCE									1
F	*Eleleis haddadi* Rodrigues & Rheims, 2020	6	DDT	SAE	FSE		1							
F	*Eleleis leleupi* Rodrigues & Rheims, 2020	6	DDT	SAE	WCE									1
B	*Eleleis limpopo* Rodrigues & Rheims, 2020	1	LC	AE				1		1				
F	*Namundra murphyi* Haddad, 2022	5	DDT	SAE	NCE							1		

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Prodidomus capensis* Purcell, 1904	3	LC	SAE		1				1				1
F	*Prodidomus flavipes* Lawrence, 1952	5	DDT	SAE	KZNE				1					
B	*Prodidomus purpurascens* Purcell, 1904	4	LC	SAE								1		1
M	*Prodidomus simoni* Dalmas, 1919	6	DDT	SAE	LE					1				
M	*Purcelliana cederbergensis* Rodrigues & Rheims, 2020	6	DDT	SAE	WCE									1
M	*Purcelliana problematica* Cooke, 1964	5	DDT	SAE	WCE									1
M	*Theuma ababensis* Tucker, 1923	2	LC	STHE			1			1		1		1
F	*Theuma aprica* Simon, 1893	6	DDT	SAE	?									
B	*Theuma capensis* Purcell, 1907	2	LC	STHE			1		1			1		1
F	*Theuma cedri* Purcell, 1907	3	LC	SAE			1			1		1		1
F	*Theuma elucubata* Tucker, 1923	3	LC	SAE			1	1	1	1			1	
F	*Theuma foveolata* Tucker, 1923	2	LC	STHE			1		1	1		1		
B	*Theuma fusca* Purcell, 1907	2	LC	STHE			1		1	1	1	1		1
B	*Theuma maculata* Purcell, 1907	2	LC	STHE			1	1	1	1	1	1		1
F	*Theuma mutica* Purcell, 1907	6	DDT	SAE	WCE									1
M	*Theuma parva* Purcell, 1907	2	LC	STHE				1	1	1		1		
F	*Theuma purcelli* Tucker, 1923	3	LC	SAE						1	1	1	1	1
F	*Theuma pusilla* Purcell, 1908	2	DDT	STHE								1		
B	*Theuma schreineri* Purcell, 1907	2	LC	STHE?		1	1					1	1	1
F	*Theuma schultzei* Purcell, 1908	2	LC	STHE								1	1	
M	*Theuma tragardhi* Lawrence, 1947	3	LC	SAE				1	1					
F	*Theuma zuluensis* Lawrence, 1947	5	DDT	SAE	KZNE				1					
FAMILY PYCNOTHELIDAE Chamberlin, 1917														
B	*Pionothele straminea* Purcell, 1902	5	VU	SAE	WCE									1
FAMILY SALTICIDAE Blackwall, 1841														
B	*Aelurillus cristatopalpus* Simon, 1902	4	DD	SAE								1		1
B	*Afraflacilla altera* (Wesołowska, 2000)	2	LC	STHE					1	1				
F	*Afraflacilla bipunctata* (Peckham & Peckham, 1903)	4	DDT	SAE		1								1
B	*Afraflacilla braunsi* (Peckham & Peckham, 1903)	1	LC	AE		1								1
B	*Afraflacilla elegans* (Wesołowska & Cumming, 2008)	2	LC	STHE			1	1		1			1	
F	*Afraflacilla histrionica* (Simon, 1902)	6	DDT	SAE	WCE?									1
B	*Afraflacilla imitator* (Wesołowska & Haddad, 2013)	5	DD	SAE	ECE	1								
M	*Afraflacilla karinae* (Haddad & Wesołowska, 2011)	6	DDT	SAE	FSE		1							
B	*Afraflacilla venustula* (Wesołowska& Haddad, 2009)	4	LC	SAE					1	1	1			
B	*Afraflacilla zuluensis* (Haddad & Wesołowska, 2013)	6	DD	SAE	KZNE				1					
B	*Afromarengo coriacea* (Simon, 1900)	1	LC	AE					1					
F	*Araegeus mimicus* Simon, 1901	6	DDT	SAE	LE					1				
F	*Asemonea amatola* Wesołowska & Haddad, 2013	6	DDT	SAE	ECE	1								
F	*Asemonea clara* Wesołowska & Haddad, 2013	3	LC	SAE		1			1	1	1	1		
B	*Asemonea murphyae* Wanless, 1980	1	LC	AE					1					
B	*Asemonea stella* Wanless, 1980	1	LC	AE		1			1		1			
B	*Baryphas ahenus* Simon, 1902	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Belippo calcarata* (Roewer, 1942)	1	LC	AE					1	1				
B	*Belippo meridionalis* Wesołowska & Haddad, 2013	3	LC	SAE				1	1	1				
B	*Belippo pulchra* Haddad & Wesołowska, 2013	3	LC	SAE					1	1				
B	*Bianor albobimaculatus* (Lucas, 1846)	0	LC	C		1		1		1			1	1
F	*Bianor eximius* Wesołowska & Haddad, 2009	2	LC	STHE					1					
M	*Vicirionessa mustela* (Simon, 1902)	2	LC	STHE		1			1		1			
B	*Thyene mutica* (Simon, 1902)	1	LC	AE				1	1	1	1	1		
F	*Carrhotus singularis* Simon, 1902	4	DDT	SAE		1	1							

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Cembalea heteropogon* (Simon, 1910)	2	LC	STHE								1		
B	*Cembalea plumosa* (Lessert, 1925)	1	LC	AE	SAE?									
B	*Cembalea triloris* Haddad & Wesołowska, 2011	2	LC	STHE								1		
M	*Chinophrys trifasciata* Wesołowska, Azarkina & Russell-Smith, 2014	5	Rare	SAE	WCE									1
B	*Cyrba boveyi* Lessert, 1933	1	LC	AE			1		1	1	1		1	
F	*Cyrba dotata* Peckham & Peckham, 1903	5	DDT	SAE	WCE									1
B	*Cyrba lineata* Wanless, 1984	2	LC	STHE					1	1	1			
B	*Cyrba nigrimana* Simon, 1900	3	LC	SAE		1	1	1	1	1	1		1	
B	*Dendryphantes acutus* Wesołowska & Haddad, 2014	2	DD	STHE			1							
B	*Dendryphantes hararensis* Wesołowska & Cumming, 2008	2	LC	STHE			1	1	1	1	1		1	
F	*Dendryphantes limpopo* Wesołowska & Haddad, 2013	3	DDT	SAE					1	1				
B	*Dendryphantes matumi* Haddad & Wesołowska, 2013	5	DD	SAE	KZNE				1					
M	*Dendryphantes neethlingi* Haddad & Wesołowska, 2013	6	DDT	SAE	KZNE				1					
B	*Dendryphantes purcelli* Peckham & Peckham, 1903	1	LC	AE		1	1		1					1
B	*Dendryphantes rafalskii* Wesołowska, 2000	2	LC	STHE			1	1						
B	*Dendryphantes schultzei* Simon, 1910	2	DD	STHE								1		
B	*Dendryphantes silvestris* Wesołowska & Haddad, 2013	3	LC	SAE		1		1	1					
B	*Euophrys bifida* Wesołowska, Azarkina & Russell-Smith, 2014	5	VU	SAE	ECE	1								
?	*Euophrys capicola* Simon, 1901	6	DDT	SAE	WCE?									1
B	*Euophrys cochlea* Wesołowska, Azarkina & Russell-Smith, 2014	5	LC	SAE	WCE									1
B	*Euophrys elizabethae* Wesołowska, Azarkina & Russell-Smith, 2014	6	DD	SAE	WCE									1
B	*Euophrys falciger* Wesołowska, Azarkina & Russell-Smith, 2014	5	LC	SAE	KZNE				1					
B	*Euophrys gracilis* Wesołowska, Azarkina & Russell-Smith, 2014	2	LC	STHE					1					
B	*Euophrys leipoldti* Peckham & Peckham, 1903	4	LC	SAE								1		1
B	*Euophrys limpopo* Wesołowska, Azarkina & Russell-Smith, 2014	4	DD	SAE						1	1			
F	*Euophrys menemerella* Strand, 1909	6	DDT	SAE	WCE									1
B	*Euophrys meridionalis* Wesołowska, Azarkina & Russell-Smith, 2014	4	LC	SAE			1		1					
F	*Euophrys miranda* Wesołowska, Azarkina & Russell-Smith 2014	6	DDT	SAE	ECE	1								
M	*Euophrys nana* Wesołowska, Azarkina & Russell-Smith, 2014	6	DDT	SAE	WCE									1
B	*Euophrys purcelli* Peckham & Peckham, 1903	4	LC	SAE		1								1
M	*Euophrys recta* Wesołowska, Azarkina & Russell-Smith, 2014	5	DDT	SAE	WCE									1
B	*Euophrys subtilis* Wesołowska, Azarkina & Russell-Smith, 2014	5	DD	SAE	KZNE				1					
M	*Euophrys uphami* (Peckham & Peckham, 1903)	6	DDT	SAE	WCE									1
F	*Evarcha alba* (Peckham & Peckham, 1903)	2	LC	STHE			1		1					
M	*Evarcha acuta* Wesołowska, 2006	2	LC	STHE			1							
M	*Evarcha amanzi* Wesołowska & Haddad, 2018	6	DDT	SAE	FSE		1							
F	*Evarcha annae* (Peckham & Peckham, 1903)	3	LC	SAE		1			1					1
M	*Evarcha brinki* Haddad & Wesołowska, 2011	5	DDT	SAE	NCE							1		
B	*Evarcha denticulata* Haddad & Wesołowska, 2013	4	LC	SAE		1	1							1
B	*Evarcha flagellaris* Haddad & Wesołowska, 2011	1	LC	AE		1	1	1		1			1	
B	*Evarcha ignea* Wesołowska & Cumming, 2008	1	LC	AE				1	1	1	1			
M	*Evarcha karas* Wesołowska, 2011	1	LC	STHE								1		1
M	*Evarcha mirabilis* Wesołowska & Haddad, 2009	3	LC	SAE					1					1
B	*Evarcha prosimilis* Wesołowska & Cumming, 2008	1	LC	AE			1	1	1	1	1	1	1	
B	*Evarcha striolata* Wesołowska & Haddad, 2009	3	LC	SAE				1	1	1	1			1
M	*Evarcha villosa* Wesołowska & Haddad, 2018	6	DDT	SAE	NCE							1		
M	*Evarcha vittula* Haddad & Wesołowska, 2011	3	LC	SAE			1	1	1	1				
B	*Evarcha werneri* (Simon, 1906)	1	LC	AE					1	1				
M	*Evarcha zimbabwensis* Wesołowska & Cumming, 2008	2	LC	STHE		1			1					

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Festucula haddadi* Azarkina & Foord, 2014	4	LC	SAE					1		1			
B	*Festucula leroyae* Azarkina & Foord, 2014	2	LC	STHE			1	1		1	1		1	
B	*Festucula robustus* Azarkina & Foord, 2014	3	DD	SAE				1	1					
B	*Goleba puella* (Simon, 1885)	1	LC	AE		1			1	1	1			
B	*Habrocestum africanum* Wesołowska & Haddad, 2009	3	LC	SAE					1					1
M	*Habrocestum albimanum* Simon, 1901	5	LC	SAE	WCE									1
M	*Habrocestum auricomum* Haddad & Wesołowska, 2013	5	DDT	SAE	LE					1				
B	*Habrocestum flavimanum* Simon, 1901	6	DD	SAE	WCE?									1
F	*Habrocestum laurae* Peckham & Peckham, 1903	4	DDT	SAE		1			1					
M	*Habrocestum luculentum* Peckham & Peckham, 1903	4	DDT	SAE								1		1
M	*Habrocestum sapiens* (Peckham & Peckham, 1903)	2	LC	STHE										1
B	*Habrocestum superbum* Wesołowska, 2000	2	LC	STHE						1				
B	*Habrocestum tanzanicum* Wesołowska & Russell-Smith, 2000	1	LC	AE									1	
B	*Harmochirus luculentus* Simon, 1885	1	LC	AE		1	1	1	1	1			1	1
B	*Hasarinella distincta* Haddad & Wesołowska, 2013	4	LC	SAE						1			1	
B	*Hasarius adansoni* (Audouin, 1826)	0	LC	C		1	1	1	1	1		1	1	
B	*Heliophanus aberdarensis* Wesołowska, 1986	1	LC	AE		1	1	1	1		1			
F	*Heliophanus africanus* Wesołowska, 1986	5	EN	SAE	GE			1						
M	*Heliophanus bellus* Wesołowska, 1986	6	DDT	SAE	WCE									1
F	*Heliophanus berlandi* Lawrence, 1937	6	DDT	SAE	KZNE				1					
B	*Heliophanus bisulcus* Wesołowska, 1986	2	LC	STHE										1
F	*Heliophanus capensis* Wesołowska, 1986	4	LC	SAE								1		1
B	*Heliophanus capicola* Simon, 1901	2	LC	STHE										1
B	*Heliophanus charlesi* Wesołowska, 2003	3	LC	SAE			1	1	1	1	1	1	1	1
B	*Heliophanus claviger* Simon, 1901	4	LC	SAE					1					1
M	*Heliophanus deamatus* Peckham & Peckham, 1903	2	LC	STHE				1		1			1	
B	*Heliophanus debilis* Simon, 1901	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Heliophanus demonstrativus* Wesołowska, 1986	1	LC	AE		1	1	1	1	1	1			
B	*Heliophanus deserticola* Simon, 1901	3	LC	SAE	WCE	1					1	1		
F	*Heliophanus designatus* Peckham & Peckham, 1903	6	DDT	SAE		1								
B	*Heliophanus fascinatus* Wesołowska, 1986	1	LC	AE		1	1	1	1					
B	*Heliophanus gramineus* Wesołowska & Haddad, 2013	3	LC	SAE		1			1	1	1			
B	*Heliophanus hamifer* Simon, 1886	1	LC	AE						1				
B	*Heliophanus hastatus* Wesołowska, 1986	1	LC	STHE		1	1	1	1	1	1			
F	*Heliophanus horrifer* Wesołowska, 1986	5	DDT	SAE	WCE									1
B	*Heliophanus infaustus* (Peckham & Peckham, 1903)	1	LC	AE		1		1						
B	*Heliophanus insperatus* Wesołowska, 1986	1	LC	AE				1		1	1			1
B	*Heliophanus lesserti* Wesołowska, 1986	1	LC	AE					1	1	1			1
M	*Heliophanus marshalli* Peckham & Peckham, 1903	6	DDT	SAE	KZNE				1					
M	*Heliophanus mirabilis* Wesołowska, 1986	5	DDT	SAE	WCE									1
B	*Heliophanus modicus* Peckham & Peckham, 1903	1	LC	AE		1	1							1
B	*Heliophanus nanus* Wesołowska, 2003	2	LC	STHE		1	1	1		1	1	1	1	
B	*Heliophanus ndumoensis* Haddad & Wesołowska, 2013	3	LC	SAE					1	1				
B	*Heliophanus orchesta* Simon, 1885	1	LC	AE		1		1	1	1	1		1	
B	*Heliophanus patellaris* Simon, 1901	2	LC	STHE		1	1	1	1		1	1		1
B	*Heliophanus pauper* Wesołowska, 1986	1	LC	AE		1			1		1		1	
B	*Heliophanus peckhami* Simon, 1902	5	LC	SAE	WCE									1
B	*Heliophanus pistaciae* Wesołowska, 2003	2	LC	STHE			1	1	1	1	1	1	1	
M	*Heliophanus portentosus* Wesołowska, 1986	6	DDT	SAE	WCE									1
M	*Heliophanus pratti* Peckham & Peckham, 1903	2	LC	STHE		1					1			1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Heliophanus proszynskii* Wesołowska, 2003	2	LC	STHE		1	1	1	1	1	1	1		1
B	*Heliophanus pygmaeus* Wesołowska & Russell-Smith, 2000	1	LC	AE					1	1	1			
F	*Heliophanus redimitus* Simon, 1910	6	DDT	SAE	NCE							1		
B	*Heliophanus sororius* Wesołowska, 2003	2	LC	STHE		1	1				1			
B	*Heliophanus termitophagus* Wesołowska & Haddad, 2002	4	LC	SAE			1					1		
M	*Heliophanus thaleri* Wesołowska, 2009	6	DDT	SAE	FSE		1							
B	*Heliophanus transvaalicus* Simon, 1901	3	LC	SAE			1	1	1	1		1	1	
B	*Heliophanus trepidus* Simon, 1910	2	LC	STHE			1	1	1	1	1	1	1	1
B	*Hispo georgius* (Peckham & Peckham, 1892)	1	LC	AE		1	1	1	1	1	1			
B	*Holcolaetis vellerae* Simon, 1910	1	LC	AE				1		1				
B	*Holcolaetis zuluensis* Lawrence, 1937	1	LC	AE			1	1	1	1	1			
F	*Homalattus coriaceus* Simon, 1902	1	LC	AE		1								
F	*Homalattus obscurus* Peckham & Peckham, 1903	6	DDT	SAE	WCE									1
F	*Homalattus punctatus* Peckham & Peckham, 1903	5	EN	SAE	KZNE				1					
B	*Hyllus argyrotoxus* Simon, 1902	1	LC	AE		1		1	1	1	1		1	1
B	*Hyllus brevitarsis* Simon, 1902	1	LC	AE		1		1	1	1	1			
B	*Hyllus dotatus* (Peckham & Peckham, 1903)	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Hyllus flavescens* Simon, 1902	6	DDT	SAE	KZNE?				1					
B	*Hyllus treleaveni* Peckham & Peckham, 1903	1	LC	AE		1	1	1	1	1	1		1	1
F	*Icius dendryphantoides* Strand, 1909	6	DDT	SAE	WCE									1
F	*Icius desertorum* Simon, 1901	6	DDT	SAE	WCE									1
B	*Icius insolidus* (Wesołowska, 1999)	2	LC	STHE		1	1	1	1	1	1	1	1	
B	*Icius nigricaudus* Wesołowska & Haddad, 2009	5	EN	SAE	KZNE				1					
M	*Icius pulchellus* Haddad & Wesołowska, 2011	4	DDT	SAE			1					1		
M	*Iranattus principalis* Wesołowska, 2000	1	LC	AE						1				
M	*Kima africana* Peckham & Peckham, 1902	2	LC	STHEb		1						1		1
B	*Kima atra* Peckham & Peckham, 1903	1	LC	AE				1						
M	*Kima variabilis* Peckham & Peckham, 1903	3	LC	SAE		1			1		1			1
F	*Langelurillus cedarbergensis* Haddad & Wesołowska, 2013	6	DDT	SAE	WCE									1
F	*Langelurillus krugeri* Wesołowska & Haddad, 2013	6	DDT	SAE	LE					1				
B	*Langelurillus minutus* Wesołowska & Cumming, 2011	2	LC	STHE						1				
F	*Langelurillus namibicus* Wesołowska, 2011	2	LC	STHE								1		
B	*Langellurillus squamiger* Wesołowska & Haddad, 2018	6	DD	SAE	KZNE				1					
B	*Langona bethae* Wesołowska & Cumming, 2011	2	LC	STHE						1				
F	*Langona bisecta* Lawrence, 1927	2	LC	STHE						1				1
B	*Langona hirsuta* Haddad & Wesołowska, 2011	3	LC	SAE			1			1	1	1		1
B	*Langona lotzi* Haddad & Wesołowska, 2011	2	LC	STHE			1		1					
M	*Langona manicata* Simon, 1901	6	DDT	SAE	LE					1				
B	*Langona tortuosa* Wesołowska, 2011	2	LC	STHE						1	1			
B	*Langona warchalowskii* Wesołowska , 2007	2	LC	STHE		1	1	1	1	1		1		1
B	*Manzuna botswana* Azarkina, 2019	2	LC	STHE						1				
B	*Manzuna petroae* Azarkina, 2019	3	LC	SAE		1		1	1	1				
F	*Massagris contortuplicata* Wesołowska & Haddad, 2013	5	DDT	SAE		1								
B	*Massagris honesta* Wesołowska, 1993	4	LC	SAE		1								1
B	*Massagris maculosa* Wesołowska & Haddad, 2018	6	DD	SAE	WCE									1
B	*Massagris mirifica* Peckham & Peckham, 1903	3	LC	SAE		1			1					1
B	*Massagris natalensis* Wesołowska & Haddad, 2009	5	VU	SAE	KZNE				1					
M	*Massagris schisma* Maddison & Zhang, 2006	6	DDT	SAE	NCE							1		
M	*Massagris separata* Wesołowska, 1993	6	DDT	SAE	ECE	1								
B	*Meleon kenti* (Lessert, 1925)	1	LC	AE					1					

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Menemerus bifurcus* Wesołowska, 1999	1	LC	AE		1		1		1	1		1	
B	*Menemerus bivittatus* (Dufour, 1831)	0	LC	C		1		1	1					1
F	*Menemerus carlini* (Peckham & Peckham, 1903)	2	LC	STHE		1								
B	*Menemerus congoensis* Lessert, 1927	1	LC	AE				1						
B	*Menemerus eburnensis* Berland & Millot, 1941	1	LC	AE				1		1				
B	*Menemerus fagei* Berland & Millot, 1941	1	LC	AE			1		1	1				
M	*Menemerus lesnei* Lessert, 1936	2	LC	STHE					1					
F	*Menemerus lesserti* Lawrence, 1927	2	LC	STHE						1		1		
M	*Menemerus meridionalis* Wesołowska, 1999	6	DDT	SAE	LE					1				
B	*Menemerus minshullae* Wesołowska, 1999	1	LC	AE					1	1				
M	*Menemerus natalis* Wesołowska, 1999	3	LC	SAE					1	1				
M	*Menemerus pilosus* Wesołowska, 1999	2	LC	STHE								1		
B	*Menemerus rubicundus* Lawrence, 1928	2	LC	STHE			1					1		
B	*Menemerus transvaalicus* Wesołowska, 1999	2	LC	STHE		1	1	1		1	1	1		1
B	*Menemerus zimbabwensis* Wesołowska, 1999	2	LC	STHE					1	1	1		1	
B	*Mexcala elegans* Peckham & Peckham, 1903	1	LC	AE		1		1	1	1	1	1		1
F	*Mexcala meridiana* Wesołowska, 2009	4	DDT	SAE						1	1			
F	*Mexcala quadrimaculata* (Lawrence, 1942)	2	LC	STHE					1	1				
M	*Mexcala rufa* Peckham & Peckham, 1902	2	LC	STHE						1		1		1
B	*Microbianor furcatus* Haddad & Wesołowska, 2013	4	LC	SAE		1	1							
M	*Microbianor globosus* Haddad & Wesołowska, 2011	6	DDT	SAE	NCE							1		
M	*Microbianor simplex* Wesołowska & Haddad, 2018	6	DDT	SAE	WCE									1
B	*Modunda staintoni* (O.P.-Cambridge, 1872)	0	LC	C					1					
M	*Mogrus albogularis* Simon, 1901	3	LC	SAE				1	1	1		1	1	
B	*Mogrus mathisi* (Berland & Millot, 1941)	0	LC	C		1	1	1	1	1			1	
M	*Myrmarachne albosetosa* Wanless, 1978	6	DDT	SAE	WCE									1
B	*Myrmarachne foreli* Lessert, 1925	1	LC	AE		1			1	1				
B	*Myrmarachne ichneumon* (Simon, 1886)	1	LC	AE			1		1	1	1			
M	*Myrmarachne inflatipalpis* Wanless, 1978	1	LC	AE		1								
B	*Myrmarachne laurentina* Bacelar, 1953	2	LC	STHE		1			1	1				1
M	*Myrmarachne leleupi* Wanless, 1978	3	LC	SAE				1	1		1			1
M	*Myrmarachne lesserti* Lawrence, 1938	4	LC	SAE		1			1					
B	*Myrmarachne lulengana* Roewer, 1965	1	LC	AE					1	1				
B	*Myrmarachne marshalli* Peckham & Peckham, 1903	1	LC	AE		1		1	1	1	1			
F	*Myrmarachne natalica* Lessert, 1925	5	DDT	SAE	KZNE				1					
B	*Myrmarachne solitaria* Peckham & Peckham, 1903	2	LC	STHE		1	1	1	1	1			1	1
B	*Myrmarachne uvira* Wanless, 1978	1	LC	AE			1	1		1			1	1
B	*Natta chionogaster* (Simon, 1901)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Natta horizontalis* Karsch, 1879	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Neaetha irreperta* Wesołowska & Russell-Smith, 2000	1	LC	AE					1	1				
B	*Nigorella hirsuta* Wesołowska, 2009	2	LC	STHE		1	1	1	1	1	1		1	
B	*Oviballus vidae* Azarkina & Haddad, 2020	3	LC	SAE		1	1		1					
B	*Pachyballus castaneus* Simon, 1900	2	LC	STHE					1		1			
B	*Pachyballus flavipes* Simon, 1910	1	LC	AE					1	1				
B	*Pachyballus miniscutulus* Wesołowska, Azarkina & Wiśniewski, 2020	4	LCC	SAE			1		1					
B	*Pachyballus transversus* Simon, 1900	1	LC	AE						1	1			
M	*Pachypoessa lacertosa* Simon, 1902	1	LC	AE					1					
M	*Parajotus obscurofemoratus* Peckham & Peckham, 1903	3	LC	SAE					1	1	1			
B	*Parajotus refulgens* Wesołowska, 2000	1	LC	AE						1	1			
B	*Pellenes beani* Peckham & Peckham, 1903	3	DD	SAE		1					1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Pellenes bulawayoensis* Wesołowska, 2000	2	LC	STHE			1	1	1	1	1	1	1	
B	*Pellenes cingulatus* Wesołowska & Russell-Smith, 2000	1	LC	AE					1					
B	*Pellenes epularis* (O.P.-Cambridge, 1872)	0	LC	C			1		1			1	1	1
B	*Pellenes geniculatus* (Simon, 1868)	0	LC	C		1	1	1	1	1		1	1	1
B	*Pellenes modicus* Wesołowska & Russell-Smith, 2000	1	LC	AE			1	1	1	1		1	1	
B	*Pellenes pulcher* Logunov, 1995	0	LC	C					1	1				
M	*Pellenes rufoclypeatus* Peckham & Peckham, 1903	6	DDT	SAE	KZNE				1					
B	*Pellenes tharinae* Wesołowska, 2006	2	LC	STHE		1	1	1	1	1	1	1	1	1
B	*Peplometus chlorophthalmus* Simon, 1900	1	LC	AE					1					
B	*Phintella aequipes* (Peckham & Peckham, 1903)	1	LC	AE		1			1	1	1			1
B	*Phintella australis* (Simon, 1902)	3	LC	SAE			1			1				1
F	*Phintella lajuma* Haddad & Wesołowska, 2013	3	LC	SAE					1	1				
M	*Phlegra albostriata* Simon, 1901	2	LC	STHE						1		1		1
M	*Phlegra arborea* Wesołowska & Haddad, 2009	6	DDT	SAE	KZNE				1					
F	*Phlegra bairstowi* Simon, 1885	6	DDT	SAE	ECE	1								
B	*Phlegra bresnieri* (Lucas, 1846)	0	LC	C		1	1		1			1		1
M	*Phlegra certa* Wesołowska & Haddad, 2009	1	LC	AE				1	1	1	1			
B	*Phlegra etosha* Logunov & Azarkina, 2006	2	LC	STHE			1			1		1		
F	*Phlegra imperiosa* Peckham & Peckham, 1903	3	LC	SAE		1		1	1	1	1			1
B	*Phlegra karoo* Wesołowska, 2006	2	LC	STHE		1	1	1	1	1	1	1	1	
B	*Phlegra nuda* Próchniewicz & Heciak, 1994	1	LC	AE		1		1	1					
B	*Phlegra pusilla* Wesołowska & van Harten, 1994	1	LC	AE				1		1				
M	*Phlegra simplex* Wesołowska & Russell-Smith, 2000	1	LC	AE						1	1			
F	*Phlegra varia* Wesołowska & Russell-Smith, 2000	1	LC	AE						1				
M	*Pignus pongola* Wesołowska & Haddad, 2009	2	LC	STHE					1					1
B	*Pignus simoni* (Peckham & Peckham, 1903)	2	LC	STHE		1	1	1	1	1	1	1	1	
B	*Planamarengo bimaculata* (Peckham & Peckham, 1903)	3	LC	SAE		1		1	1	1	1		1	
B	*Plexippus petersi* (Karsch, 1878)	0	LC	C							1			
M	*Plexippus rubrogularis* Simon, 1902	6	DDT	SAE	LE					1				
F	*Plexippus tsholotsho* Wesołowska, 2011	2	LC	STHE					1	1				
B	*Portia schultzi* Karsch, 1878	1	LC	AE		1		1	1	1	1			1
M	*Propiomarengo foordi* Azarkina & Haddad, 2020	6	DDT	SAE	FSE					1				
F	*Propiomarengo plana* (Haddad & Wesołowska, 2013)	5	DDT	SAE			1							
B	*Psenuc dependens* (Haddad & Wesołowska, 2011)	3	LC	SAE			1			1		1		1
F	*Psenuc solitarius* (Haddad & Wesołowska, 2011)	2	LC	STHE			1							
F	*Pseudicius adustus* Wesołowska, 2006	2	LC	STHE								1		
B	*Pseudicius africanus* Peckham & Peckham, 1903	2	LC	STHE		1						1		
B	*Pseudicius dentatus* Wesołowska & Haddad, 2013	3	LC	SAE			1		1		1			
F	*Pseudicius femineus* Wesołowska & Haddad, 2013	4	DDT	SAE		1								1
M	*Pseudicius flabellus* Wesołowska & Haddad, 2013	5	DDT	SAE	WCE									1
B	*Pseudicius gracilis* Haddad & Wesołowska, 2011	3	LC	SAE			1	1	1				1	
B	*Pseudicius maculatus* Haddad & Wesołowska, 2011	2	LC	STHE		1	1		1					1
M	*Pseudicius marshi* (Peckham & Peckham, 1903)	4	LC	SAE								1		1
B	*Pseudicius matabelensis* Wesołowska, 2011	2	LC	STHE					1			1		
F	*Pseudicius musculus* Simon, 1901	1	LC	AE					1			1		
B	*Pseudicius procerus* Wesołowska & Haddad, 2018	6	DD	SAE	NCE									
B	*Pseudicius squamatus* Haddad & Wesołowska, 2013	6	DD	SAE	KZNE				1					
B	*Pseudicius zebra* Simon, 1902	6	DD	SAE	ECE	1								
B	*Rhene amanzi* Wesołowska & Haddad, 2011	6	DD	SAE	FSE		1							
M	*Rhene banksi* Peckham & Peckham, 1902	4	DDT	SAE			1							

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
M	*Rhene biguttata* Peckham & Peckham, 1903	4	DD	SAE		1			1					1
F	*Rhene capensis* Strand, 1909	5	DDT	SAE	WCE									
F	*Rhene cooperi* Lessert, 1925	5	DDT	SAE	KZNE				1					
B	*Rhene facilis* Wesołowska & Russell-Smith, 2000	1	LC	AE	FSE?				1					
J	*Rhene foai* Simon, 1902	6	DDT	SAE	?		?							
B	*Rhene konradi* Wesołowska, 2009	5	LC	SAE			1							
M	*Rhene legitima* Wesołowska & Haddad, 2018	6	DDT	SAE	ECE	1								
M	*Rhene lingularis* Haddad & Wesołowska, 2011	3	LC	SAE		1	1					1		
B	*Rhene machadoi* Berland & Millot, 1941	1	LC	AE		1				1	1			
B	*Rhene pinguis* Wesołowska & Haddad, 2009	5	DD	SAE	KZNE				1					
M	*Rhene punctatus* Wesołowska & Haddad, 2013	5	DDT	SAE	KZNE				1					
F	*Rhene timidus* Wesołowska & Haddad, 2013	4	LC	SAE		1			1					
M	*Rumburak bellus* Wesołowska, Azarkina & Russell-Smith, 2014	5	DDT	SAE	WCE									1
B	*Rumburak hilaris* Wesołowska, Azarkina & Russell-Smith, 2014	4	LC	SAE		1								1
B	*Rumburak lateripunctatus* Wesołowska, Azarkina & Russell-Smith, 2014	5	Rare	SAE	WCE									1
B	*Rumburak laxus* (Zhang & Maddison, 2012)	3	LC	SAE				1		1	1			
B	*Rumburak mirabilis* Wesołowska, Azarkina & Russell-Smith, 2014	5	Rare	SAE	ECE	1								
M	*Rumburak tuberatus* Wesołowska, Azarkina & Russell-Smith, 2014	5	DDT	SAE	LE					1				
B	*Rumburak virilis* Wesołowska, Azarkina & Russell-Smith, 2014	5	DD	SAE	ME						1			
M	*Salticus annulatus* (Giebel, 1870)	6	DDT	SAE	KZNE?				1					
B	*Schenkelia modesta* Lessert, 1927	1	LC	AE					1	1	1			
M	*Sibianor kenyaensis* Logunov, 2001	1	LC	AE					1					
B	*Sibianor victoriae* Logunov, 2001	1	LC	AE		1					1			1
B	*Sonoita lightfooti* Peckham & Peckham, 1903	1	LC	AE				1	1	1				
B	*Stenaelurillus guttiger* (Simon, 1901)	2	LC	STHE			1	1	1	1	1		1	
B	*Stenaelurillus modestus* Wesołowska, 2014	5	DD	SAE	KZNE				1					
B	*Stenaelurillus termitophagus* (Wesołowska & Cumming, 1999)	2	LC	STHE			1	1		1				
M	*Tanzania meridionalis* Haddad & Wesołowska, 2011	6	DDT	SAE	FSE									
M	*Tanzania minutus* (Wesołowska & Russell-Smith, 2000)	1	LC	AE										
B	*Tanzania mkomaziensis* (Wesołowska & Russell-Smith, 2000)	1	LC	AE										
B	*Tanzania parvulus* Wesołowska, Azarkina, Russell-Smith, 2014	3	LC	SAE										1
B	*Tanzania striatus* Wesołowska, Azarkina, Russell-Smith, 2014	5	Rare	SAE	WCE									
M	*Tenuiballus coronatus* Azarkina & Haddad, 2020	6		SAE	KZNE				1					
M	*Tenuiballus minor* Azarkina & Haddad, 2020	6		SAE	KZNE				1					
M	*Thyene aperta* (Peckham & Peckham, 1903)	1	LC	AE		1	1	1	1			1		
B	*Thyene australis* Peckham & Peckham, 1903	1	LC	AE		1				1				
F	*Thyene bilineata* Lawrence, 1927	2	LC	STHE				1		1		1		
M	*Thyene bucculenta* (Gerstäcker, 1873)	1	LC	AE		1		1	1	1		1		1
B	*Thyene coccineovittata* (Simon, 1885)	1	LC	AE		1	1		1	1	1			1
M	*Thyene coronata* Simon, 1902	6	DDT	SAE	KZNE?				1					
F	*Thyene dakarensis* (Berland & Millot, 1941)	1	LC	AE						1	1			
B	*Thyene imperialis* (Rossi, 1846)	1	LC	AE					1	1		1		
B	*Thyene inflata* (Gerstäcker, 1873)	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Thyene leighi* Peckham & Peckham, 1903	1	LC	AE		1			1	1				
B	*Thyene natalii* Peckham & Peckham, 1903	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Thyene ogdeni* Peckham & Peckham, 1903	1	LC	AE		1		1	1	1	1			1
B	*Thyene semiargentea* (Simon, 1884)	1	LC	AE		1	1	1	1	1	1		1	
B	*Thyene thyenioides* (Lessert, 1925)	1	LC	AE		1	1	1		1	1	1	1	
B	*Thyenula alotama* Wesołowska, Azarkina & Russell-Smith, 2014	3	LC	SAE		1			1		1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Thyenula armata* Wesołowska, 2001	2	LC	STHE		1	1		1					
B	*Thyenula aurantiaca* (Simon, 1902)	2	LC	STHE		1	1	1	1	1	1	1	1	1
M	*Thyenula cheliceroides* Wesołowska, Azarkina, Russell-Smith, 2014	3	LC	SAE					1					1
B	*Thyenula clarisognata* Wesołowska, Azarkina, Russell-Smith, 2014	4	EN	SAE		1			1					
M	*Thyenula dentatidens* Wesołowska, Azarkina, Russell-Smith, 2014	5	DDT	SAE	KZNE				1					
B	*Thyenula fidelis* Wesołowska & Haddad, 2009	2	LC	STHE		1			1	1				
B	*Thyenula haddadi* Wesołowska, Azarkina & Russell-Smith, 2014	5	LC	SAE	KZNE				1					
B	*Thyenula juvenca* Simon, 1902	3	LC	SAE		1			1	1				
B	*Thyenula leighi* (Peckham & Peckham, 1903)	4	LC	SAE		1			1					
M	*Thyenula magna* Wesołowska & Haddad, 2009	4	LC	SAE		1			1					
M	*Thyenula natalica* (Simon, 1902)	3	LC	SAE		1		1	1		1			
B	*Thyenula oranjensis* Wesołowska, 2001	3	LC	SAE			1		1	1	1			1
B	*Thyenula rufa* Wesołowska, Azarkina & Russell-Smith, 2014	4	EN	SAE		1			1					
B	*Thyenula sempiterna* Wesołowska, 2000	2	LC	STHE					1	1				
B	*Thyenula splendens* Wesołowska & Haddad, 2018	6	DD	SAE	ECE	1								
F	*Thyenula tenebrica* Wesołowska, Azarkina, Russell-Smith, 2014	6	DDT	SAE	ECE	1								
M	*Thyenula virgulata* Wesołowska, Azarkina, Russell-Smith, 2014	5	LC	SAE	KZNE				1					
B	*Thyenula vulnifica* Wesołowska, Azarkina, Russell-Smith, 2014	5	DD	SAE	ECE	1								
B	*Thyenula wesolowskae* Zhang & Maddison, 2012	4	LC	SAE						1	1			
B	*Tomomingi szutsi* Wesołowska & Haddad, 2013	4	DD	SAE						1	1			
B	*Tusitala barbata* Peckham & Peckham, 1902	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Tusitala hirsuta* Peckham & Peckham, 1902	1	LC	AE		1			1	1	1			
B	*Tusitala lyrata* (Simon, 1903)	1	LC	AE					1					1
B	*Ureta quadrispinosa* (Lawrence, 1938)	4	LC	SAE		1			1					
B	*Veissella durbani* (Peckham & Peckham, 1903)	3	LC	SAE		1			1	1	1			
F	*Viciria flavipes* Peckham & Peckham, 1903	4	DDT	SAE		1			1					
M	*Wandawe australis* Azarkina & Haddad, 2020	4	DDT	SAE		1			1					
B	*Wandawe benjamini* (Wesołowska & Haddad, 2013)	4	LC	SAE		1			1					
B	*Xuriella prima* Wesołowska & Russell-Smith, 2000	1	LC	AE		1			1					1
M	*Yimbulunga foordi* Wesołowska, Azarkina & Russell-Smith, 2014	6	DDT	SAE	KZNE				1					
F	*Zulunigma incognita* (Wesołowska & Haddad, 2009)	6	DDT	SAE	KZNE				1					
FAMILY SCYTODIDAE Blackwall, 1864														
B	*Scytodes arenacea* Purcell, 1904	2	LC	STHE				1				1		
F	*Scytodes broomi* Pocock, 1902	4	DDT	SAE			1					1		
B	*Scytodes caffra* Purcell, 1904	3	LC	SAE				1	1	1	1			1
F	*Scytodes cedri* Purcell 1904	4	LC	SAE		1								1
F	*Scytodes clavata* Benoit, 1965	1	LC	AE						1	1			
J	*Scytodes constellata* Lawrence, 1938	3	LC	SAE		1			1	1	1			
F	*Scytodes drakensbergensis* Lawrence, 1947	4	LC	SAE			1		1					
M	*Scytodes elizabethae* Purcell, 1904	3	LC	SAE		1	1	1			1			1
B	*Scytodes flagellata* Purcell, 1904	3	LC	SAE				1	1		1			1
B	*Scytodes fusca* Walckenaer, 1837	0	LC	C		1		1	1	1				1
B	*Scytodes gooldi* Purcell, 1904	5	VU	SAE	WCE									1
F	*Scytodes karrooica* Purcell, 1904	6	DDT	SAE	WCE									1
M	*Scytodes lanceolata* Purcell, 1904	6	DDT	SAE	NCE							1		
B	*Scytodes lawrencei* Lessert, 1939	1	LC	AE		1				1				
F	*Scytodes leipoldti* Purcell, 1904	4	DDT	SAE								1		1
F	*Scytodes lycosella* Purcell, 1904	6	DDT	SAE	KZNE				1					
B	*Scytodes lyriformis* Purcell, 1904	6	DD	SAE	NCE							1		
F	*Scytodes maritima* Lawrence, 1938	3	LC	SAE					1	1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Scytodes marshalli* Pocock, 1902	6	DDT	SAE	KZNE				1					
F	*Scytodes montana* Purcell, 1904	5	Rare	SAE	WCE									1
J	*Scytodes quinqua* Lawrence, 1927	2	LC	STHE						1				
B	*Scytodes rubra* Lawrence, 1937	4	LC	SAE		1			1					
F	*Scytodes schultzei* Purcell 1908	5	DDT	SAE	NCE							1		
B	*Scytodes silvatica* Purcell, 1904	4	LC	SAE		1								1
B	*Scytodes subulata* Purcell, 1904	6	DD	SAE	WCE									1
F	*Scytodes symmetrica* Lawrence, 1938	6	DDT	SAE	KZNE				1					
B	*Scytodes testudo* Purcell, 1904	5	LC	SAE	WCE									1
B	*Scytodes thoracica* (Latreille, 1802)	0	LC	C				1	1	1				
F	*Scytodes triangulifera* Purcell, 1904	4	DDT	SAE		1								1
F	*Scytodes trifoliata* Lawrence, 1938	3	LC	SAE		1			1					
FAMILY SEGESTRIIDAE Simon, 1893														
F	*Ariadna bilineata* Purcell, 1904	3	LC	SAE				1		1			1	1
F	*Ariadna capensis* Purcell, 1904	5	DDT	SAE	WCE									1
B	*Ariadna corticola* Lawrence, 1952	3	LC	SAE		1	1	1	1	1	1			
F	*Ariadna dentigera* Purcell, 1904	5	DDT	SAE	WCE									1
F	*Ariadna gryllotalpa* (Purcell, 1904)	6	DDT	SAE	WCE									1
F	*Ariadna hottentotta* Purcell, 1908	3	LC	SAE		1						1		1
F	*Ariadna insularis* Purcell, 1908	2	LC	STHE								1		
F	*Ariadna jubata* Purcell, 1904	5	LC	SAE	NCE							1		
B	*Ariadna karrooica* Purcell, 1904	3	LC	SAE		1	1					1		
F	*Ariadna kolbei* Purcell, 1904	6	DDT	SAE	ECE	1								
F	*Ariadna lightfooti* Purcell, 1904	3	LC	SAE		1						1		1
F	*Ariadna scabripes* Purcell,1904	6	DDT	SAE	NCE							1		
F	*Ariadna segestrioides* Purcell, 1904	6	DDT	SAE	ECE	1								
F	*Ariadna similis* Purcell, 1908	6	DDT	SAE	NWE								1	
F	*Ariadna umtalica* Purcell, 1904	2	LC	STHE								1		
FAMILY SELENOPIDAE Simon, 1897														
M	*Anyphops alticola* (Lawrence, 1940)	4	DDT	SAE					1		1			
B	*Anyphops amatolae* (Lawrence, 1940)	6	DD	SAE	ECE	1								
B	*Anyphops atomarius* (Simon, 1887)	4	LC	SAE		1								1
F	*Anyphops barbertonensis* (Lawrence, 1940)	1	LC	AE			1	1	1	1	1			1
F	*Anyphops barnardi* (Lawrence, 1940)	2	LC	STHE				1				1		
F	*Anyphops basutus* (Pocock, 1901)	2	LC	STHE		1			1					
B	*Anyphops bechuanicus* (Lawrence, 1940)	3	DD	SAE					1				1	
F	*Anyphops braunsi* (Lawrence, 1940)	3	LC	SAE		1					1			
B	*Anyphops broomi* (Pocock, 1900)	3	LC	SAE			1					1		1
F	*Anyphops caledonicus* (Lawrence, 1940)	5	DDT	SAE	WCE									1
B	*Anyphops capensis* (Lawrence, 1940)	4	LC	SAE		1								1
F	*Anyphops civicus* (Lawrence, 1940)	3	LC	SAE		1	1		1					
B	*Anyphops decoratus* (Lawrence, 1940)	3	LC	SAE		1			1	1	1			
F	*Anyphops dubiosus* (Lawrence, 1952)	5	DDT	SAE	KZNE				1					
M	*Anyphops fitzsimonsi* (Lawrence, 1940)	3	LC	SAE				1			1		1	
M	*Anyphops gilli* (Lawrence, 1940)	3	LC	SAE		1			1					1
F	*Anyphops helenae* (Lawrence, 1940)	6	DDT	SAE	WCE									1
F	*Anyphops hessei* (Lawrence, 1940)	3	LC	SAE			1					1		1
F	*Anyphops hewitti* (Lawrence, 1940)	2	LC	STHE		1				1				
M	*Anyphops immaculatus* (Lawrence, 1940)	3	LC	SAE		1	1	1						1
B	*Anyphops karrooicus* (Lawrence, 1940)	3	LC	SAE		1	1					1		

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Anyphops kraussi* (Pocock, 1898)	5	Rare	SAE	WCE									1
B	*Anyphops lawrencei* (Roewer, 1951)	2	LC	STHE			1	1	1					
F	*Anyphops leleupi* Benoit, 1972	4	LC	SAE						1	1	1		
M	*Anyphops lesserti* (Lawrence, 1940)	5	DDT	SAE	WCE									1
M	*Anyphops lignicola* (Lawrence, 1937)	4	LC	SAE					1		1			
F	*Anyphops lochiel* Corronca, 2000	4	DDT	SAE						1	1			
F	*Anyphops longipedatus* (Roewer, 1955)	3	DDT	SAE				1	1				1	
F	*Anyphops lucia* Corronca, 2005	6	DDT	SAE	KZNE				1					
F	*Anyphops lycosiformis* (Lawrence, 1937)	3	LC	SAE					1	1				
F	*Anyphops maculosus* (Lawrence, 1940)	3	LC	SAE		1						1		1
M	*Anyphops marshalli* (Pocock, 1902)	3	LC	SAE		1			1					1
B	*Anyphops minor* (Lawrence, 1940)	3	LC	SAE					1					1
F	*Anyphops montanus* (Lawrence, 1940)	5	DDT	SAE	WCE									1
M	*Anyphops mumai* (Corronca, 1996)	5	DDT	SAE	ECE	1								
B	*Anyphops namaquensis* (Lawrence, 1940)	4	LC	SAE								1		1
F	*Anyphops narcissi* Benoit, 1972	2	LC	STHE					1		1			
B	*Anyphops natalensis* (Lawrence, 1940)	6	DD	SAE	KZNE				1					
B	*Anyphops ngome* Corronca, 2005	3	LC	SAE					1	1				
F	*Anyphops parvulus* (Pocock, 1900)	4	DDT	SAE		1								1
B	*Anyphops phallus* (Lawrence, 1952)	6	DD	SAE	KZNE				1					
M	*Anyphops pococki* (Lawrence, 1940)	3	LC	SAE					1		1		1	
F	*Anyphops purcelli* (Lawrence, 1940)	4	DDT	SAE		1								1
F	*Anyphops regalis* (Lawrence, 1940)	5	DDT	SAE	WCE									1
F	*Anyphops reservatus* (Lawrence, 1937)	3	LC	SAE					1	1				
F	*Anyphops rubicundus* (Lawrence, 1940)	3	LC	SAE							1			1
F	*Anyphops schoenlandi* (Pocock, 1902)	5	LC	SAE	ECE	1								
B	*Anyphops septemspinatus* (Lawrence, 1937)	2	LC	STHE					1	1				
F	*Anyphops sexspinatus* (Lawrence, 1940)	5	LC	SAE	NCE							1		
B	*Anyphops silvicolellus* (Strand, 1913)	1	LC	AE						1				
F	*Anyphops spenceri* (Pocock, 1896)	3	LC	SAE		1			1	1				
B	*Anyphops stauntoni* (Pocock, 1902)	1	LC	AE		1		1	1				1	
M	*Anyphops stridulans* (Lawrence, 1940)	6	DDT	SAE	NCE							1		
F	*Anyphops thornei* (Lawrence, 1940)	5	DDT	SAE	WCE									1
F	*Anyphops transvaalicus* (Lawrence, 1940)	5	LC	SAE	ME						1			
F	*Anyphops tuckeri* (Lawrence, 1940)	3	LC	SAE		1	1	1		1	1		1	
F	*Anyphops tugelanus* (Lawrence, 1942)	6	DDT	SAE	KZNE				1					
F	*Anyphops whiteae* (Pocock, 1902)	4	LC	SAE		1			1					
F	*Selenops ansieae* Corronca, 2002	5	DDT	SAE	LE					1				
F	*Selenops brachycephalus* Lawrence, 1940	2	LC	STHE						1	1			
F	*Selenops dilon* Corronca, 2002	4	DDT	SAE						1	1			
F	*Selenops feron* Corronca, 2002	2	LC	STHE				1						
F	*Selenops ilcuria* Corronca, 2002	1	LC	AE						1	1			
B	*Selenops intricatus* Simon, 1910	1	LC	AE					1					
F	*Selenops kruegeri* Lawrence, 1940	1	LC	AE				1		1	1	1		
B	*Selenops lesnei* Lessert, 1936	1	LC	AE						1	1			
F	*Selenops ovambicus* Lawrence, 1940	1	LC	AE						1				
B	*Selenops radiatus* Latreille, 1819	0	LC	C		1	1	1	1	1	1	1	1	
F	*Selenops tenebrosus* Lawrence, 1940	2	LC	STHE						1				
F	*Selenops tonteldoos* Corronca, 2005	6	DDT	SAE	ME						1			
B	*Selenops zuluanus* Lawrence, 1940	2	LC	STHE					1	1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY SICARIIDAE Keyserling, 1880														
B	*Hexophthalma albospinosa* (Purcell, 1908)	2	LC	STHE								1		
B	*Hexophthalma hahni* (Karsch, 1878)	2	LC	STHE						1	1	1	1	1
F	*Hexophthalma leroyi* Lotz, 2018	6	DDT	SAE	NCE							1		
B	*Hexophthalma spatulatus* (Pocock, 1900)	4	LC	SAE		1								1
F	*Loxosceles cederbergensis* Lotz, 2017	5	DDT	SAE	WCE									1
B	*Loxosceles dejagerae* Lotz, 2017	3	LC	SAE		1						1		1
M	*Loxosceles haddadi* Lotz, 2017	6	DDT	SAE	LE					1				
F	*Loxosceles makapanensis* Lotz, 2017	5	DDT	SAE	LE					1				
B	*Loxosceles parramae* Newlands, 1981	4	LC	SAE			1	1						
B	*Loxosceles pilosa* Purcell, 1908	2	LC	STHE								1		
B	*Loxosceles rufescens* (Dufour, 1820)	0	LC	C										1
B	*Loxosceles simillima* Lawrence, 1927	1	LC	AE			1	1	1	1	1	1	1	
B	*Loxosceles speluncarum* Simon, 1893	5	VU	SAE	GE			1						
B	*Loxosceles spinulosa* Purcell, 1904	4	LC	SAE		1								1
FAMILY SPARASSIDAE Bertkau, 1872														
B	*Arandisa deserticola* Lawrence, 1938	2	LC	STHE								1		
F	*Eusparassus borakalalo* Moradmand, 2013	4	LC	SAE				1		1				
B	*Eusparassus jaegeri* Moradmand, 2013	2	LC	STHE				1	1	1	1	1	1	
B	*Eusparassus schoemanae* Moradmand, 2013	2	LC	STHE		1	1					1		
B	*May bruno* Jäger & Krehenwinkel, 2015	5	LC	SAE	NCE							1		
B	*Olios auricomis* (Simon, 1880)	1	LC	AE				1	1	1	1			
B	*Olios biarmatus* Lessert, 1925	4	LC	SAE		1			1					
M	*Olios brachycephalus* Lawrence, 1938	4	LC	SAE		1			1					
M	*Olios chelifer* Lawrence, 1937	3	LC	SAE					1	1	1			
M	*Olios chubbi* Lessert, 1923	2	LC	STHE					1	1				
B	*Olios correvoni nigrifrons* Lawrence, 1928	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Olios fasciculatus* Simon, 1880	5	LC	SAE	KZNE				1					
F	*Olios fonticola* (Pocock, 1902)	6	DDT	SAE	ECE	1								
M	*Olios freyi* Lessert, 1929	1	LC	AE						1				
B	*Olios kruegeri* (Simon, 1897)	6	DD	SAE	GE			1						
B	*Olios kunzi* Jager, 2020	1	LC	AE			1							
B	*Olios lacticolor* Lawrence, 1952	4	DD	SAE		1			1					
B	*Olios machadoi* Lawrence, 1952	3	LC	SAE		1			1		1			1
F	*Olios marshalli* (Pocock, 1898)	6	DDT	SAE	KZNE				1					
M	*Olios sherwoodi* Lessert, 1929	1	LC	AE								1		
B	*Olios sjostedti* Lessert, 1921	1	LC	AE				1	1	1	1			
M	*Olios stictopus* (Pocock, 1898)	6	DDT	SAE	KZNE				1					
F	*Olios zulu* Simon, 1880	6	DDT	SAE	KZNE				1					
F	*Palystella namaquensis* Lawrence, 1938	6	DDT	SAE	NCE							1		
B	*Palystella pallida* Lawrence, 1938	4	LC	SAE								1		1
B	*Palystes ansiedippenaarae* Croeser, 1996	4	LC	SAE					1		1			
B	*Palystes castaneus* (Latreille, 1819)	2	LC	STHE										1
B	*Palystes crawshayi* Pocock, 1902	2	LC	STHE			1							
B	*Palystes karooensis* Croeser, 1996	3	LC	SAE		1			1					1
B	*Palystes kreuzmanni* Jäger & Kunz, 2010	5	EN	SAE	WCE									1
B	*Palystes leppanae* Pocock, 1902	4	LC	SAE		1								1
M	*Palystes leroyorum* Croeser, 1996	3	LC	SAE	EC?		1	1		1				
F	*Palystes lunatus* Pocock, 1896	6	DDT	SAE	?	1								
B	*Palystes martinfilmeri* Croeser, 1996	5	Rare	SAE	WCE									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Palystes perornatus* Pocock, 1900	3	LC	SAE		1			1					1
B	*Palystes stilleri* Croeser, 1996	4	LC	SAE								1		1
B	*Palystes stuarti* Croeser, 1996	4	DD	SAE								1		1
B	*Palystes superciliosus* L. Koch, 1875	2	LC	STHE		1	1	1	1	1	1	1	1	1
F	*Panaretella distincta* (Pocock, 1896)	4	NT	SAE		1			1					
B	*Panaretella immaculata* Lawrence, 1952	3	LC	SAE					1	1	1			
B	*Panaretella minor* Lawrence, 1952	4	LC	SAE		1			1					
F	*Panaretella scutata* (Pocock, 1902)	4	DDT	SAE		1			1					
B	*Panaretella zuluana* Lawrence, 1937	4	LC	SAE					1		1			
B	*Parapalystes cultrifer* (Pocock, 1900)	3	LC	SAE								1		1
B	*Parapalystes euphorbiae* Croeser, 1996	5	LC	SAE	NCE							1		
B	*Parapalystes lycosinus* (Pocock, 1900)	4	LC	SAE		1								1
B	*Parapalystes megacephalus* (C.L. Koch, 1845)	6	DD	SAE	WCE									1
M	*Parapalystes whiteae* (Pocock, 1902)	6	DDT	SAE	ECE	1								
B	*Pseudomicrommata longipes* (Bösenberg & Lenz, 1895)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Pseudomicrommata vittigera* (Simon, 1897)	2	LC	STHE						1	1	1		
FAMILY STASIMOPIDAE Bond, Opatova & Hedin, 2020														
F	*Stasimopus artifex* Pocock, 1902	5	DDT	SAE	ECE	1								
B	*Stasimopus astutus* Pocock, 1902	5	DD	SAE	ECE	1								
F	*Stasimopus bimaculatus* Purcell, 1903	3	LC	SAE		1								1
B	*Stasimopus brevipalpis* Purcell, 1903	5	DD	SAE	WCE									1
F	*Stasimopus caffrus* (C.L. Koch, 1842)	6	DDT	SAE	WCE									1
F	*Stasimopus castaneus* Purcell, 1903	6	DDT	SAE	ECE	1								
F	*Stasimopus coronatus* Hewitt, 1915	3	DDT	SAE			1	1		1			1	
F	*Stasimopus dreyeri* Hewitt, 1915	6	DDT	SAE	FSE		1							
B	*Stasimopus erythrognathus* Purcell, 1903	6	DD	SAE	WCE									1
B	*Stasimopus filmeri* Engelbrecht & Prendini, 2012	4	EN	SAE				1					1	
B	*Stasimopus gigas* Hewitt, 1915	4	DD	SAE			1						1	
B	*Stasimopus griswoldi* Engelbrecht & Prendini, 2012	5	EN	SAE	NWE								1	
B	*Stasimopus hewitti* Engelbrecht & Prendini, 2012	5	VU	SAE	GE			1						
B	*Stasimopus insculptus* Pocock, 1901	5	DD	SAE	ECE	1								
B	*Stasimopus kentanicus* Purcell, 1903	6	DD	SAE	ECE	1								
F	*Stasimopus kolbei* Purcell, 1903	6	DDT	SAE	ECE	1								
F	*Stasimopus leipoldti* Purcell, 1902	6	DDT	SAE	WCE									1
M	*Stasimopus longipalpis* Hewitt, 1917	4	DDT	SAE	NCE		1					1		
B	*Stasimopus mandelai* Hendrixson & Bond, 2004	6	CR	SAE	ECE	1								
B	*Stasimopus maraisi* Hewitt, 1914	4	DD	SAE								1		1
F	*Stasimopus meyeri* (Karsch, 1879)	6	DDT	SAE	NCE							1		
M	*Stasimopus minor* Hewitt, 1915	5	DDT	SAE	FSE		1							
F	*Stasimopus nanus* Tucker, 1917	6	DDT	SAE	FSE		1							
B	*Stasimopus nigellus* Pocock, 1902	4	DD	SAE			1							
F	*Stasimopus obscurus* Purcell, 1908	4	DDT	SAE			1					1		
F	*Stasimopus oculatus* Pocock, 1897	3	LC	SAE			1	1		1	1	1	1	
M	*Stasimopus palpiger* Pocock, 1902	6	DDT	SAE	ECE	1								
B	*Stasimopus patersonae* Hewitt, 1913	5	DD	SAE	ECE	1								
F	*Stasimopus poweri* Hewitt, 1915	6	DDT	SAE	NCE							1		
M	*Stasimopus purcelli* Tucker, 1917	6	DDT	SAE	WCE									1
F	*Stasimopus quadratimaculatus* Purcell, 1903	6	DDT	SAE	WCE									1
B	*Stasimopus qumbu* Hewitt, 1913	6	DD	SAE	ECE	1								
B	*Stasimopus robertsi* Hewitt, 1910	5	LC	SAE	GE			1						

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Stasimopus rufidens* (Ausserer, 1871)	5	DDT	SAE	KZNE				1					
B	*Stasimopus schoenlandi* Pocock, 1900	5	DD	SAE	ECE	1								
B	*Stasimopus schreineri* Purcell, 1903	3	DDT	SAE		1						1		
F	*Stasimopus schultzei* Purcell, 1908	5	DDT	SAE	NCE							1		
B	*Stasimopus spinipes* Hewitt, 1917	6	DD	SAE	ECE	1								
B	*Stasimopus spinosus* Hewitt, 1914	5	DD	SAE	ECE	1								
M	*Stasimopus steynburgensis* Hewitt, 1915	6	DDT	SAE	ECE	1								
F	*Stasimopus suffuscus* Hewitt, 1916	6	DDT	SAE	GE			1						
B	*Stasimopus tysoni* Hewitt, 1919	6	DD	SAE	ECE	1								
B	*Stasimopus unispinosus* Purcell, 1903	5	DD	SAE	NCE							1		
F	*Stasimopus umtaticus* Purcell, 1903	5	DDT	SAE	ECE	1								
FAMILY SYMPHYTOGNATHIDAE Hickman, 1931														
F	*Symphytognatha imbulunga* Griswold, 1987	6	DDT	SAE	KZNE				1					
B	*Cangoderces globosa* Wang, Li & Haddad, 2018	6	DD	SAE	ME						1			
FAMILY TELEMIDAE Fage, 1913														
B	*Cangoderces lewisi* Harington, 1951	6	CR	SAE	WCE									1
FAMILY TETRAGNATHIDAE Menge, 1866														
B	*Diphya foordi* Omelko, Marusik & Lyle, 2020	3	LC	SAE		1		1		1	1			1
M	*Diphya leroyorum* Omelko, Marusik & Lyle, 2020	6	DDT	SAE	ME						1			
B	*Diphya simoni* Kauri, 1950	3	LC	SAE		1				1				1
F	*Diphya vanderwaltae* Omelko, Marusik & Lyle, 2020	6	DDT	SAE	ECE	1								
B	*Diphya wesolowskae* Omelko, Marusik & Lyle, 2020	3	RARE	SAE		1			1					
B	*Glenognatha argyrostilba* (O.P.-Cambridge, 1876)	0	LC	C					1	1				
B	*Leucauge argyrescens* Benoit, 1978	1	LC	AE		1			1	1				
F	*Leucauge auronotum* Strand, 1907	3	LC	SAE				1	1	1	1			
B	*Leucauge decorata* (Blackwall, 1864)	0	LC	C		1	1	1	1	1	1	1		1
B	*Leucauge festiva* (Blackwall, 1866)	1	LC	AE		1	1	1	1	1	1	1	1	1
J	*Leucauge fishoekensis* Strand, 1909	6	DDT	SAE	WCE									1
B	*Leucauge kibonotensis* Tullgren, 1910	1	LC	AE						1				1
B	*Leucauge levanderi* (Kulczynski, 1901)	1	LC	AE		1		1	1	1	1			
F	*Leucauge medjensis* Lessert, 1930	1	LC	AE		1			1	1	1			
M	*Leucauge thomeensis* Kraus, 1960	1	LC	AE		1			1	1	1			1
F	*Meta meruensis* Tullgren, 1910	1	LC	AE	WCE				1	1	1			
B	*Metellina haddadi* Marusik & Larsen, 2018	5	DD	SAE										1
B	*Pachygnatha leleupi* Lawrence, 1952	1	LC	AE						1				
B	*Pachygnatha zappa* Bosmans & Bosselaers, 1994	1	LC	AE					1	1				
B	*Tetragnatha bogotensis* Keyserling, 1865	0	LC	C		1	1	1	1	1	1	1	1	1
J	*Tetragnatha caffra* (Strand, 1909)	6	DDT	SAE	WCE									1
B	*Tetragnatha ceylonica* O.P.-Cambridge, 1869	0	LC	C		1	1		1					1
B	*Tetragnatha demissa* L. Koch, 1872	0	LC	C		1	1	1	1	1	1			1
B	*Tetragnatha isidis* (Simon, 1880)	0	LC	C				1	1	1				1
B	*Tetragnatha jaculator* Tullgren, 1910	0	LC	C			1	1	1	1				
B	*Tetragnatha keyserlingi* Simon, 1890	0	LC	C		1			1	1	1			1
B	*Tetragnatha nitens* (Audouin, 1826)	0	LC	C		1	1	1	1	1				1
B	*Tetragnatha subsquamata* Okuma, 1985	1	LC	AE		1		1	1	1	1		1	
F	*Tetragnatha taylori* O.P.-Cambridge, 1890	6	DDT	SAE	?									
B	*Tetragnatha unicornis* Tullgren, 1910	1	LC	AE		1			1					
B	*Tetragnatha vermiformis* Emerton, 1884	0	LC	C		1			1		1			1
FAMILY THERAPHOSIDAE Thorell, 1869														
B	*Augacephalus breyeri* (Hewitt, 1919)	2	LC	STHE					1	1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Augacephalus junodi* (Simon, 1904)	2	LC	STHE		1		1	1	1	1			
F	*Brachionopus annulatus* Purcell, 1903	6	DDT	SAE	ECE	1								
F	*Brachionopus pretoriae* Purcell, 1904	3	DDT	SAE				1		1	1			
B	*Brachionopus robustus* Pocock, 1897	3	DDT	SAE					1	1	1			
F	*Brachionopus tristis* Purcell, 1903	3	DDT	SAE					1	1	1			
B	*Ceratogyrus brachycephalus* Hewitt, 1919	4	DDT	SAE						1	1			
B	*Ceratogyrus darlingi* Pocock, 1897	2	LC	STHE					1	1	1	1	1	
B	*Ceratogyrus paulseni* Gallon, 2005	5	VUD2	SAE	LE					1				
B	*Harpactira atra* (Latreille, 1832)	5	LC	SAE	WCE									1
B	*Harpactira baviana* Purcell, 1903	4	LC	SAE		1						1		
B	*Harpactira cafreriana* (Walckenaer, 1837)	5	LC	SAE	WCE									1
M	*Harpactira chrysogaster* Pocock, 1897	5	LC	SAE	WCE									1
B	*Harpactira curator* Pocock, 1898	5	LC	SAE	KZN				1					
F	*Harpactira curvipes* Pocock, 1897	5	LC	SAE	ECE	1								
B	*Harpactira dictator* Purcell, 1902	5	LC	SAE	WCE									1
F	*Harpactira gigas* Pocock, 1898	3	LC	SAE					1	1	1			
B	*Harpactira hamiltoni* Pocock, 1902	3	LC	SAE			1	1	1	1	1		1	
F	*Harpactira lineata* Pocock, 1897	6	DDT	SAE	?									
F	*Harpactira lyrata* (Simon, 1892)	6	DDT	SAE	?									
B	*Harpactira marksi* Purcell, 1902	5	LC	SAE	WCE									1
B	*Harpactira namaquensis* Purcell, 1902	4	LC	SAE								1		1
F	*Harpactira pulchripes* Pocock, 1901	5	LC	SAE	ECE	1								
B	*Harpactira tigrina* Ausserer, 1875	4	LC	SAE		1			1					
F	*Harpactirella domicola* Purcell, 1903	6	DDT	SAE	WCE									1
B	*Harpactirella helenae* Purcell, 1903	5	DDT	SAE	WCE									1
B	*Harpactirella karrooica* Purcell, 1902	5	DDT	SAE	WCE									1
B	*Harpactirella lapidaria* Purcell, 1908	5	DDT	SAE	NCE							1		
F	*Harpactirella lightfooti* Purcell, 1902	5	DDT	SAE	WCE									1
B	*Harpactirella longipes* Purcell, 1902	5	DDT	SAE	WCE									1
F	*Harpactirella magna* Purcell, 1903	5	DDT	SAE	ECE	1								
B	*Harpactirella overdijki* Gallon, 2010	3	DDT	SAE						1	1		1	
F	*Harpactirella schwarzi* Purcell, 1904	6	DDT	SAE	ECE	1								
B	*Harpactirella spinosa* Purcell, 1908	5	DDT	SAE	NCE							1		
B	*Harpactirella treleaveni* Purcell, 1902	5	DDT	SAE	WCE									1
B	*Idiothele mira* Gallon, 2010	5	Rare	SAE	KZNE				1					
B	*Idiothele nigrofulva* (Pocock, 1898)	2	LC	STHE		1	1		1	1	1	1	1	1
B	*Pterinochilus lapalala* Gallon & Engelbrecht, 2011	5	Rare	SAE	LE					1				
B	*Pterinochilus lugardi* Pocock, 1900	1	LC	AE						1				
B	*Trichognathella schoenlandi* (Pocock, 1900)	2	LC	AE		1								
FAMILY THERIDIIDAE Sundevall, 1833														
B	*Achaearanea globispira* Henschel & Jocqué, 1994	4	DD	SAE								1		1
B	*Anelosimus nelsoni* Agnarsson, 2006	3	LC	SAE		1	1	1	1	1	1		1	1
B	*Argyrodes argyrodes* (Walckenaer, 1841)	0	LC	C		1		1	1					
B	*Argyrodes convivans* Lawrence, 1937	2	LC	STHE		1			1	1	1	1		1
B	*Argyrodes sextuberculosus* Strand, 1908	1	LC	AE					1	1				
M	*Argyrodes stridulator* Lawrence, 1937	5	LC	SAE	KZNE				1					
B	*Argyrodes zonatus* (Walckenaer, 1841)	1	LC	AE		1		1		1	1			1
F	*Araimnes campestratus* Simon, 1903	1	LC	AE					1	1				
F	*Chorizopella tragardhi* Lawrence, 1947	3	LC	SAE		1		1	1	1	1		1	
F	*Enoplognatha inornata* O.P.-Cambridge, 1904	3	LC	SAE		1		1		1		1		1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Enoplognatha molesta* O.P.-Cambridge, 1904	3	LC	SAE		1	1	1	1	1	1	1	1	1
J	*Episinus bilineatus* Simon, 1894	2	LC	STHE		1	1	1	1	1	1	1	1	1
M	*Episinus bishopi* (Lessert, 1929)	1	LC	AE		1	1		1					1
F	*Episinus marignaci* (Lessert, 1933)	2	LC	STHE		1			1	1				
B	*Euryopis episinoides* (Walckenaer, 1847)	0	LC	C		1	1	1	1	1	1	1		1
B	*Euryopis funebris* (Hentz, 1850)	0	LC	C		1		1	1	1	1	1		1
B	*Histagonia deserticola* Simon, 1895	2	LC	STHE								1		
B	*Latrodectus cinctus* Blackwall, 1865	0	LC	C		1			1	1				1
B	*Latrodectus geometricus* C.L. Koch, 1841	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Latrodectus indistinctus* O.P.-Cambridge, 1904	2	LC	STHE		1						1		1
B	*Latrodectus karrooensis* Smithers, 1944	4	LC	SAE								1		1
B	*Latrodectus renivulvatus* Dahl, 1902	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Latrodectus rhodesiensis* Mackay, 1972	2	LC	STHE			1	1		1				
B	*Latrodectus umbukwane* M.Wright, C.Wright, Lyle & Engelbrecht, 2019	5	DDT	SAE	KZNE				1					
B	*Meotipa pulcherrima* (Mello-Leitão, 1917)	0	LC	C		1			1					
B	*Parasteatoda lunata* (Clerck, 1757)	0	LC	C					1					
B	*Parasteatoda tepidariorum* (C.L. Koch, 1841)	0	LC	C		1		1						1
B	*Phoroncidia capensis* (Simon, 1895)	6	DD	SAE	WCE									1
F	*Phoroncidia eburnea* (Simon, 1895)	3	LC	SAE		1	1	1	1	1	1	1		
M	*Phoroncidia truncatula* (Strand, 1909)	6	DDT	SAE	WCE									1
B	*Phycosoma martinae* (Roberts, 1983)	0	LC	C		1	1	1	1	1	1		1	1
B	*Phycosoma spundana* (Roberts, 1978)	1	LC	AE						1	1			
B	*Platnickina mneon* (Bösenberg & Strand, 1906)	0	LC	C		1			1	1	1		1	
F	*Rhomphaea affinis* Lessert, 1936	2	LC	STHE			1		1					
B	*Rhomphaea nasica* (Simon, 1873)	0	LC	C		1		1	1	1				1
B	*Steatoda capensis* Hann, 1990	0	LC	C		1	1	1	1	1	1	1	1	1
F	*Steatoda connexa* (O.P.-Cambridge, 1904)	6	DDT	SAE	WCE?									1
B	*Steatoda erigoniformis* (O.P.-Cambridge, 1872)	0	LC	C		1	1	1	1	1	1	1		1
F	*Steatoda fagei* (Lawrence, 1964)	5	DDT	SAE	WCE									1
B	*Steatoda foravae* Dippenaar-Schoeman & Müller, 1992	4	LC	SAE		1								1
B	*Steatoda grossa* (C.L. Koch, 1838)	1	LC	C				1		1	1	1		1
B	*Steatoda lawrencei* Brignoli, 1983	5	DDT	SAE	KZNE				1					
F	*Steatoda mormorata* (Simon, 1910)	5	LC	SAE	NCE							1		
B	*Steatoda triangulosa* (Walckenaer, 1802)	3	LC	AE					1	1				
J	*Theridion albidorsum* Strand, 1909	6	DDT	SAE	WCE									1
F	*Theridion auberti* Simon, 1904	6	DDT	SAE	LE					1				
B	*Theridion bradyanum* Strand, 1907	6	DD	SAE	WCE									1
B	*Theridion dedux* O.P.-Cambridge, 1904	6	DD	SAE	KZNE				1					
F	*Theridion delicatum* O.P.-Cambridge, 1904	4	LC	SAE		1								1
F	*Theridion durbanicum* Lawrence, 1947	6	DDT	SAE	KZNE				1					
J	*Theridion octoferum* Strand, 1909	6	DDT	SAE	WCE									1
B	*Theridion pictum* (Walckenaer, 1802)	0	LC	C				1	1	1	1			
F	*Theridion proximum* Lawrence, 1964	6	DDT	SAE	WCE									1
B	*Theridion purcelli* O.P.-Cambridge, 1904	3	LC	SAE		1	1	1	1	1	1	1	1	1
B	*Theridion retreatense* Strand, 1909	6	DD	SAE	WCE									1
F	*Theridion vanhoeffeni* Strand, 1909	6	DDT	SAE	WCE									1
B	*Tidarren cuneolatum* (Tullgren, 1910)	1	LC	AE			1	1	1	1	1	1		1
F	*Tidarren scenicum* (Thorell, 1899)	1	LC	AE		1		1	1	1	1			
F	*Tidarren ubickorum* Knoflach & Van Harten, 2006	2	DDT	STHE							1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
FAMILY THOMISIDAE Sundevall, 1833														
B	*Ansiae tuckeri* (Lessert, 1919)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Avelis hystriculus* Simon, 1895	3	LC	SAE		1				1			1	1
F	*Borboropactus australis* (Lawrence, 1937)	5	DDT	SAE	KZN				1					
B	*Borboropactus silvicola* (Lawrence, 1938)	3	LC	SAE		1			1	1	1			
F	*Borboropactus squalidus* Simon, 1884	2	LC	STHE		1			1					
B	*Camaricus nigrotesselatus* Simon, 1895	1	LC	AE		1	1	1	1	1	1		1	1
M	*Cynathea bicolor* Simon, 1895	1	LC	AE		1			1					
B	*Diaea puncta* Karsch, 1884	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Diaea rohani* Fage, 1923	2	LC	STHE		1	1	1	1					
F	*Diaea viridipes* Strand, 1909	4	DDT	SAE		1								1
M	*Firmicus abnormis* (Lessert, 1923)	3	LC	SAE		1	1					1		1
M	*Firmicus bipunctatus* Caporiacco, 1941	1	LC	AE		1	1	1						
B	*Firmicus bragantinus* (Brito Capello, 1866)	1	LC	AE					1	1	1			1
B	*Geraesta congoensis* (Lessert, 1943)	1	LC	AE		1			1	1	1			
B	*Heriaeus allenjonesi* Van Niekerk & Dippenaar-Schoeman, 2013	3	LC	SAE			1					1		1
B	*Heriaeus copricola* Van Niekerk & Dippenaar-Schoeman, 2013	2	LC	STHE				1	1	1	1			
B	*Heriaeus crassispinus* Lawrence, 1942	1	LC	AE		1		1	1	1	1	1	1	
B	*Heriaeus foordi* Van Niekerk & Dippenaar-Schoeman, 2013	3	LC	SAE					1	1	1			
M	*Heriaeus muizenberg* Van Niekerk & Dippenaar-Schoeman, 2013	6	CR	SAE	WCE									1
B	*Heriaeus peterwebbi* Van Niekerk & Dippenaar-Schoeman, 2013	2	LC	STHE			1	1		1	1		1	
F	*Heriaeus sossusvlei* Van Niekerk & Dippenaar-Schoeman, 2013	2	LC	STHE										1
B	*Heriaeus transvaalicus* Simon, 1895	4	LC	SAE						1			1	
B	*Heriaeus xanderi* Van Niekerk & Dippenaar-Schoeman, 2013	1	LC	AE					1		1			
B	*Heriaeus zanii* Van Niekerk & Dippenaar-Schoeman, 2013	3	LC	SAE			1					1		1
F	*Hewittia gracilis* Lessert, 1928	1	LC	AE		1	1	1	1	1	1		1	1
B	*Holopelus albibarbis* Simon, 1895	1	LC	AE		1						1		1
B	*Holopelus almiae* Dippenaar-Schoeman, 1986	4	LC	SAE		1								1
B	*Misumenops rubrodecoratus* Millot, 1942	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Monaeses austrinus* Simon, 1910	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Monaeses fuscus* Dippenaar-Schoeman, 1984	3	LC	SAE					1	1	1			1
B	*Monaeses gibbus* Dippenaar-Schoeman, 1984	3	LC	SAE		1	1		1	1	1			
B	*Monaeses griseus* Pavesi, 1897	1	LC	AE		1			1	1	1			
B	*Monaeses paradoxus* Lucas, 1864	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Monaeses pustulosus* Pavesi, 1895	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Monaeses quadrituberculatus* Lawrence, 1927	1	LC	AE		1	1	1	1	1	1	1	1	
B	*Mystaria flavogutatta* (Lawrence, 1952)	1	LC	AE		1			1	1				
B	*Mystaria irmatrix* Lewis & Dippenaar-Schoeman, 2014	2	LC	STHE		1			1					
B	*Mystaria lata* (Lawrence, 1927)	2	LC	STHE		1			1	1				
B	*Mystaria lindaicapensis* Lewis & Dippenaar-Schoeman, 2014	4	VU	SAE		1								1
B	*Mystaria mnyama* Lewis & Dippenaar-Schoeman, 2014	5	DD	SAE	KZNE				1					
B	*Mystaria occidentalis* (Millot, 1942)	1	LC	AE		1			1					
B	*Mystaria rufolimbata* Simon, 1895	1	LC	AE					1	1			1	
B	*Mystaria savannensis* Lewis & Dippenaar-Schoeman, 2014	1	LC	AE				1	1	1			1	
B	*Mystaria variabilis* (Lessert, 1919)	1	LC	AE		1			1					
F	*Nyctimus trimeni* (Simon 1895)	6	DDT	SAE	?									
B	*Oxytate argenteooculata* (Simon, 1886)	1	LC	AE		1	1	1	1	1	1			1
F	*Oxytate concolor* (Caporiacco, 1947)	1	LC	AE		1		1	1	1	1			1
B	*Oxytate leruthi* (Lessert, 1943)	1	LC	AE				1			1			1
B	*Oxytate ribes* (Jézéquel, 1964)	1	LC	AE		1		1	1	1				1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
B	*Ozyptila caenosa* Jézéquel, 1966	1	LC	AE		1			1			1		1
B	*Pactactes compactus* Lawrence, 1947	3	LC	SAE		1			1	1	1			
B	*Pactactes obesus* Simon, 1895	1	LC	AE		1			1					1
M	*Pactactes trimaculatus* Simon, 1895	1	LC	AE		1			1	1	1			1
B	*Parabomis elsae* Dippenaar-Schoeman & Foord, 2020	3	LC	SAE				1	1	1	1			
B	*Parabomis martini* Lessert, 1919	1	LC	AE		1		1	1	1	1		1	
B	*Parabomis megae* Dippenaar-Schoeman & Foord, 2020	2	LC	STHE						1				
B	*Parabomis pilosus* Dippenaar-Schoeman & Foord, 2020	2	LC	STHE						1				
B	*Parasmodix quadrituberculata* Jézéquel, 1966	1	LC	AE				1	1	1	1			1
M	*Phaenopoma nigropunctatum* (O.P.-Cambridge, 1883)	3	LC	SAE		1			1	1				1
B	*Pherecydes carinae* Dippenaar-Schoeman, 1980	4	LC	SAE					1		1			
F	*Pherecydes ionae* Dippenaar-Schoeman, 1980	1	LC	AE		1			1	1				
B	*Pherecydes lucinae* Dippenaar-Schoeman, 1980	3	LC	SAE		1			1	1			1	
B	*Pherecydes nicolaasi* Dippenaar-Schoeman, 1980	3	LC	SAE		1			1	1				
B	*Pherecydes tuberculatus* O.P.-Cambridge, 1883	2	LC	STHE		1	1		1	1		1		1
B	*Pherecydes zebra* Lawrence, 1927	1	LC	AE					1	1	1			
B	*Phrynarachne melloleitaoi* Lessert, 1933	2	LC	STHE		1			1		1	1		1
B	*Phrynarachne rugosa* (Latreille, 1804)	1	LC	AE		1			1					
F	*Platythomisus deserticola* Lawrence, 1936	2	LC	STHE							1			
F	*Platythomisus jubbi* Lawrence, 1968	3	LC	SAE		1			1	1	1			
F	*Platythomisus sibayius* Lawrence, 1968	6	DDT	SAE	KZNE				1					
B	*Runcinia aethiops* (Simon, 1901)	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Runcinia depressa* Simon, 1906	1	LC	AE			1	1	1	1	1	1	1	
B	*Runcinia erythrina* Jézéquel, 1964	1	LC	AE		1	1	1	1	1	1		1	1
B	*Runcinia flavida* (Simon, 1881)	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Runcinia grammica* (L. Koch, 1937)	0	LC	C		1	1		1	1	1			1
B	*Runcinia insecta* (L. Koch, 1875)	0	LC	C		1	1		1	1	1		1	1
B	*Runcinia johnstoni* Lessert, 1919	1	LC	AE		1			1	1	1	1	1	1
B	*Runcinia tropica* Simon, 1907	1	LC	AE				1	1		1			
B	*Simorcus capensis* Simon, 1895	1	LC	AE		1						1		1
B	*Simorcus cotti* Lessert, 1936	1	LC	AE		1		1	1	1	1		1	
B	*Simorcus haddadi* Van Niekerk & Dippenaar-Schoeman, 2010	5	NT	SAE	WCE									1
B	*Simorcus lotzi* Van Niekerk & Dippenaar-Schoeman, 2010	2	LC	STHE			1			1				
B	*Smodicinus coroniger* Simon, 1895	1	LC	AE		1			1	1				
F	*Stiphropella gracilis* Lawrence, 1952	3	LC	SAE					1		1		1	
B	*Stiphropus affinis* Lessert, 1923	2	LC	STHE		1	1	1	1	1	1	1		
M	*Stiphropus bisigillatus* Lawrence, 1952	2	LC	STHE		1	1		1	1	1		1	
M	*Stiphropus drassiformis* (O.P.-Cambridge, 1883)	6	DDT	SAE	ECE?	1								
B	*Stiphropus intermedius* Millot, 1942	1	LC	AE				1	1				1	
B	*Sylligma ndumi* Honiball & Dippenaar-Schoeman 2011	2	LC	STHE		1			1	1	1		1	
B	*Synema decens* (Karsch, 1878)	2	LC	STHE		1		1	1	1	1	1	1	1
B	*Synema diana* (Audouin, 1826)	1	LC	AE				1	1	1	1			
B	*Synema imitatrix* (Pavesi, 1883)	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Synema langheldi* Dahl, 1907	1	LC	AE		1		1	1	1	1			1
B	*Synema mandibulare* Dahl, 1907	2	LC	AE		1			1	1	1			
B	*Synema marlothi* Dahl, 1907	2	LC	STHE		1		1	1		1			1
M	*Synema nigrotibiale* Lessert, 1919	1	LC	AE		1		1	1	1	1		1	1
M	*Synema riflense* Strand, 1909	3	LC	SAE		1					1	1		1
F	*Synema simoneae* Lessert, 1919	1	LC	AE		1		1	1	1		1		1
F	*Synema vallotoni* Lessert, 1923	2	LC	STHE		1		1	1	1	1			

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Tagulis granulosus* Simon, 1895	1	LC	AE					1					
J	*Talaus limbatus* Simon, 1895	6	DDT	SAE	LE					1				
B	*Thomisops bullatus* Simon, 1895	2	LC	STHE		1		1	1	1	1			
B	*Thomisops granulatus* Dippenaar-Schoeman, 1989	2	LC	STHE		1			1					
B	*Thomisops lesserti* Millot, 1942	1	LC	AE		1		1	1					
B	*Thomisops melanopes* Dippenaar-Schoeman, 1989	3	LC	SAE		1			1		1		1	1
B	*Thomisops pupa* Karsch, 1879	1	LC	AE		1		1	1	1	1			
B	*Thomisops senegalensis* Millot, 1942	1	LC	AE		1			1		1	1		1
B	*Thomisops sulcatus* Simon, 1895	1	LC	AE		1	1	1	1	1	1			1
B	*Thomisus australis* Comellini, 1957	1	LC	AE		1	1	1	1	1	1	1		1
B	*Thomisus blandus* Karsch, 1880	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Thomisus citrinellus* Simon, 1875	0	LC	C		1	1	1	1	1	1	1	1	1
M	*Thomisus congoensis* Comellini, 1957	1	LC	AE			1	1	1	1	1		1	
B	*Thomisus dalmasi* Lessert, 1919	1	LC	AE			1	1	1	1	1	1		1
B	*Thomisus daradioides* Simon, 1890	0	LC	C			1	1	1	1	1	1		1
B	*Thomisus granulatus* Karsch, 1880	1	LC	AE		1		1	1	1	1			1
B	*Thomisus kalaharinus* Lawrence, 1936	1	LC	AE		1	1	1	1	1	1	1	1	1
M	*Thomisus machadoi* Comellini, 1959	1	LC	AE		1	1	1		1		1		1
F	*Thomisus natalensis* Lawrence, 1942	2	LC	STHE					1					
F	*Thomisus schultzei* Simon, 1910	1	LC	STHE			1	1				1	1	
B	*Thomisus scrupeus* (Simon, 1886)	1	LC	AE		1		1	1	1	1		1	1
B	*Thomisus spiculosus* Pocock, 1901	1	LC	AE				1	1	1	1			
B	*Thomisus stenningi* Pocock, 1900	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Thomisus zuluanus* Lawrence, 1942	5	DDT	SAE	KZNE				1					
B	*Tmarus africanus* Lessert, 1919	1	LC	AE		1		1	1	1	1	1	1	1
B	*Tmarus cameliformis* Millot, 1942	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Tmarus cancellatus* Thorell, 1899	1	LC	AE		1			1	1	1			
B	*Tmarus comellinii* Garcia-Neto, 1989	1	LC	AE		1	1	1	1	1	1	1	1	1
B	*Tmarus foliatus* Lessert, 1928	1	LC	AE		1	1	1	1	1	1		1	1
M	*Tmarus guineensis* Millot, 1942	1	LC	AE								1		1
B	*Tmarus longicaudatus* Millot, 1942	1	LC	AE					1	1		1		1
M	*Tmarus natalensis* Lessert, 1925	3	LC	SAE		1			1	1				1
B	*Tmarus planetarius* Simon, 1903	1	LC	AE					1	1				1
B	*Trichopagis manicata* Simon, 1886	1	LC	AE					1	1				
B	*Xysticus fagei* Lessert, 1919	1	LC	AE				1		1	1		1	
F	*Xysticus havilandi* Lawrence, 1942	3	LC	SAE		1	1	1	1					1
F	*Xysticus lucifugus* Lawrence, 1937	3	LC	SAE					1	1	1			1
F	*Xysticus mulleri* Lawrence, 1952	2	LC	STHE		1		1	1	1	1	1	1	
F	*Xysticus namaquensis* Simon, 1910	5	DDT	SAE	NCE							1		
B	*Xysticus natalensis* Lawrence, 1938	2	LC	STHE		1	1	1	1	1	1		1	
B	*Xysticus sagittifer* Lawrence, 1927	2	LC	STHE			1							1
J	*Xysticus simonstownensis* Strand, 1909	6	DDT	SAE	WCE									1
F	*Xysticus tugelanus* Lawrence, 1942	2	LC	STHE		1		1	1					1
B	*Xysticus urbensis* Lawrence, 1952	2	LC	STHE		1		1					1	1
FAMILY TRACHELIDAE Simon, 1897														
B	*Afroceto africana* (Simon 1910)	2	LC	STHE		1	1		1	1		1		1
M	*Afroceto ansieae* Lyle, 2015	6	DDT	SAE	KZNE				1					
M	*Afroceto bisulca* Lyle & Haddad, 2010	6	DDT	SAE	WCE									1
F	*Afroceto bulla* Lyle & Haddad, 2010	6	DDT	SAE	ECE	1								
B	*Afroceto capensis* Lyle & Haddad, 2010	5	Rare	SAE	WCE									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Afroceto coenosa* (Simon, 1897)	4	LC	SAE					1		1			
F	*Afroceto corcula* Lyle & Haddad, 2010	4	DDT	SAE								1		1
B	*Afroceto croeseri* Lyle & Haddad, 2010	4	DD	SAE		1			1					
M	*Afroceto dippenaarea* Lyle, 2015	6	DDT	SAE	WCE									1
B	*Afroceto flabella* Lyle & Haddad, 2010	6	DD	SAE	ECE	1								
M	*Afroceto gracilis* Lyle & Haddad, 2010	5	DDT	SAE	ME						1			
B	*Afroceto martini* (Simon, 1897)	2	LC	STHE		1	1	1	1	1	1	1		1
B	*Afroceto plana* Lyle & Haddad, 2010	1	LC	AE		1	1		1	1				1
M	*Afroceto porrecta* Lyle & Haddad, 2010	4	DDT	SAE		1								1
B	*Afroceto rotunda* Lyle & Haddad, 2010	6	DD	SAE	NCE							1		
B	*Afroceto spicula* Lyle & Haddad, 2010	5	DD	SAE	NCE							1		
B	*Capobula capensis* Haddad, Jin, Platnick & Booysen, 2021	5	LC	SAE	WCE									1
B	*Capobula infima* (Simon, 1896)	5	LC	SAE	WCE									1
B	*Capobula capensis* Haddad, Jin, Platnick & Booysen, 2021	5	LC	SAE	WCE									1
B	*Capobula infima* (Simon, 1896)	5	LC	SAE	WCE									1
B	*Capobula montana* Haddad, Jin, Platnick & Booysen, 2021	3	LC	SAE		1	1							
F	*Capobula neethlingi* Haddad, Jin, Platnick & Booysen, 2021	5	LC	SAE	WCE									1
F	*Capobula ukhahlamba* Haddad, Jin, Platnick & Booysen, 2021	5	LC	SAE	KZNE				1					
B	*Fuchiba aquilonia* Haddad & Lyle, 2008	2	LC	STHE					1	1	1			
B	*Fuchiba capensis* Haddad & Lyle, 2008	4	LC	SAE		1								1
B	*Fuchiba montana* Haddad & Lyle, 2008	2	LC	STHE		1	1							1
F	*Fuchiba similis* Haddad & Lyle, 2008	6	DDT	SAE	KZNE				1					
B	*Fuchiba tortilis* Haddad & Lyle, 2008	5	DD	SAE	ECE	1								
B	*Fuchiba venteri* Haddad & Lyle, 2008	6	DD	SAE	WCE									1
B	*Fuchibotulus bicornis* Haddad & Lyle, 2008	5	VU	SAE	WCE									1
F	*Fuchibotulus haddadi* Lyle, 2013	6	DDT	SAE	ECE	1								
B	*Fuchibotulus kigelia* Haddad & Lyle, 2008	2	LC	STHE			1	1	1	1			1	
B	*Jocquestus capensis* Lyle & Haddad, 2018	5	DD	SAE	WCE									1
F	*Jocquestus harrisi* Lyle & Haddad, 2018	6	DDT	SAE	LE					1				
B	*Jocquestus incurvus* Lyle & Haddad, 2018	4	Rare	SAE						1	1			
B	*Jocquestus roeweri* (Lawrence, 1938)	5	DD	SAE	KZNE				1					
B	*Jocquestus schenkeli* (Lessert, 1923)	1	LC	AE		1			1	1	1			
B	*Orthobula arca* Haddad, Jin & Platnick, 2022	3	LC	SAE			1	1	1					
B	*Orthobula radiata* Simon, 1897	1	LC	AE				1	1	1	1	1	1	
B	*Patelloceto secutor* Lyle & Haddad, 2010	2	LC	STHE					1		1			
B	*Planochelas haddadi* Khoza & Lyle, 2019	6	DD	SAE	KZNE									1
B	*Planochelas neethling* Khoza & Lyle, 2019	6	DD	SAE	KZNE									1
F	*Poachelas montanus* Haddad & Lyle, 2008	3	LC	SAE		1	1							1
F	*Poachelas refugus* Haddad, 2010	3	DDT	SAE					1	1				
B	*Poachelas striatus* Haddad & Lyle, 2008	3	LC	SAE			1				1	1		
B	*Spinotrachelas capensis* Haddad, 2006	5	LC	SAE	WCE									1
F	*Spinotrachelas confinis* Lyle, 2011	6	DDT	SAE	WCE									1
B	*Spinotrachelas montanus* Haddad, Neethling & Lyle, 2011	4	VU	SAE			1		1					
F	*Spinotrachelas namaquensis* Lyle, 2011	6	DDT	SAE	NCE							1		
M	*Spinotrachelas similis* Lyle, 2011	6	DDT	SAE	WCE									1
B	*Thysanina absolva* Lyle & Haddad, 2006	5	LC	SAE	FSE		1							
B	*Thysanina capensis* Lyle & Haddad, 2006	4	LC	SAE								1		1
B	*Thysanina gracilis* Lyle & Haddad, 2006	2	LC	STHE			1					1		
B	*Thysanina serica* Simon, 1910	2	LC	STHE				1		1				
B	*Thysanina transversa* Lyle & Haddad, 2006	3	LC	SAE		1			1	1	1			1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
M	*Trachelas pusillus* Lessert, 1923	1	LC	AE		1	1	1	1	1	1	1	1	1
F	*Trachelas scopulifer* Simon, 1896	3	DDT	SAE						1				1
FAMILY TROCHANTERIIDAE Karsch, 1879														
B	*Platyoides alpha* Lawrence, 1928	2	LC	STHE						1				
F	*Platyoides costeri* Tucker, 1923	5	DDT	SAE	WCE									1
B	*Platyoides leppanae* Pocock, 1902	1	LC	AE		1			1					1
F	*Platyoides pictus* Pocock, 1902	4	DDT	SAE		1								1
F	*Platyoides pirie* Platnick, 1985	3	LC	SAE		1			1					
B	*Platyoides pusillus* Pocock, 1898	1	LC	AE		1			1	1				1
F	*Platyoides quinquedentatus* Purcell, 1907	5	DDT	SAE	WCE									1
F	*Platyoides robertsi* Haddad, 2022	6	DD	SAE	NCE							1		
F	*Platyoides rossi* Platnick, 1985	6	DDT	SAE	ECE	1								
B	*Platyoides walteri* (Karsch, 1886)	1	LC	AE		1	1	1	1	1	1	1	1	1
FAMILY ULOBORIDAE Thorell, 1869														
F	*Hyptiotes akermani* Wiehle, 1964	3	LC	SAE		1		1	1	1				
F	*Miagrammopes brevicaudus* O.P.-Cambridge, 1882	2	LC	STHE		1		1	1	1	1	1	1	1
F	*Miagrammopes constrictus* Purcell, 1904	3	LC	SAE		1		1	1	1	1	1	1	1
B	*Miagrammopes longicaudus* O.P.-Cambridge, 1882	2	LC	STHE		1								
M	*Philoponella angolensis* (Lessert, 1933)	1	LC	AE		1			1	1				1
F	*Philoponella operosa* (Simon, 1896)	6	DDT	SAE	WCE									1
F	*Uloborus planipedius* Simon, 1896	3	LC	SAE		1			1	1	1			1
B	*Uloborus plumipes* Lucas, 1846	0	LC	C		1	1	1	1	1	1	1	1	1
B	*Uloborus walckenaerius* Latreille, 1806	0	LC	C			1					1		1
B	*Zosis geniculata* (Olivier, 1789)	1	LC	C		1			1	1				1
FAMILY ZODARIIDAE Thorell, 1881														
F	*Akyttara homunculus* Jocqué, 1991	2	LC	STHE						1		1		
M	*Australutica africana* Jocqué, 2008	5	Rare	SAE	LE					1				
M	*Australutica normanlarseni* Jocqué, 2008	5	DDT	SAE	WCE									1
B	*Ballomma erasmus* Jocqué & Henrard, 2015	6	DDT	SAE	LE					1				
B	*Ballomma haddadi* Jocqué & Henrard, 2015	4	DD	SAE						1	1			
F	*Ballomma legala* Jocqué & Henrard, 2015	6	DD	SAE	LE					1				
B	*Ballomma neethlingi* Jocqué & Henrard, 2015	5	DD	SAE	LE					1				
F	*Caesetius bevisi* (Hewitt, 1916)	2	LC	STHE					1	1				
B	*Caesetius biprocessiger* (Lawrence, 1952)	5	LC	SAE	KZNE				1					
B	*Caesetius flavoplagiatus* Simon, 1910	2	LC	STHE								1		
M	*Caesetius globicoxis* (Lawrence, 1942)	3	LC	SAE					1	1	1			1
B	*Caesetius inflatus* Jocqué, 1991	1	LC	AE					1	1	1			
B	*Caesetius murinus* Simon, 1893	4	LC	SAE		1								1
J	*Caesetius politus* Simon, 1893	6	DDT	SAE	LE					1				
B	*Caesetius schultzei* Simon, 1910	4	LC	SAE								1		1
F	*Caesetius spenceri* (Pocock, 1900)	3	LC	SAE		1					1			
M	*Capheris abrupta* Jocqué, 2009	4	LC	SAE								1		
B	*Capheris crassimana* (Simon, 1887)	2	LC	STHE						1		1		
B	*Capheris decorata* Simon, 1904	1	LC	AE			1	1	1	1	1	1		1
B	*Capheris fritzsimonsi* Lawrence, 1936	2	LC	STHE						1		1		
F	*Capheris langi* Lawrence, 1936	2	LC	STHE						1				
B	*Capheris subtilis* Jocqué, 2009	2	LC	STHE						1				
B	*Chariobas cylindraceus* Simon, 1893	1	LC	AE				1	1	1			1	1
J	*Chariobas lineatus* Pocock, 1900	4	LC	SAE		1								1
J	*Chariobas mamillatus* Strand, 1909	6	DDT	SAE	WCE									1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
J	*Chariobas navigator* Strand, 1909	6	DDT	SAE	WCE									1
B	*Cicynethus acer* Jocqué & Henrard, 2018	2	LC	STHE						1	1			
B	*Cicynethus decoratus* (Lawrence, 1952)	4	DD	SAE		1			1					
B	*Cicynethus floriumfontis* Jocqué, 1991	5	DD	SAE	ECE	1								
F	*Cicynethus peringueyi* (Simon, 1893)	5	DDT	SAE	WCE									1
B	*Cicynethus subtropicalis* (Lawrence, 1952)	5	LC	SAE	KZNE				1					
F	*Cydrela friedlanderae* Hewitt, 1914	5	DDT	SAE	NCE							1		
B	*Cydrela schoemanae* Jocqué, 1991	3	LC	SAE				1		1				
F	*Cydrela spinifrons* Hewitt, 1915	3	LC	SAE						1		1		
M	*Cydrela spinimana* Pocock, 1898	3	LC	SAE					1	1	1			
M	*Cydrela unguiculata* (O.P.-Cambridge, 1870)	6	DDT	SAE	KZNE					1				
M	*Cyrioctea lotzi* Jocqué, 2013	5	DDT	SAE	FSE		1							
M	*Cyrioctea marken* Platnick & Jocqué, 1992	5	DDT	SAE	LE					1				
M	*Cyrioctea sawadee* Jocqué, 2013	6	DDT	SAE	WCE									1
B	*Diores annetteae* Jocqué, 1990	3	LC	SAE				1		1	1			
B	*Diores auricula* Tucker, 1920	2	LC	STHE						1				
B	*Diores bifurcatus* Tucker, 1920	5	LC	SAE	WCE									1
F	*Diores bivattatus* Simon, 1893	6	DDT	SAE	WCE									1
M	*Diores capensis* Tucker, 1920	5	VU	SAE	WCE									1
M	*Diores cognatus* O.P.-Cambridge, 1904	6	DDT	SAE	WCE?									1
B	*Diores decipiens* Jocqué, 1990	6	DD	SAE	WCE									1
B	*Diores dowsetti* Jocqué, 1990	5	VU	SAE	WCE									1
B	*Diores femoralis* Jocqué, 1990	3	LC	SAE			1	1					1	
F	*Diores godfreyi* Hewitt, 1919	6	DDT	SAE	WCE									1
M	*Diores griswoldorum* Jocqué, 1990	2	LC	STHE								1		
B	*Diores jonesi* Tucker, 1920	3	LC	SAE					1	1				
B	*Diores leleupi* Jocqué, 1990	5	Rare	SAE	WCE									1
B	*Diores lesserti* Lawrence, 1952	3	LC	STHE					1	1	1			
B	*Diores magicus* Jocqué & Dippenaar-Schoeman, 1992	2	LC	STHE						1				
B	*Diores pauper* Jocqué, 1990	3	LC	SAE		1				1				
B	*Diores poweri* Tucker, 1920	2	LC	STHE		1	1	1	1		1	1	1	1
B	*Diores radulifer* Simon, 1910	5	LC	SAE	NCE							1		
B	*Diores rectus* Jocqué, 1990	1	LC	AE				1		1	1			
B	*Diores recurvatus* Jocqué, 1990	2	LC	STHE			1	1		1	1		1	
B	*Diores russelli* Jocqué, 1990	2	LC	STHE			1							
M	*Diores sequax* Jocqué, 1990	6	DDT	SAE	KZNE				1					
F	*Diores setosus* Tucker, 1920	5	Rare	SAE	WCE								1	
B	*Diores silvestris* Jocqué, 1990	5	Rare	SAE	WCE								1	
B	*Diores simoni* O.P.-Cambridge, 1904	5	LC	SAE	WCE								1	
B	*Diores simplicior* Jocqué, 1990	1	LC	AE					1					
F	*Diores spinulosus* Jocqué, 1990	5	DDT	SAE	ECE	1								
B	*Diores termitophagus* Jocqué & Dippenaar-Schoeman, 1992	4	DD	SAE		1	1							
B	*Diores triangulifer* Simon, 1910	2	LC	STHE			1	1		1		1	1	
B	*Diores triarmatus* Lessert, 1929	1	LC	AE			1	1		1	1			
B	*Diores youngai* Jocqué, 1990	5	LC	SAE	WCE									1
F	*Heradida bicincta* Simon, 1910	2	LC	STHE						1		1		
F	*Heradida extima* Jocqué, 1987	4	LC	SAE		1						1		1
F	*Heradida loricata* Simon, 1893	4	LC	SAE			1							
F	*Heradida speculigera* Jocqué, 1987	4	LC	SAE				1						1

		D	CON STATUS	ENDEMICITY	PROV	EC	FS	G	KZN	L	M	NC	NW	WC
F	*Heradida xerampelina* Benoit, 1974	6	DDT	SAE	WCE									1
B	*Hermippus loricatus* Simon, 1893	1	LC	AE										
F	*Hermippus septemguttatus* Lawrence, 1942	5	LC	SAE	KZNE				1					
M	*Hermippus tenebrosus* Jocqué, 1986	3	DDT	SAE					1	1				
B	*Mallinus nitidiventris* Simon, 1893	3	LC	SAE		1	1					1		1
B	*Palfuria caputlari* Szűts & Jocqué, 2001	1	LC	AE						1				
F	*Palfuria retusa* Simon, 1910	2	LC	STHE								1		
B	*Palfuria spirembolis* Szűts & Jocqué, 2001	2	LC	STHE						1	1			
F	*Procydrela limacola* Jocqué, 1999	5	DDT	SAE	WCE									1
B	*Procydrela precursor* Jocqué, 1999	4	EN	SAE		1								1
B	*Psammoduon arenicola* (Simon, 1910)	5	VU	SAE	WCE									1
B	*Psammoduon canosum* (Simon, 1910)	2	LC	STHE								1		1
B	*Psammoduon deserticola* (Simon, 1910)	2	LC	STHE						1		1		
F	*Psammorygma aculeatum* (Karsch, 1878)	3	LC	SAE					1	1	1			
F	*Psammorygma rutilans* (Simon, 1887)	6	DDT	SAE	?									
B	*Ranops caprivi* Jocqué, 1991	2	LC	STHE			1	1	1	1	1			
B	*Ranops robinae* Jocqué & Henrard, 2020	3	LC	SAE			1	1		1	1			
B	*Rotundrela orbiculata* Jocqué, 1999	5	Rare	SAE	WCE									1
B	*Rotundrela rotunda* Jocqué, 1999	5	EN	SAE	WCE									1
B	*Systenoplacis fagei* (Lawrence, 1936)	3	LC	SAE					1	1				1
B	*Systenoplacis vandami* (Hewitt, 1916)	3	LC	SAE		1		1	1	1		1	1	
F	*Thaumastochilus martini* Simon, 1897	5	DDT	SAE	KZNE				1					
B	*Thaumastochilus termitomimus* Jocqué, 1994	3	DD	SAE				1	1					
FAMILY ZOROPSIDAE Bertkau, 1882														
B	*Griswoldia acaenata* (Griswold, 1991)	4	LC	SAE		1								1
B	*Griswoldia disparilis* (Lawrence, 1952)	4	DD	SAE		1			1					
F	*Griswoldia leleupi* (Griswold, 1991)	5	LC	SAE	LE					1				
B	*Griswoldia meikleae* (Griswold, 1991)	6	DD	SAE	WCE									1
F	*Griswoldia melana* (Lawrence, 1938)	5	VU	SAE	KZNE				1					
M	*Griswoldia natalensis* (Lawrence, 1938)	6	DDT	SAE	KZNE				1					
B	*Griswoldia punctata* (Lawrence, 1942)	5	NT	SAE	KZNE				1					
B	*Griswoldia robusta* (Simon, 1898)	5	LC	SAE	WCE									1
B	*Griswoldia sibyna* (Griswold, 1991)	5	DD	SAE	WCE									1
B	*Griswoldia transversa* (Griswold, 1991)	4	DD	SAE		1			1					
B	*Griswoldia urbensis* (Lawrence, 1942)	4	LC	SAE		1			1					
B	*Griswoldia zuluensis* (Lawrence, 1938)	5	EN	SAE	KZNE				1					
B	*Phanotea cavata* Griswold, 1994	4	LC	SAE		1								1
B	*Phanotea ceratogyna* Griswold, 1994	5	Rare	SAE	WCE									1
B	*Phanotea digitata* Griswold, 1994	5	LC	SAE	WCE									1
B	*Phanotea knysna* Griswold, 1994	5	DD	SAE	WCE									1
F	*Phanotea lata* Griswold, 1994	6	DDT	SAE	WCE									1
F	*Phanotea latebricola* Lawrence, 1952	6	DDT	SAE	KZNE				1					
F	*Phanotea margarita* Griswold, 1994	6	DDT	SAE	WCE									1
B	*Phanotea natalensis* Lawrence, 1951	6	DD	SAE	KZNE				1					
M	*Phanotea orestria* Griswold, 1994	6	DDT	SAE	WCE									1
B	*Phanotea peringueyi* Simon, 1896	5	Rare	SAE	WCE									1
B	*Phanotea sathegyna* Griswold, 1994	5	DD	SAE	ECE	1								
F	*Phanotea simoni* Lawrence, 1951	5	DDT	SAE	KZNE				1					
B	*Phanotea xhosa* Griswold, 1994	4	LC	SAE		1								1

Discussion and conclusion

South African spider systematics and ecology are in an exploratory phase and the traditional approach to mapping diversity has enabled spider ecological research results in South Africa to generate species lists that are often resolved to species level. This descriptive phase provides the foundation for more integrative work and any attempts to ignore the importance of providing baseline diversity and taxonomic data will hamper subsequent attempts to develop a deeper understanding and appreciation of this unique heritage. This checklist provides a framework and context for future work.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

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Author contributions

ASD conceptualized, compiled data and wrote the first draft; CRH data curation and commented on drafts of the manuscript; RB data curation and commented on drafts of the manuscript; LL data curation and commented on drafts of the manuscript; RS data curation and commented on drafts of the manuscript; SHF data analysis and commented on drafts of the manuscript.

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Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

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Supplementary material 1

Reference list and citations for all the studies mentioned in the paper that are in grey literature

Authors: Ansie S. Dippenaar-Schoeman, Charles R. Haddad, Leon N. Lotz, Ruan Booysen, Rudolph C. Steenkamp, Stefan H. Foord

Data type: docx

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Link: https://doi.org/10.3897/AfrInvertebr.64.111047.suppl1

Supplementary material 2

List of Photo Identification Guides available from the World Spider Catalogue or Zenodo

Authors: Ansie S. Dippenaar-Schoeman, Charles R. Haddad, Leon N. Lotz, Ruan Booysen, Rudolph C. Steenkamp, Stefan H. Foord

Data type: docx

Explanation note: Each guide contains information on the genera and species recorded from South Africa with information on global distribution, South African distribution with a map, lifestyle, conservation and taxonomic information and images and photographs.

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Research Article

A survey of Gnaphosidae (Arachnida, Araneae) from Ascension Island with description of a new species of *Australoechemus* Schmidt & Piepho, 1994

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Abstract

An updated checklist of the gnaphosid spiders of Ascension Island – comprising five species, in five genera – is presented, based on examination of historical and new specimens from across Ascension, including its islet Boatswain Bird Island. The continued presence of the non-native *Marinarozelotes jaxartensis* (Kroneberg, 1875), *Urozelotes rusticus* (L. Koch, 1842), and *Zelotes laetus* (O. Pickard-Cambridge, 1872), previously recorded by prior workers, is confirmed. Two species are newly recorded from the island: *Synaphosus syntheticus* (Chamberlin, 1924) from the mainland and *Australoechemus vickyae* **sp. nov.** (♂♀) from both Boatswain Bird Island and the mainland.

Key words: Aranei, distribution, gnaphosid, inventory, new species, taxonomy, United Kingdom Overseas Territories

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Introduction

Ascension Island is a remote volcanic island in the South Atlantic Ocean, part of the United Kingdom Overseas Territory of Saint Helena, Ascension and Tristan da Cunha. The invertebrate biodiversity of the island is understudied, especially the spider fauna which has hitherto only received general faunistic attention in three prior works (Duffey 1964; Ashmole and Ashmole 1997, 2000). The family Gnaphosidae Banks, 1892, commonly known as ground spiders, currently contains 150 genera and 2458 species (WSC 2023) of which only a handful have been recorded from the island, either at species or generic level (Duffey 1964; Ashmole and Ashmole 1997, 2000).

Recently, the senior author has been sorting the collection of spiders made by E. A. Duffey from Ascension Island (Duffey 1964), an expedition which also included one of the other authors (Philip Ashmole); also sorting newly collected

specimens sent by author Adam Sharp. This has revealed a total of five species of gnaphosid, three previously known from the island (sometimes under names which are now considered as junior synonyms) and two new for Ascension Island. Of these newly recorded morphospecies, one corresponds to *Synaphosus syntheticus* (Chamberlin, 1924) and the other is an undescribed species of *Australoechemus* Schmidt & Piepho, 1994.

In this work, we provide an updated list of the gnaphosid spiders of Ascension based on examined specimens, also providing complimentary photographs of the genitalia of *S. syntheticus* and describing *Australoechemus vickyae* sp. nov. from both sexes.

Materials and methods

Images of specimens were made using a Canon EOS 6D Mark II attached to a Leica MZ12.5 stereomicroscope, with images stacked using Helicon Focus software. Abbreviations: **Ah** = anterior hood; **Al** = tegular anteroprolateral membranous lamellar extension; **ASC** = Ascension Island Conservation Directorate collection, Georgetown, Ascension Island (it is intended in the future that the ASC invertebrate collection will be donated and moved to the Saint Helena National Trust, Jamestown, Saint Helena); **Bc** = bursa copulatrix; **Co** = conductor; **Em** = embolus; **Fe** = femur; **MMUE** = Manchester Museum, Manchester, United Kingdom; **Mt** = metatarsus; **NHMUK** = Natural History Museum, London, United Kingdom; **OUMNH** = Oxford University Museum of Natural History, Oxford, United Kingdom; **Pa** = patella; **Re** = receptacles; **Sd** = sperm duct; **St** = subtegulum; **Ta** = tarsus; **Ti** = tibia; **WSC** = World Spider Catalog. Setation: d = dorsal, p = prolateral, r = retrolateral, v = ventral, macrosetae are scored in posterior, median and anterior thirds (e.g. 1–2–3). Total lengths include chelicerae but exclude spinnerets. Leg measurements are presented as: total length (femur, patella, tibia, metatarsus, tarsus). All measurements in mm. References are provided for most relevant publications dealing with taxonomy of the species. Authors' emphases in [ ].

Results

Australoechemus Schmidt & Piepho, 1994

Australoechemus Schmidt & Piepho in Schmidt et al. 1994: 101.

Type species. *Australoechemus oecobiophilus* Schmidt & Piepho, 1994 by original designation.

Diagnosis (tentative, based on species known from adults but not the type species, known only from an immature holotype). Murphy (2007: 13) placed *Australoechemus* into the *Drassodes* Group; genera of this informal grouping were defined based on to their relatively large size, "plain, tawny-light, light greyish abdomen, occasionally patterned", presence of a notch at the apex of the trochanters, and absence of a dorsal scutum in the male. At the generic level, *Australoechemus* (*sensu* Murphy, 2007) differs from other gnaphosids by the male palp with a U-shaped sperm duct not meandering, running along margins of the tegulum (vs. meandering, or not located on the margin of the tegulum in other presently known genera) and the presence of a tegular an-

teroprolateral membranous lamellar extension (*AI*) (*AI* absent in other presently known genera).

Remarks. This genus was described to accommodate two new species from the Cape Verde Islands: *A. oecobiophilus* Schmidt & Piepho, 1994 and *A. celer* Schmidt & Piepho, 1994. *Australoechemus oecobiophilus* was chosen as the type species although its holotype is a juvenile specimen. The diagnosis and illustrations given in the paper are very poor, for instance the generic diagnosis comprises solely the following short and vague sentence: "Von *Echemus* Simon, 1878 durch das Fehlen eines Putzkammes an Metatarsus III und IV unterschieden. Die Gattung scheint *Xerophaeus* Purcell, 1907 nahezustehen" ["Distinguished from *Echemus* Simon, 1878 by the absence of preening combs on metatarsi III and IV. The genus appears to be close to *Xerophaeus* Purcell, 1907"] (Schmidt et al. 1994: 101). A detailed illustrated redescription is given for *A. celer* by Murphy (2007), and it is on this greatly improved work that we decide to place *A. vickyae* sp. nov. in this genus, given general similarities in the overall structure of the male palp and female epigyne. It is essential that future workers clarify the taxonomy of the type species *A. oecobiophilus* based on adult specimens. This is outside the scope of the present work.

***Australoechemus vickyae* sp. nov.**
https://zoobank.org/66935B7B-7C32-4276-9A5B-1707D8DA4ACC
Figs 1–4

Material examined. *Holotype*: Ascension Island • 1♂; Boatswain Bird Island; -7.936, -14.307; 12–31.IX.1957; E. Duffey leg.; No. 91; NHMUK.

***Paratypes*:** Ascension Island • 1♀; Boatswain Bird Island; -7.936, -14.307; 12–31.IX.1957; E. Duffey leg.; [no number]; NHMUK • 1♀; Boatswain Bird Island; -7.936, -14.307; 12–31.IX.1957; E. Duffey leg.; No. 35; NHMUK • 1♀; Boatswain Bird Island; -7.936, -14.307; 12–31.IX.1957; E. Duffey leg.; No. 51; NHMUK • 1♂, 1 immature ♂; Boatswain Bird Island; -7.936, -14.307; 12–31.IX.1957; E. Duffey leg.; No. 65; NHMUK • 1♀; Boatswain Bird Island; -7.936, -14.307; 26–27.V.1995; P. Ashmole and M. Ashmole leg.; ?*Nodocion* sp. det. J. A. Murphy; Murphy collection No. 23865; MMUE G7572.13477 • 1♀; South Gannet Hill, Ascension Island; -7.983, -14.399; 15.V.2013; pitfall trap; L. F. White leg.; ASC01604 • 1♀; near English Bay Road, Ascension Island; -7.913981, -14.378077; 85 m.a.s.l; pitfall trap; 13/01/2022; A. Sharp leg.; ASC F13 2 PFJ.

Diagnosis. *Australoechemus vickyae* sp. nov. somewhat resembles *A. celer*, but males can be distinguished by having the retrolateral tibial apophysis almost as long as tibia, tapering at the tip and non-bifurcated (vs. shorter than tibia, broad at tip and bifurcated). Females of *A. vickyae* sp. nov. differ from those of *A. celer* by having the epigynal fovea widest in anterior part (vs. wider in midpart) and having receptacles smaller than the bursa copulatrix (vs. receptacles larger than bursa copulatrix).

Etymology. The specific epithet is a matronym in honour of British conservationist and entomologist Vicky Wilkins (Species Recovery Trust, Salisbury, UK, and co-chair of the IUCN SSC Mid-Atlantic Islands Invertebrate Specialist Group) for her enduring and significant contributions to the conservation of invertebrates in the United Kingdom Overseas Territories.

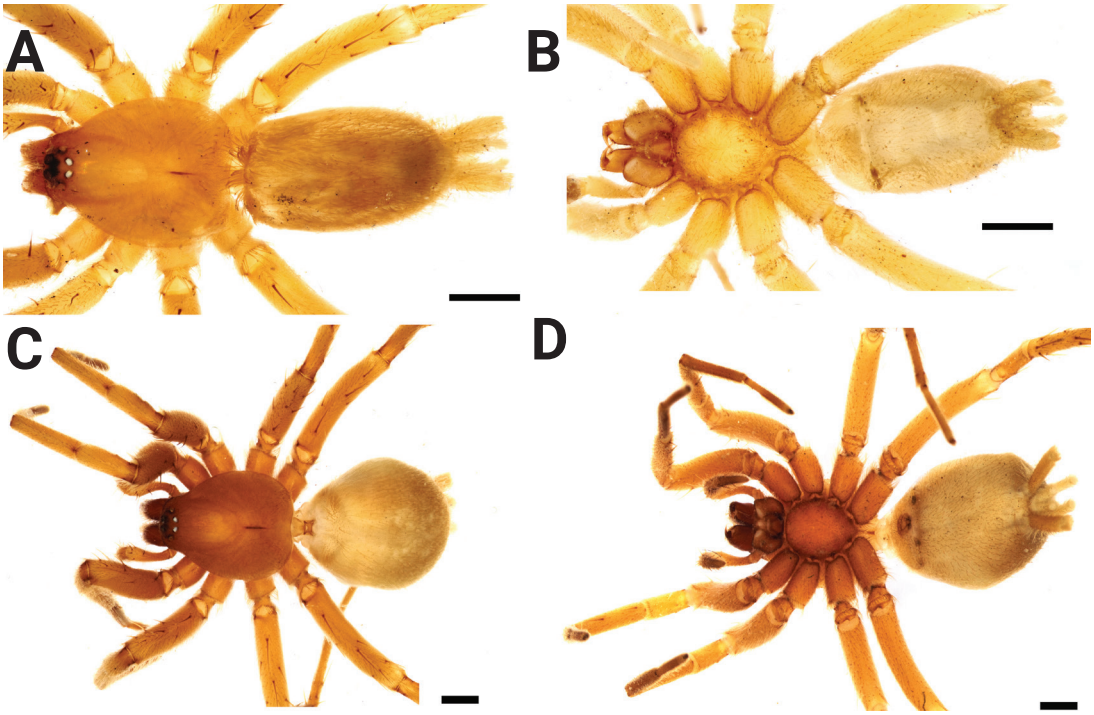

Figure 1. Habitus of *Australoechemus vickyae* sp. nov. **A** holotype male, dorsal **B** ibid, ventral **C** paratype female, dorsal **D** ibid, ventral. Scale bars: 1 mm.

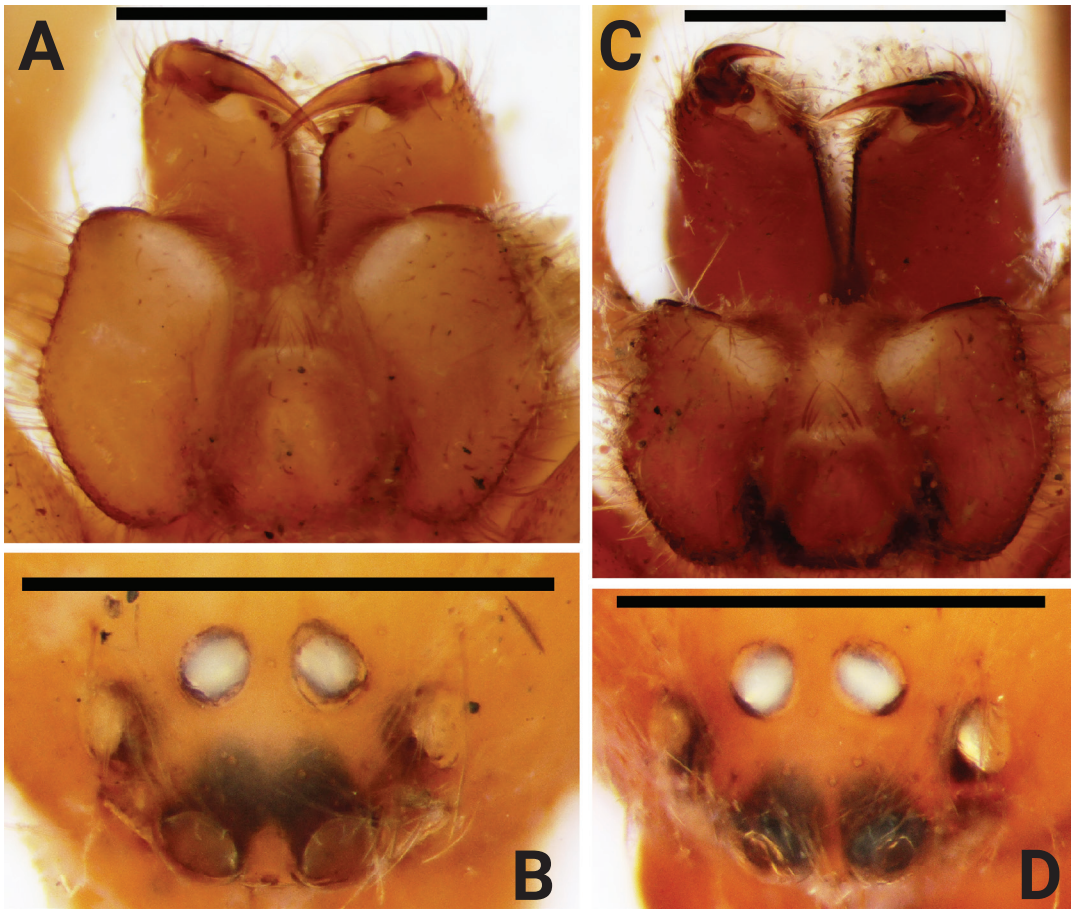

Figure 2. Ventral face of chelicera and eye pattern of *Australoechemus vickyae* sp. nov. **A** holotype male, chelicera **B** holotype male, eye pattern **C** paratype female, chelicera **D** paratype female, eye pattern. Scale bars: 1 mm.

Description. Male (holotype). ***Total length*:** 6.17. Carapace: 2.67 long, 2.12 wide. ***Abdomen*:** 2.89 long, 1.75 wide. ***Leg measurements*:** I 8.12 (2.28, 1.05, 2.02, 1.63, 1.14), II 8.00 (2.28, 1.03, 1.44, 1.93, 1.32), III 7.49 (2.21, 0.87, 1.24, 1.85, 1.32), IV 11.02 (2.36, 1.56, 2.39, 3.28, 1.43). ***Setation*:** for legs see Table 1. ***Colour*** (in alcohol): overall light brown, abdomen slightly darker than carapace, chelicerae and legs (Fig. 1A, B). Chelicera with three promarginal teeth and one retromarginal tooth (Fig. 2A). ***Eyes*:** AME largest, PME not reduced in size (Fig. 2B).

Palp (Figs 3A, 4A–D).

Patella longer than tibia, about 2× longer than wide; tibia about 2× longer than wide with non-bifurcated retrolateral apophysis gradually tapering toward tip, tip slightly bent prolaterally; cymbium elongate, gradually tapering, 2.2× longer than wide; bulb oval, 1.8× longer than wide, with very long subtegulum (*St*); sperm duct (*Sd*) U-shaped, retrolaterally as wide as tibial apophysis, gradually tapering prolaterally; anteroprolateral part of tegulum with elongate tegular anteroprolateral membranous lamellar extension (*Al*); conductor (*Co*) small, membranous, located near tip of embolus; embolus (*Em*) originates at about 9 o'clock position, straight, tip slightly bent. ***Palpal setation*:** femur d 0–1–4 r0–1–0 p0–1–0; patella d0–0–1; tibia d1–1–0.

Female (paratype). ***Total length*:** 8.27. ***Carapace*:** 3.53 long, 2.62 wide. ***Abdomen*:** 3.67 long, 2.63 wide. ***Leg measurements*:** I 8.96 (2.02, 1.63, 1.87, 1.98, 1.46), II 8.79 (2.01, 1.65, 1.72, 1.96, 1.45), III 8.29 (1.17, 1.38, 2.19, 2.00, 1.55), IV 11.42 (3.03, 1.82, 2.66, 2.57, 1.34). ***Setation*:** for legs see Table 2, palp: femur d0–1–2 r0–1–0 p0–1–0; patella p0–1–0 r0–1–0; tibia d1–1–1 p0–2–1; tarsus d2–0–0 p2–1–0. Colour (in alcohol): carapace and legs light brown, abdomen beige, chelicerae reddish-brown (Figs 1C, D, 2C). Chelicera with three promarginal teeth and one retromarginal tooth (Fig. 2C). ***Eyes*:** AME largest, PME not reduced in size (Fig. 2D).

Epigyne. Epigynal plate almost as long as wide, with large fovea and distinct anterior hood (*Ah*); fovea widest anteriorly, posterior part 2× thinner than anterior; bursa copulatrix (*Bc*) oval, longer than wide, spaced by about ½ of length; receptacles (*Re*) oval, transversal, locate at posterior margin of endogyne, spaced by about one width (Fig. 3B, C).

Table 1. Leg setation.

	Fe	Pa	Ti	Mt
I	d1–1–1 r0–1–0 p0–0–2	–	v1–2–2	v 2–0–0
II	d1–1–1 r0–1–1 p0–1–1	–	v0–0–2	–
III	d1–1–1 r0–1–1 p0–1–1	–	d1–2–3 v3–2–2	v 2–1–3
IV	d1–1–1 r0–1–1 p0–1–1	–	d1–2–1 v2–2–2	d2–4–3 v2–4–5

Table 2. Leg setation.

	Fe	Pa	Ti	Mt	Ta
I	d1–1–1 r0–1–0 p0–0–2	–	v1–1–1	v2–0–0	–
II	d1–1–1 r0–1–0 p0–2–1	–	v1–1–1	v2–0–0	–
III	d1–1–1 r0–1–1 p0–1–1	r0–1–0	d1–2–2 v2–2–3	d0–2–2 v2–1–2	–
IV	d1–1–1 r0–1–1 p0–1–1	r0–1–0	d2–2–3 v2–2–2	d2–3–2 v2–4–3	–

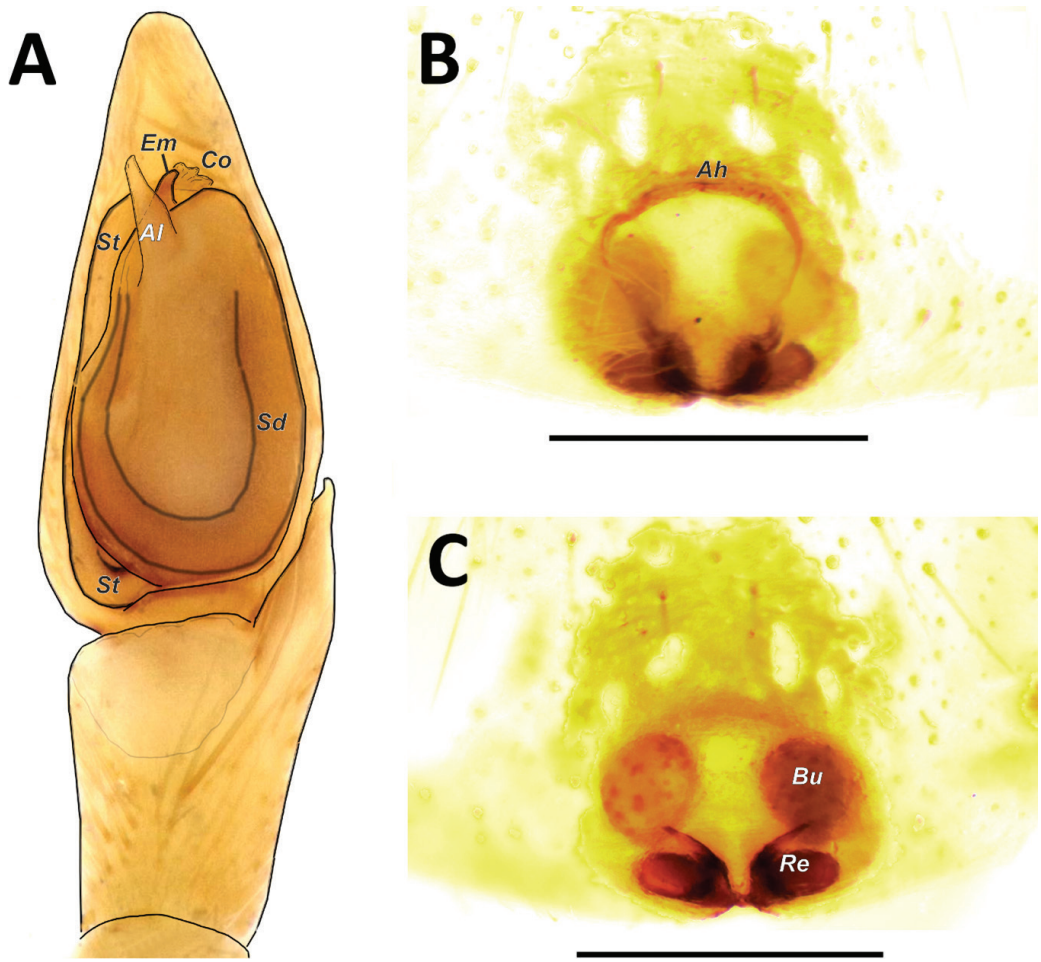

Figure 3. Copulatory organs of *Australoechemus vickyae* sp. nov. **A** holotype male palp, ventral, structures traced **B** paratype female epigyne (cleared), ventral **C** ibid, endogyne (cleared), dorsal. Abbreviations: *Ah* = anterior hood, *Al* = tegular anteroprolateral membranous lamellar extension, *Bu* = bursa copulatrix, *Co* = conductor, *Em* = embolus, *Re* = receptacle, *Sd* = sperm duct, *St* = subtegulum. Scale bars: 0.5 mm (**B, C**).

Total length variation. 6.17–7.37 (adult males, n=2); 8.25–9.24 (adult females; n=6).

Distribution. Known only from Ascension Island, South Atlantic Ocean.

Remarks. The holotype and most of the paratypes were collected from Boatswain Bird Island, an islet which hosts several unique endemic arachnids (Ashmole and Ashmole 2000). The paratype used for description is the unnumbered female in the Duffey collection, and it has also been clearly marked as the female used in the description on a newly added type label by DS. Two paratype females were collected recently on the mainland, indicating this species occurs more widely on Ascension. Eric Duffey was a prolific collector who collected all over the main island and Boatswain Bird Island (Stonehouse 1960; PA pers. obs.). Fortunately, the type specimens of *A. vickyae* sp. nov. collected by Duffey were explicitly collected by him only from Boatswain Bird Island (as "*Drassodes* spp." in Duffey 1964) enabling us to be sure of their collecting locality. This species represents an interesting new record of a genus only thought to occur in the Cape Verde Islands previously (WSC 2023). Molecular work in the future could further elucidate the higher-level relationships of this genus to other gnaphosids. Nonetheless, the morphology of the new species clearly favours placement in

Figure 4. General views of palp of holotype of *Australoechemus vickyae* sp. nov. **A** ventral **B** retrolateral **C** prolateral **D** dorsal. Scale bars: 1 mm.

Australoechemus under its current definitions for the following reasons: (1) the similar shape of the tegular anteroprolateral membranous lamellar extension, the shape of the cymbium, the U-shaped sperm duct, and the absence of a scutum in the male, (2) the epigyne with a large fovea and distinct anterior hood in the female and (3) the presence of a trochanteral notch in both sexes.

Marinarozelotes jaxartensis **(Kroneberg, 1875)**

Trachyzelotes jaxartensis: Levy 1998: 107, figs 25–28 (♂♀).
Marinarozelotes jaxartensis: Ponomarev and Shmatko 2020: 135, figs 7–8, 11, 30, 37–38, 50, 59 (♂♀).

Other citations. For complete list of taxonomic references, see WSC (2023).

Material examined. Ascension Island • 1♀; Packers Hole Cueva [Cave]; -7.923, -14.368; 13–17.III.1990; P. Ashmole and M. Ashmole leg.; ASC 0173 • 1♀; South Gannet Pools; -7.9865, -14.4015; 24.III.1990; P. Ashmole and M. Ashmole leg.; ASC 0259 • 1 immature; Letterbox [Peninsula]; -7.947, -14.297; 18–22 March 1990; P. Ashmole and M. Ashmole leg.; ASC 0745 • 1♀; Scouts Path [Green Mountain National Park]; -7.944, -14.346; 28.I.2020; [no collector stated]; ASC • 1♂; South Gannet Upper; -7.983, -14.399; 23–27 March 1990; P. Ashmole and M. Ashmole leg.; ASC 0592 • 1♀ Ascension Island; -7.94, -14.37; August–November 1957; E. A. Duffey leg.; No. 25; NHMUK.

Remarks. Invasive. Originally described from Central Asia, but now known to also occur across the Holarctic, and in Hawaii, India, Mexico, and South Africa (WSC 2023). First recorded from Ascension Island as *Camillina acanthognathus* (Purcell, 1907) by Duffey (1964). Specimens collected in the 1990s by Philip and Myrtle Ashmole (University of Edinburgh) indicate it is still found on the island.

Synaphosus syntheticus **(Chamberlin, 1924)**
Fig. 5

Synaphosus syntheticus: Ovtsharenko et al. 1994: 5, figs 1–2, 12–20 (♂♀).
Synaphosus syntheticus: Zamani et al. 2023: 355, figs 7A, 8D (♀, habitus and epigyne).

Other citations. For a complete list of taxonomic references, see WSC (2023).

Material examined. Ascension Island • 1♀ Travellers Hill; -7.94, -14.37; pitfall trap baited with fish; 11th March 2022; A. Sharp leg.; ASC G16 1 PFF • 1♂ same data as preceding but baited with meat; ASC G16 1 PFM.

Remarks. Invasive. Newly recorded for Ascension. *Synaphosus syntheticus* was initially described in *Nodocion* Chamberlin, 1922 and is often sympatric with *Nodocion s.s.* in the Nearctic. This species is also found in Africa (Egypt, Libya), the Middle East (Israel and Saudi Arabia) and is thought introduced to the United States and Mexico, although it was originally described from California (Ovtsharenko et al. 1994; WSC 2023). We provide complimentary images of the palp (Fig. 5A, B) and epigyne (Fig. 5C, D).

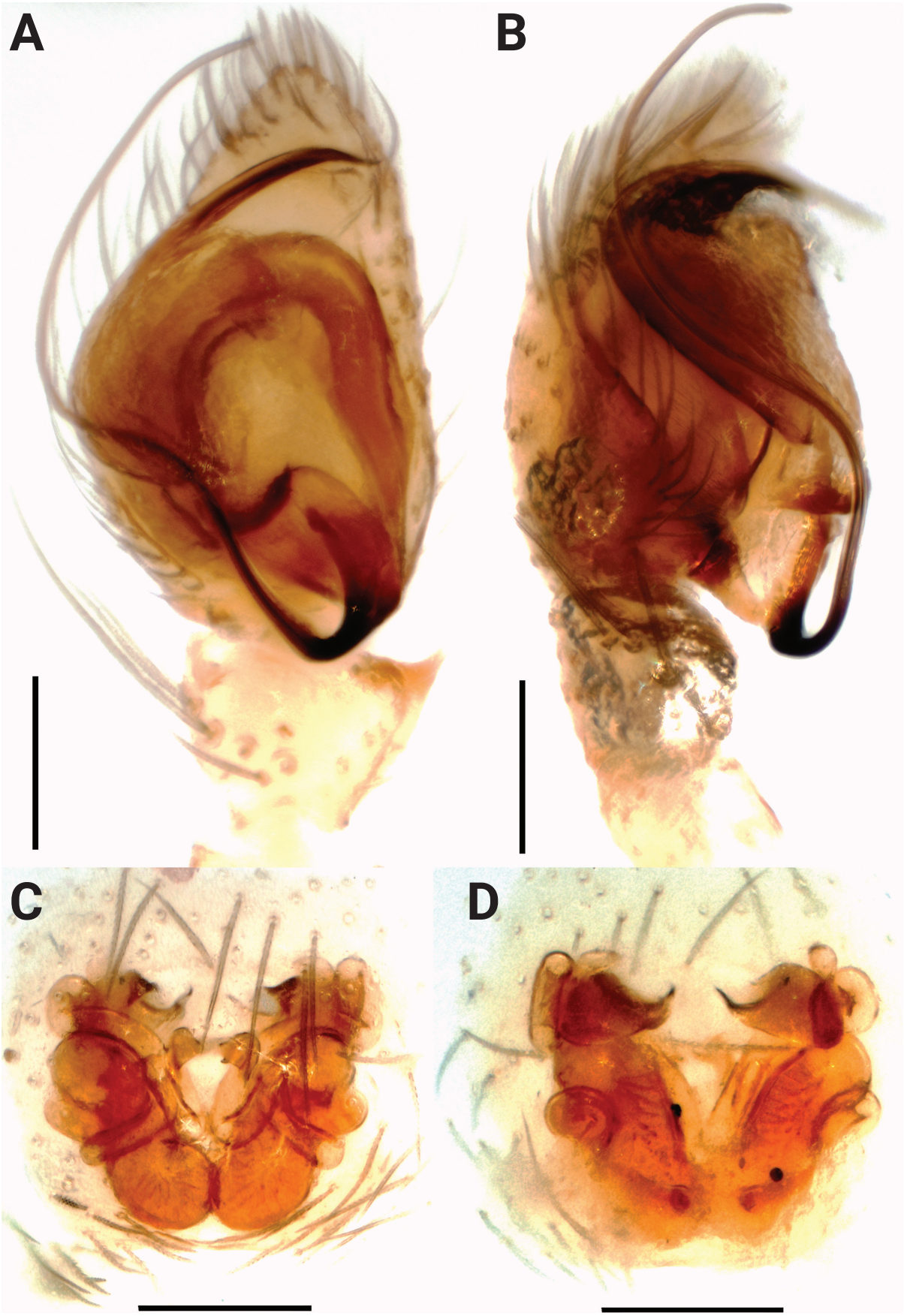

Figure 5. Copulatory organs of *Synaphosus syntheticus* Chamberlin, 1922 specimens from Ascension **A** male palp, ventral **B** Ibid, prolateral **C** epigyne (cleared), ventral **D** endogyne (cleared), dorsal. Scale bars: 0.1 mm.

Urozelotes rusticus (L. Koch, 1842)

Urozelotes rusticus: Platnick and Murphy 1984: 24, figs 55–58 (♂♀).
Zelotes rusticus: Grimm 1985: 221, figs 217b, 218b, 244, 272–273 (♂♀).

Other citations. For complete list of taxonomic references, see WSC (2023).

Material examined. Ascension Island • 1♀ Lady Hill; -7.95, -14.37; pitfall trap baited with meat; 3rd March 2022; A. Sharp leg.; ASC F17 3 PFM • 1♂ Cricket Valley; -7.95, -14.34; pitfall trap; 24th March 2022; A. Sharp leg.; ASC K17 3 PFU • 1♀ North of Cocoanut Bay; -7.97, -14.34; pitfall trap baited with jam; 14th January 2022; A. Sharp leg.; ASC K19 3 PFJ • 1♂ Lower Valley Crater; -7.92, -14.34; subterranean pipe trap baited with blue cheese; 29th March 2023; A Sharp leg.; ASC LVC PTC • 1♂, 1♀; Boatswain Bird Island; -7.936, -14.307; hand collected; P. Ashmole and M. Ashmole leg.; ASC 1161 • 1♂ Ascension Island; -7.94, -14.37; collected between August-November 1957; E. A. Duffey leg.; No. 25; NHMUK.

Remarks. Invasive. Originally described from Tirol in Central Europe, but now known from all continents (WSC 2023). First recorded from Ascension Island by Duffey (1964) as *Zelotes rusticus*. Recent specimens confirm it is still to be found on the main island.

Zelotes laetus (O. Pickard-Cambridge, 1872)

Zelotes laetus: Levy 1998: 122, figs 58–61 (♂♀).

Other citations. For complete list of taxonomic references, see WSC (2023).

Material examined. Ascension Island • 1♂, 1♀; South Gannet Pools; -7.9865, -14.4015; 23–27 March 1990; Lava; P. Ashmole and M. Ashmole leg.; ASC 0830 • 1♂ Perfect Crater; -7.92, -14.36; pitfall trap baited with meat; 27th January 2022; A. Sharp leg. ASC H14 3 PFM • 1♂ Georgetown; -7.93, -14.41; pitfall trap baited with fish; 4th February 2022; A. Sharp leg.; ASC B15 2 PFF • 1♂ Dark Crater; -7.96, -14.39; pitfall trap; 15th February 2022; A. Sharp leg.; ASC E19 1 PFU • 1 imm. Ascension Island; -7.97, -14.39; 15.II.2022; superficial deposits; collected by hand; A. Sharp leg.; ASC E19 2 HC • 1 imm. Ascension Island; -7.93, -14.41; 04.II.2022; Intermediate Zr/Nb mafic flows (Younger flows); collected via litter extraction; A. Sharp leg.; ASC B15 1 LC • 1 imm. ♀, 1 imm.; Ascension Island; -7.96, -14.38; Low Zr/Nb mafic flows; collected by hand; A. Sharp leg.; ASC E19 3 HC • 1♂; North East Bay; -7.919, -14.344; 29.I.2013; pitfall trap; L. F. White leg.; ASC01117 • 1♂; same data as preceding except 28.II.2013; ASC01197 • 1♀; same data as preceding except 26.III.2013; ASC01354 • 1♂; same data preceding; ASC01376 • 1 imm.; same data as preceding except 21.V.2013; ASC01683 • 1 imm.; same data as preceding except 23.IV.2023; ASC01527 • 1♂; same data as preceding except 06.XII.2012; ASC00867 • 1♂ Ascension Island; -7.94, -14.37; collected between August-November 1957; E. A. Duffey leg.; No. 11; NHMUK.

Remarks. Invasive. Originally described by Pickard-Cambridge (1872) from 'Palestine' (modern Israel), known from West Palaearctic, introduced to Hawaii, USA, Mexico, and Peru (WSC 2023). First recorded on Ascension Island by Ashmole and Ashmole (1997) under the junior synonym *Zelotes inauratus* O. Pickard-Cambridge, 1872 (types in OUMNH examined by DS) although a male

was also found in the Duffey collection at NHMUK. Recently collected specimens confirm its continued presence on the main island.

Discussion

The gnaphosid fauna of Ascension Island, like most spider families present, is depauperate. Nonetheless, since records of some taxa given by Duffey (1964) had no precise locality, the study of more recent specimens from the 1990s and twenty first century allows us to provide the first precise locality records for some species on the island. Of note, Duffey (1964) also recorded *Gnaphosa funerea* (Dalmas, 1921) [using its original name, which is a homonym, *Pterochroa lugubris* O. Pickard-Cambridge, 1873] from Ascension Island but all of the relevant specimens labelled as this species in the Natural History Museum, London are immature gnaphosids of indeterminate genus (DS pers. obs.) and therefore *G. funerea* is hereby removed from the list of Ascension Island's spiders, as it cannot be confirmed from indeterminate juvenile specimens.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

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Author contributions

DS identified the specimens, produced plates, made diagnoses and descriptions, wrote the first draft of the manuscript and edited the revised maniscript. YMM identified specimens, produced plates, and edited the revised manuscript. AS collected specimens, provided locality and ecological data and edited the revised manuscript. PA collected specimens and provided locality and ecological data and edited the revised manuscript.

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Data availability

All of the data that support the findings of this study are available in the main text.

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African Invertebrates 64(3): 303–322 (2023)
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Research Article

Afrotropical *Atrichops* Verrall (Diptera, Athericidae) with description of a new species

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Abstract

Atrichops Verrall, 1909 of the Afrotropical Region are revised, and *Atherix adamastor* Stuckenberg, 1960, previously unplaced, is transferred to *Atrichops* based on morphological characters and feeding ecology. A new South African species, *A. intermedius* **sp. nov.** is also described from Mpumalanga Province.

Key words: haematophagy, new species, taxonomy, water snipe flies

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Introduction

Atrichops Verrall, 1909 is a haematophagous genus of athericine water snipe flies. Females of the genus are all known to feed on frogs (Nagatomi 1964). Presently there are 11 extant species of *Atrichops*, and one fossil species. Yang et al. (2016) most recently described three new species from China. Thomas and Gagneur (1982) described the first species of African *Atrichops*, *A. numidicus* from Algeria. Two years later (Nagatomi 1984b) described the first Afrotropical species *A. stuckenbergi* Nagatomi, 1984 from Kenya.

Note that there are at present five *incertae sedis* species of Athericidae known from South Africa (Stuckenberg 1980), previously placed in *Atherix*, and frequently still referred to as such. One of these five, *Atherix adamastor* Stuckenberg, 1960 has been informally placed in *Atrichops* by Picker et al. (2004: 274), and is also listed as such on data aggregators. The informal classification of this species can be largely attributed to Brian Stuckenberg. In his contributions to the "Field Guide to Insects of South Africa" by Picker et al. (2004), he categorised the species as an *Atrichops* when providing information on Athericidae. However, it is important to note that no formal taxonomic treatment or revision has been proposed for this species until the publication of this paper. Woodley (2017: 887) provides an identification key to the current Afrotropical genera of the family. We describe a new species *A. intermedius* sp. nov., which shares affinities with both *A. stuckenbergi* and *A. adamastor* comb. nov., from

Mpumalanga Province, South Africa. A key to these three species of Afrotropical *Atrichops* is also provided. A discussion about the apparent habitat and distribution is provided.

Materials and methods

Preparation methods

Morphological terminology follows that of Cumming and Wood (2017). Terminalia were macerated in heated 10% KOH for approximately 20 minutes or until clear, and examined using a Novel compound microscope with an attached Canon 850D DSLR camera. The same camera, with a 105 mm lens and extension tubes, was used for habitus photos of specimens. Specimen photos were stacked using Helicon Focus 7. Images and plates were prepared using Adobe Photoshop CC 2023 and Adobe Illustrator CC 2023. The species distribution map (Fig. 40) was generated using QGIS 3.28.6 and Africa Terrestrial Ecosystems (Sayre 2023).

Collections

The following collection codens are used:

BMSA National Museum, Bloemfontein, South Africa
CAS California Academy of Sciences, San Francisco, U.S.A.
ICIPE International Centre of Insect Physiology and Ecology, Nairobi, Kenya
MZLU Lund University, Entomology collections, Lund, Sweden
NMSA KwaZulu-Natal Museum, Pietermaritzburg, South Africa
OMNH Osaka Museum of Natural History, Osaka, Japan
SAMC Iziko South African Museum, Cape Town, South Africa

Any additions to materials examined are placed within square-brackets.

Results

Key to species of Afrotropical *Atrichops* Verrall

1 ♂ eyes appearing holoptic (Fig. 6), ♀ frons black shiny, with apex silver-white pruinose just above antennae (Fig. 12), face and clypeus with same silver-white pruinosity; ♂♀ eyes with only some sparse ommatrichia; proepimeral process short and blunt (Figs 21, 24) ..... ***A. stuckenbergi* Nagatomi**

– ♂ eyes narrowly dichoptic (e.g. Figs 4, 5), ♀ frons dark brown with at least some sparse pruinosity (e.g. Figs 10, 11), face and clypeus with some yellowish pruinosity; ♂♀ eyes densely covered in ommatrichia, proepimeral process knoblike, not well-developed **2**

2 ♂ frons as wide as anterior ocellus (Fig. 4), fore tibia with numerous conspicuous dorsal and ventral setulae longer than tibial diameter, gonocoxite with narrow base, parameral apodeme long (Fig. 31); ♀ face and clypeus separated by transverse emargination, but no apparent transverse

line; ♂♀ wing at most with only a slight dark marking over base of discal cell (Figs 25, 26) ..... ***A. adamastor* (Stuckenberg)**

– ♂ frons narrower than anterior ocellus (Fig. 5), fore tibia with sparse, widely spaced setulae at most as long as tibial diameter, gonocoxite with narrow base, parameral apodeme shorter (Fig. 32); ♀ face and clypeus separated by transverse emargination with clearly visible darkened transverse line; ♂♀ wing with apparent dark marking over base and apex of discal cell (Figs 27, 28) ..... ***A. intermedius* sp. nov.**

Taxonomy

Atrichops Verrall, 1909

Atrichops Verrall, 1909: 291. Type-species: *Atherix crassipes* Meigen, 1820, by monotypy.

Atherix (*Heterosuragina*) Nagatomi, 1958: 61. Type-species: *Atherix* (*Heterosuragina*) *fontinalis* Nagatomi, 1958, by original designation; synonymised by Nagatomi (1964: 69).

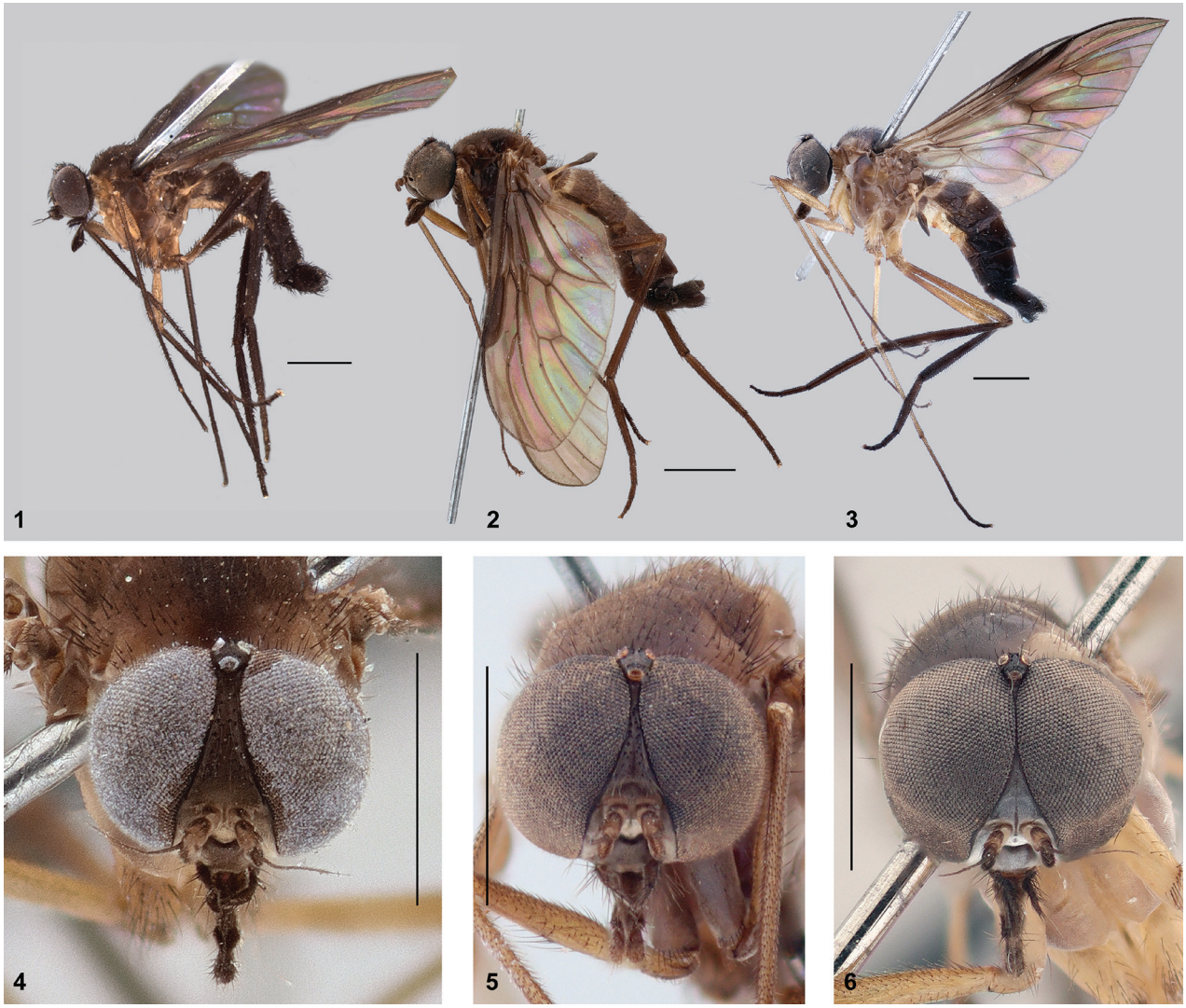
Diagnosis. *Atrichops* is similar in appearance to *Suragina* Walker, 1859 with elongated legs and hind coxa with a stout apical spine-like projection on its anteroventral surface. It is also haematophagous, with well-developed mandibles. It differs from *Suragina* by having a knob-like proepimeral process (also referred to as a ventrally projecting flap in non-Afrotropical species) (reduced in Afrotropical *Atrichops* species), antennal bases very close together (widely spaced in *Suragina*), face narrower than the frons in female, and tibial spur ratio of 0:1:2 (0:2:2 in *Suragina* – some specimens of *S. bivittata* (Bezzi, 1926) with ratio 1:2:2). *Atrichops* also has a uniform coloured frons, without the characteristic contrasting black and silver-grey of *Suragina*. *Atrichops* can be identified and distinguished from other Afrotropical genera using the generic key in Woodley (2017: 887), which is currently the most up to date key for the family in the Afrotropics.

Atrichops adamastor (Stuckenberg, 1960), comb. nov.

Figs 1, 4, 7, 10, 13, 16, 19, 22, 25, 26, 31, 34, 37

Atherix adamastor Stuckenberg, 1960: 273, fig. 86; Stuckenberg 1980: 313; Nagatomi 1984a: 94.

Identity. Stuckenberg (1960: 273) originally described *Atrichops adamastor* in *Atherix* Meigen, noting that its wings are "unlike those of any other South African species". However, it also differs from all other described southern African athericid species by possessing the key diagnostic characters of *Atrichops*, namely a combination of a ventrally projecting knob-like proepimeral process (albeit much reduced), having the face narrower than the frons in female, and antennal bases closely set, nearly touching. It also has wing vein R_{4+5} setulose dorsally, a characteristic only present in *Atrichops*, compared to other Afrotropical genera (see Stuckenberg 2000: 157). The male terminalia are also typical of

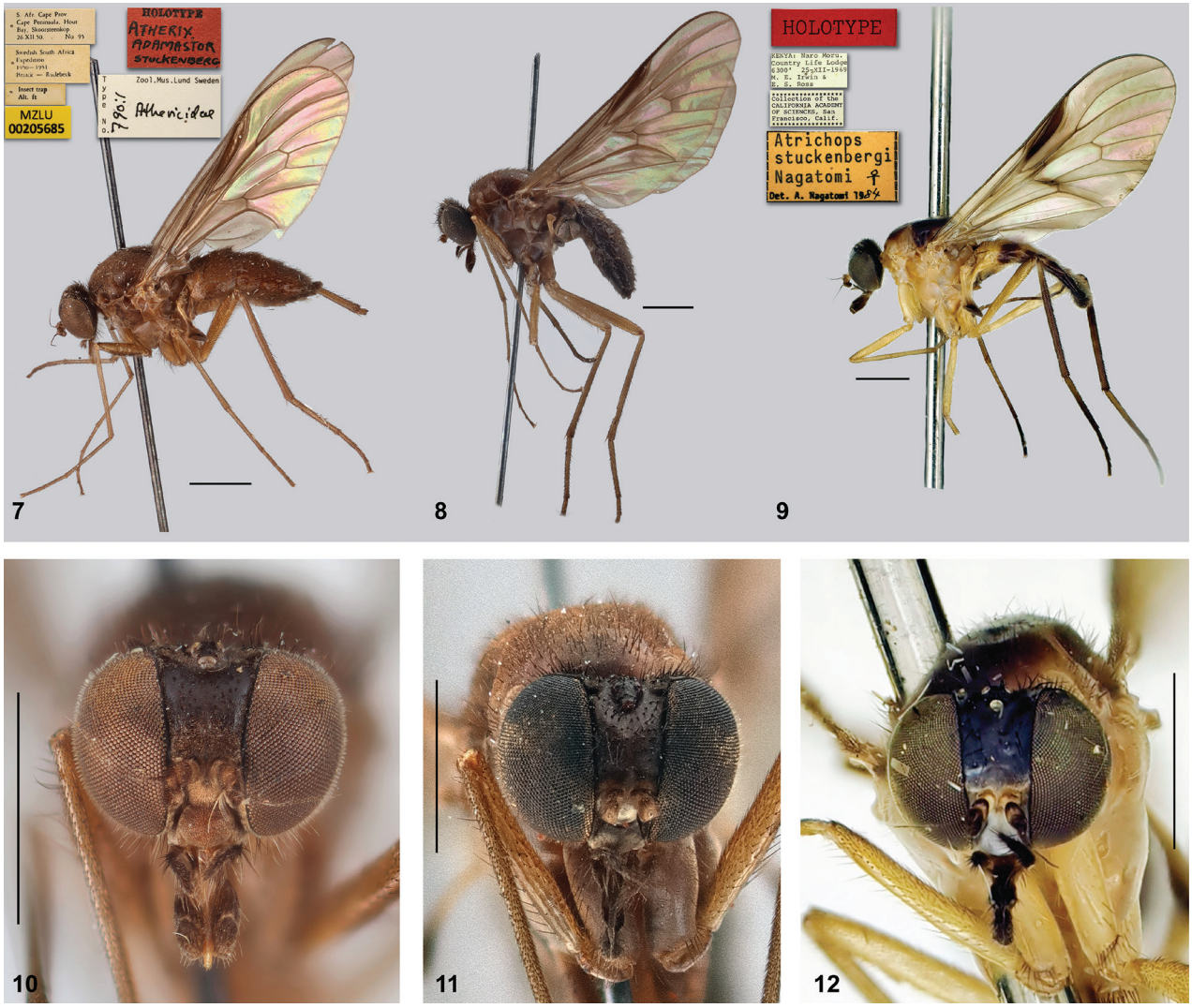


Figures 1–6. *Atrichops* spp. ♂ **1–3** lateral habitus **4–6** head anterior view **1** *A. adamastor* (BMSA(D)38908) **2** *A. intermedius* sp. nov. holotype (NMSA-Dip. 81808) **3** *A. stuckenbergi* (BMSA(D) 95848) **4** *A. adamastor* (BMSA(D)38904) **5** *A. intermedius* sp. nov. holotype (NMSA-Dip. 81808) **6** *A. stuckenbergi* (BMSA(D) 95848). Scale bars: 1 mm.

Atrichops species, with the gonostylus inserted medially on the dorsal surface of the gonocoxite (e.g., Figs 31–33), compared to other genera where it is inserted apically. *Atrichops adamastor* has also been observed blood-feeding on frogs (see biology discussion below), a behaviour typical of the genus.

Material examined. Type material examined (based on digital photos, additional data from Brinck and Rudebeck (1955: 77)): ***Holotype*:** South Africa • 1♀; Western Cape Province, Cape Peninsula, Hout Bay, Skoorsteenkop; [34°02.0684'S, 18°22.2420'E]; [300 ft]; 26 Dec. 1950; [Swedish South African Expedition leg.]; No. 95, type no. 00790:1, MZLU00205685 (see Fig. 1) (MZLU).

Additional material examined. South Africa • 2♂10♀; Western Cape Province, Gamkaskloof (Die Hel) at:; 33°21.808'S, 21°37.650'E; 336 m asl; 16–18 Oct. 2012; Kirk-Spriggs, A.H. leg.; Malaise trap in Karoo and valley *Acacia* woodland; (2♂: BMSA(D)38904, 38908; 10♀: BMSA(D)38899, 38900, 38901, 38902, 38903, 38905, 38906, 38907, 38909, 40027).



Figures 7–12. *Atrichops* spp. ♀ **7–9** lateral habitus **10–12** head anterior view **7** *A. adamastor* holotype and specimen labels (MZLU 00205685) **8** *A. intermedius* sp. nov. paratype (NMSA-Dip. 81809) **9** *A. stuckenbergi* (CAS) **10** *A. adamastor* holotype (MZLU 00205685) **11** *A. intermedius* sp. nov. paratype (NMSA-Dip. 81809) **12** *A. stuckenbergi* (BMSA(D)95856). (**7, 10** modified from originals provided by Rune Bygebjerg, MZLU, CC BY-NC 2.0 **9, 12** modified from originals provided by Rikio Matsumoto, Osaka Museum of Natural History). Scale bars: 1 mm.

Diagnosis. *Atrichops adamastor* can easily be distinguished from *A. stuckenbergi*. In *A. adamastor* the ommatrichia densely covers the eye in both sexes, the frons is dark brown, the face is brownish, and the wing is uniformly brown suffused, whereas *A. stuckenbergi* has the eye sparsely covered in ommatrichia, the frons is shiny black, the face has silver-white pruinosity, and its wing brown suffused mainly on apical half, with discal cell and cell m_3 less so (e.g., Fig. 26 vs Fig. 30). It is most similar to *A. intermedius* sp. nov., a South African contemporary (see *A. intermedius* diagnosis).

Remarks. The species was described by Stuckenberg (1960: 273), based only on a single female. The description is sufficient and no re-description is necessary. A description of its previously undescribed (but known) male follows. Mention is made of female characters should they differ from the male.

Figures 13–18. Atrichops spp. ♂♀ dorsal view **13** ♂ *A. adamastor* (BMSA(D)38908) **14** ♂ *A. intermedius* sp. nov. holotype (NMSA-Dip. 81808) **15** ♂ *A. stuckenbergi* (BMSA(D)94158) **16** ♀ *A. adamastor* holotype (MZLU 00205685) **17** ♀ *A. intermedius* sp. nov. paratype (NMSA-Dip. 81809) **18** ♀ *A. stuckenbergi* holotype (CAS). (**16** modified from original provided by Rune Bygebjerg, MZLU, CC BY-NC 2.0; **18** modified from original provided by Rikio Matsumoto, OMNH). Scale bars: 1 mm.

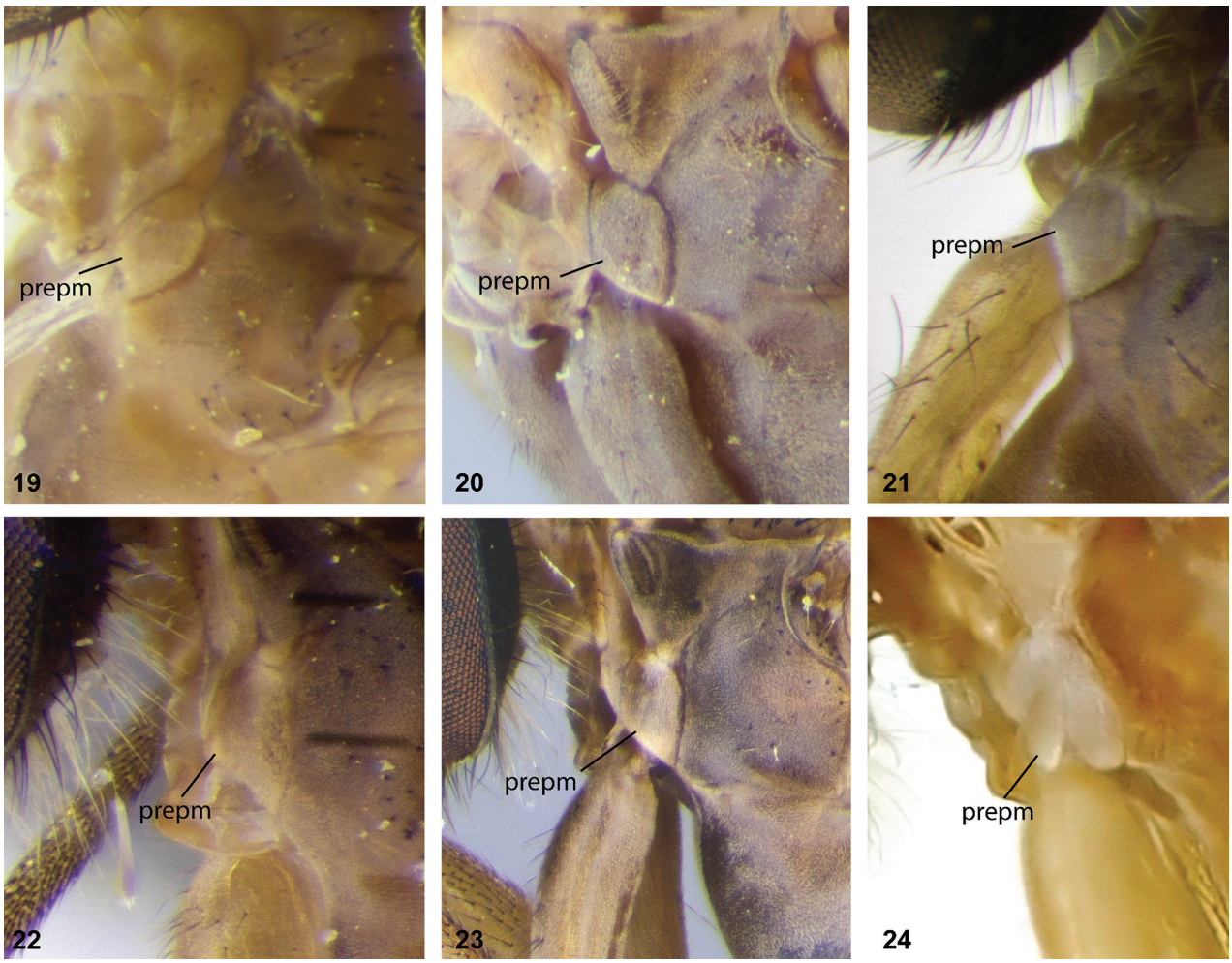
Description. ***Measurements*** (♂ n = 2, ♀ n = 10): Wing span: ♂ 4.8–5.2 mm (avg. 5.0 mm); ♀ 5.0–6.0 mm (avg. 5.4 mm); body length: ♂ 4.6 mm (avg. 4.6 mm); ♀ 3.7–4.9 mm (avg. 4.5 mm); wing span to body length ratio (avg.): ♂ 1.1; ♀ 1.2.

Male. ***Head*** (Fig. 4): Colour brown, with some light yellowish pruinosity on face; eye densely covered in ommatrichia; narrowly dichoptic, ♀ widely dichoptic; ommatidia on lower ½ of eye smaller than upper ½ (♀ ommatidia uniform,

comparatively larger than in ♂); lateral edge of eye with slight indentation, nearly absent in ♀; ocellar tubercle base level with frons, ♀ base with sunken appearance; ocellar tubercle with short dark setulae, vertex dark brown, almost appearing black, with somewhat longer setulae than ocellar tubercle; ocelli similar in size; ocellar tubercle in front of dorsal margin of eye, margin less indented than in ♀; vertex narrower than in ♀; dorsal inner edge of eye surrounding ocellar tubercle without discernible paired dark markings (present in ♀); occiput same dark brown as frons, when viewed at angle appearing shiny blackish; upper occiput with short dark setulae on dorsal margin and on rest of upper surface, lower occiput with erect, long and pale setulae, these continue ventrally on head to before mouthparts, bearing dark ventral setulae; frons dark brown, almost black, somewhat shiny, narrow area above antenna lighter brown; frons at narrowest as wide as anterior ocellus, widening towards antennal base, ♀ frons at least 2× width of ocellar tubercle, narrowing only slightly towards antennal base (Fig. 10); frons with well-developed dark setulae on surface, but only on lateral margins, very similar to ocellar and vertical setulae; ♀ frons more setulose than ♂; face lateral edges bare; gena bare; face light brown, clypeus darker brown, ♀ clypeus orange-brown; clypeus bare; face separated from clypeus by prominent transverse suture, in ♀ separated by transverse emargination, never forming suture; clypeus less prominent than in ♀; face much wider than in ♀, with prominent longitudinal emargination on side of clypeus, giving appearance that face bulges laterally; face and clypeus not visible in profile (visible in ♀); antennal bases close together, almost touching, gap somewhat larger than in ♀; scape brown, dorsally infuscate around setulae; pedicel comparatively darker than scape, dorsal surface infuscate; scape and pedicel setulae dark; 1st flagellomere reniform, appearing almost 2× height of pedicel, concolourous with pedicel, basal margins lighter brown; 2nd flagellomere arista-like, dark brown; scape and pedicel setulae similar in size; palpus dark brown, well-developed, ca 0.5× length of proboscis, with long, dark setulae; proboscis dark brown with orange-brown base.

Thorax (Figs 13, 19): Scutum with short dark setulae, posterior setulae longer than anterior setulae; scutellum with well-developed dark setulae; postpronotal lobe lighter brown than scutum, with fine dark setulae; scutum uniformly dark brown; scutellum uniformly dark brown; pleura generally brown in colour, with anepisternum, katepisternum, anatergite and katatergite somewhat darker; proepimeron with reduced flap-like process near anterior edge; notopleuron with well-developed dark setulae; area surrounding posterior spiracle dark brown, postspiracular scale dark brown almost black; proepisternum and pronotum yellow; anterior spiracle bare posteriorly; proepimeron, proepisternum bare, anepisternum with short dark setulae; katatergite with long pale setulae; rest of pleura bare; postmetacoxal bridge narrow.

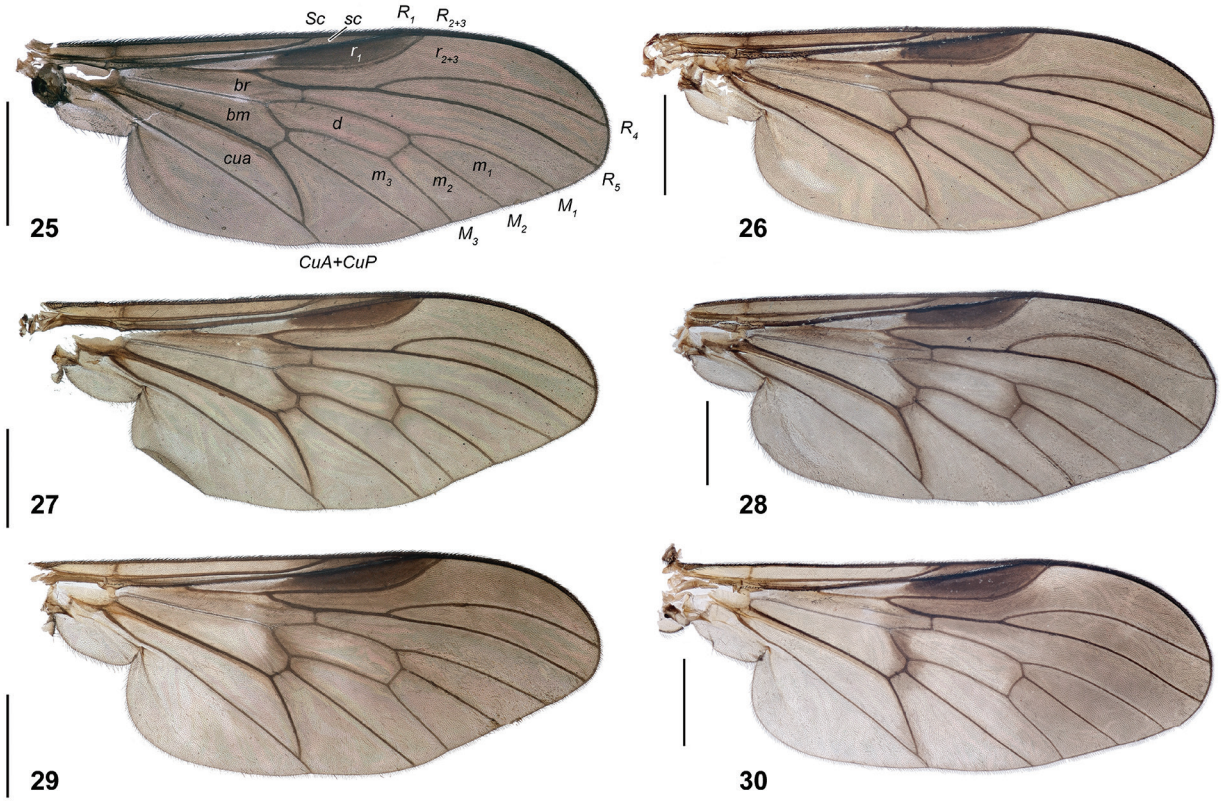
Legs (Fig. 1): Coxae yellow, anterior surface of mid- and hind coxae brown; fore coxa with short dark setulae on surface, more pronounced apically; mid coxa with long dark setulae on surface, hind coxa with dark setulae on anterior and lateral apical edges, and with well-developed anterior apical point; trochanters with some scattered short dark setulae, trochanters yellow, apical margins with darker markings, more pronounced than in ♀; fore and mid femora, and basal ¼ of hind femur yellow, apical ¾ of hind femur dark brown (♀ with all femora yellow); fore, mid and hind femora with small anterior apical dark mark;



Figures 19–24. *Atrichops* spp. ♂♀ proepimeral process, lateral view **19** ♂ *A. adamastor* (BMSA(D)38904) **20** ♂ *A. intermedius* sp. nov. paratype (NMSA-Dip. 81811) **21** ♂ *A. stuckenbergi* (BMSA(D)94158) **22** ♀ *A. adamastor* (BMSA(D)38899) **23** ♀ *A. intermedius* sp. nov. paratype (NMSA-Dip. 212861) **24** ♀ *A. stuckenbergi* holotype (CAS). (**24** modified from original provided by Rikio Matsumoto, OMNH). Abbreviations: prepm – proepimeral process.

fore, mid and hind tibiae dark brown (♀ with fore and mid tibiae yellow); all tarsi dark brown (♀ with yellow fore and mid tarsi); fore tarsal claws symmetrical, empodium at least 2× size of pulvilli on fore legs; fore femur covered with dark setulae on all surfaces, posteroventral setulae forming row, mid femur with row of longer anteroventral setae and hind femur with similar dark setulae on ventral and dorsal surfaces; hind leg stouter than fore and mid legs; fore tibia and tarsi densely covered with long setulae along dorsal and ventral surfaces, at least as long or longer than width of segments (♀ without long hairs); hind tarsomeres 1.01–1.02 (♂) and 0.87–1.02 (♀) times as long as hind tibia.

Wing (Fig. 25): Suffused brown on entire surface, with much darker pterostigma over area of veins R_1 and R_{2+3} and cell r_1; vein R_{4+5} with setulae dorsally, extending to vein R_5; veins dark brown; cell *br* and discal cell uniform in colour; costa with distinct downward flexure over pterostigma; cell *cua* closed at short distance from wing margin, cell m_3 open, veins M_1, M_2, M_3 present; halter with yellow stalk and dark knob in both sexes and ♀ holotype (Fig. 7). This differs from original description of *A. adamastor* ♀ holotype with Stuckenberg (1960: 274) noting "Halteres with pale yellowish knob."



Figures 25–30. *Atrichops* spp. ♂♀ wings dorsal view, *A. adamastor*: **25** ♂ (BMSA(D)38904) **26** ♀ (BMSA(D)38904); *A. intermedius* sp. nov.: **27** ♂ paratype (NMSA-Dip. 81811) **28** ♀ paratype (NMSA-Dip. 212861) *A. stuckenbergi*: **29** ♂ (BMSA(D)94161) **30** ♀ (BMSA(D)95861). Abbreviations: *br* – basal radial cell; *bm* – basal medial cell; *cua* – anterior cubital cell; *CuA+CuP* – anterior branch of cubital vein + posterior branch of cubital vein; *d* – discal cell; M_1 – first branch of media; m_1 – first medial cell; M_2 – second branch of media; m_2 – second medial cell; M_3 – third branch of media; m_3 – third medial cell; R_1 – anterior branch of radius; r_1 – first radial cell; R_{2+3} – second branch of radius; r_{2+3} – second radial cell; R_4 – upper branch of third branch of radius; R_5 – lower branch of third branch of radius; Sc – subcostal vein ; sc – subcostal cell. Scale bars: 1 mm.

Abdomen: Dark brown, with anterodorsal margins of tergites 1–3 as well as entirety of sternites 1–5 yellow; tergites and sternites with short dark setulae, longer dark setulae on lateral margins of tergites, as well as long pale setulae on sternites 1–3; tergite 1 without median suture.

Terminalia (Figs 31, 34): Epandrium and cercus dark brown; hypoproct dark brown; epandrium, hypandrium and cercus with dark setulae; gonostylus finger-like with rounded apex; gonocoxite narrowing from middle towards base, apically rounded with long setulae; parameral apodeme long, extending to base of gonocoxite in ventral view; gonocoxal apodeme markedly shorter than gonocoxite.

Female. *Terminalia:* Stuckenberg (1960) did not describe proepimeral process, which is reduced flap-like process (Fig. 22), much as in ♂. Cercus dark brown with dark setulae; genital fork (Fig. 37) with slender distal apodeme, median lobe with deep apical emargination, paired apical lobes with somewhat square appearance, arms each gradually rounded; 3 sclerotised, somewhat oblong, spermathecae.

Distribution. South Africa (Western Cape Province).

Biology. The species was observed and photographed feeding on the Cape River Frog, *Amieta fuscigula* Duméril & Bibron in the Garden Route Botanical Garden, George, South Africa, by Colin Ralston (iNaturalist observation 9344668). While this behaviour has been informally observed before, it is the first published photographic observation of the species exhibiting typical *Atrichops* behaviour. It also emphasizes the importance of citizen science in highlighting species interactions that might otherwise go unrecorded or unnoticed. Although an attempt was made to sample material from the Botanical Garden for study, it was unsuccessful, possibly due to heavy rains earlier in the season.

***Atrichops intermedius* Muller, sp. nov.**
https://zoobank.org/B3B59959-1502-472A-89C2-C21017DE04D1
Figs 2, 5, 8, 11, 14, 17, 20, 23, 27, 28, 32, 35, 38

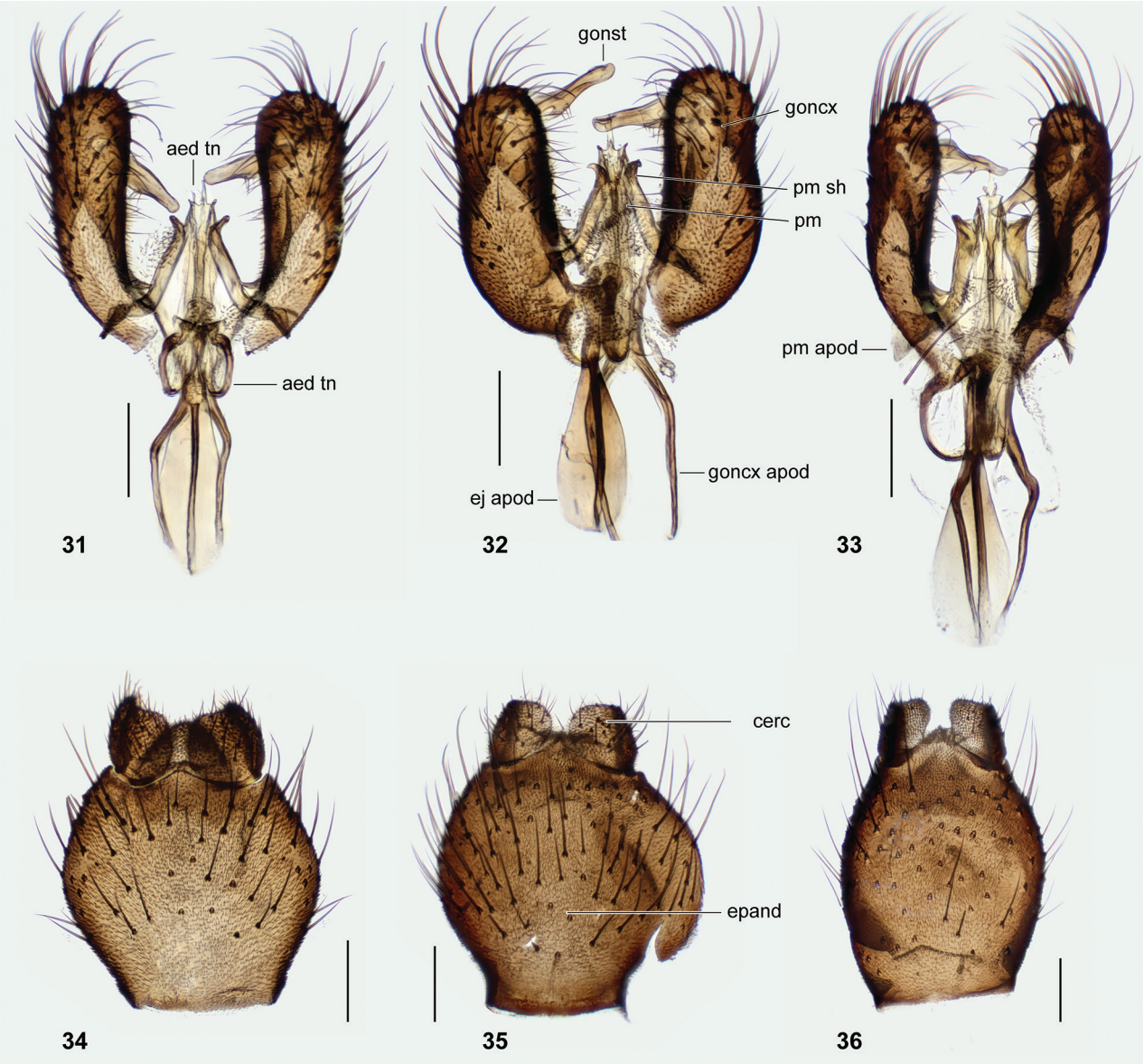
Material examined. Type material: *Holotype*: South Africa • 1♂; Mpumalanga, Gladdespruit River nr Nelspruit [Mbombela] airfield, Transvaal; [25°30.6703'S, 30°54.4575'E]; 2530Db; 2975 ft [907 m asl]; 23 Feb. 1971; Stuckenberg [Stuckenberg, B.R] leg.; streamside bush; (NMSA-DIP 81808) (NMSA).

***Paratypes*:** South Africa • 2♂2♀; Mpumalanga, Gladdespruit River nr Nelspruit [Mbombela] airfield, Transvaal; [25°30.6703'S, 30°54.4575'E]; 2530Db; 2975 ft [907 m asl]; 23 Feb.1971; Stuckenberg [Stuckenberg, B.R] leg.; streamside bush; (2♂ NMSA-DIP 81811, NMSA-DIP 28122; 1♀ NMSA-DIP 81809) (NMSA). 1♀; Mpumalanga, Graskop, [24°55.5480'S, 30°49.7993'E]; 6 Mar. 2000; Picker, M. leg.; (NMSA-DIP 212861) (NMSA).

Diagnosis. This species is most similar to *A. adamastor*. However, it differs from *A. adamastor* in several ways: ♂ more narrowly dichoptic, ♀ with a darkened transverse line between the face and clypeus, ♂ legs nearly unicolorous yellowish compared to both males of *A. adamastor* and *A. stuckenbergi*, which have yellow hind femora and all tibiae and tarsi darker; Additionally, *A. intermedius* has wings with darker brown suffusions over base and apex of the discal cell that are similar to *A. stuckenbergi*, in contrast to *A. adamastor* which has a more uniformly suffused wing. Its gonocoxite is also more rounded and less tapering toward base compared to both *A. adamastor* and *A. stuckenbergi*.

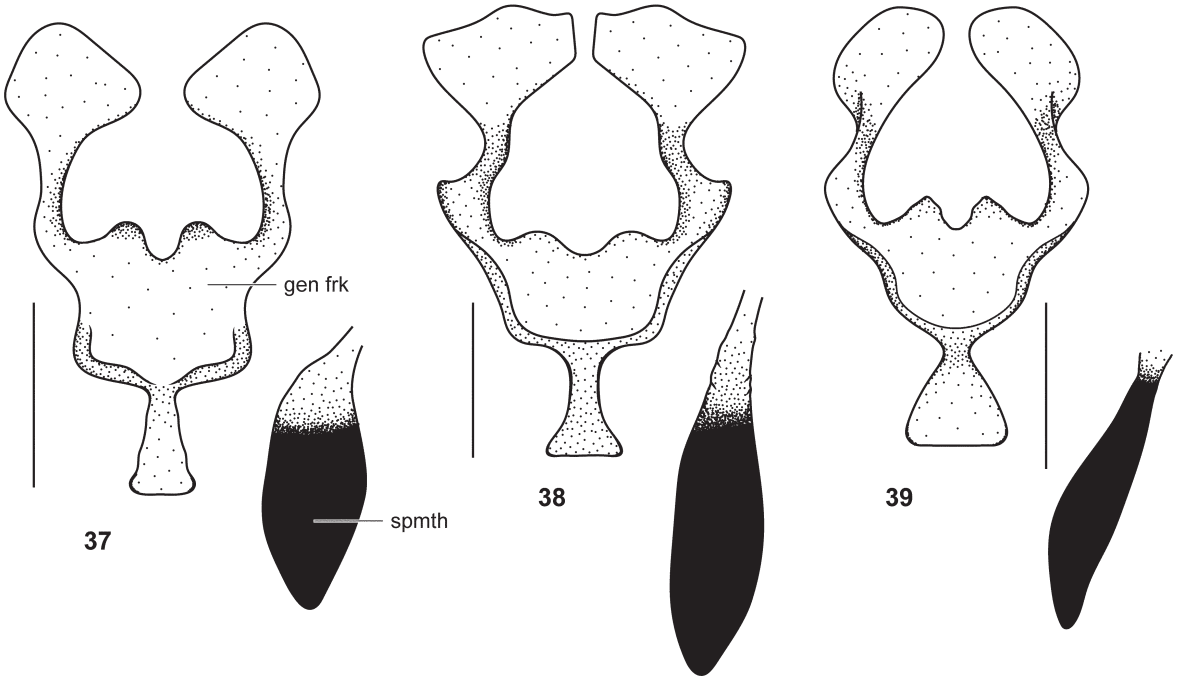
Description. ***Measurements*** (♂ n = 2, ♀ n = 2): Wing span: ♂ 5.4–5.8 mm (avg. 5.6 mm); ♀ 5.9–6.5 mm (avg. 6.2 mm); body length: ♂ 5.5 mm–6.1 mm (avg. 5.8 mm); ♀ 4.8–5.4 mm (avg. 5.1 mm); wing span to body length ratio (avg.): ♂ 0.96; ♀ 1.2.

Male. ***Head*** (Fig. 5): Colour dark brown, with some yellowish pruinosity on face; eye densely covered in ommatrichia; narrowly dichoptic; ommatidia on lower ½ of eye smaller than upper ½; lateral edge of eye with indentation (absent in ♀); ocellar tubercle base level with frons, ocellar tubercle with short dark setulae; vertex dark brown, almost appearing black, with markedly longer setulae than ocellar tubercle, similar to that of frons; ocelli similar in size; ocellar tubercle in front of dorsal margin of eye, margin less indented than in ♀; vertex narrower than in ♀; dorsal inner edge of eye, surrounding ocellar tubercle without discernible paired dark markings; occiput same dark brown as frons, when viewed at angle appearing shiny blackish; upper occiput with short dark



Figures 31–36. *Atrichops* spp., 31–33 ♂ terminalia **31** *A. adamastor* (BMSA(D)38904) **32** *A. intermedius* sp. nov. paratype (NMSA-Dip. 81811) **33** *A. stuckenbergi* (BMSA(D) 94160), 34–36 ♂ epandrium and cerci **34** *A. adamastor* (BMSA(D)38904) **35** *A. intermedius* sp. nov. paratype (NMSA-Dip. 81811) **36** *A. stuckenbergi* (BMSA(D) 94160). Abbreviations: aed tn – aedeagal tine; cerc – cercus; ej apod – ejaculatory apodeme; epand – epandrium; goncx – gonocoxite; goncx apod – gonocoxal apodeme; gonst – gonostylus; pm – paramere; pm apod – parameral apodeme; pm sh – parameral sheath.Scale bars: 0.15 mm.

setulae on dorsal margin and on rest of upper surface, lower occiput with erect, long and pale setulae, these continue ventrally on head to before mouthparts bearing dark ventral setulae; frons dark brown, almost black, narrow area above antenna lighter brown; frons at narrowest slightly narrower than anterior ocellus, widening towards antennal base; frons with well-developed dark setulae on surface except for narrow area above antennae, these similar to ocellar and vertical setulae; frons less setulose than ♀; face lateral edges bare; gena bare; face light brown, clypeus darker brown; clypeus bare; face separated from clypeus by prominent, deep, dark transverse suture; clypeus less prominent than in ♀; face much wider than in ♀, with prominent longitudinal emarginations on



Figures 37–39. *Atrichops* spp., ♀ genital fork and spermatheca **37** *A. adamastor* (BMSA(D)38899 **38** *A. intermedius* sp. nov. paratype (NMSA-Dip. 212861) **39** *A. stuckenbergi* (BMSA(D) 95855). Abbreviations: gen fk – genital fork; spmth – spermatheca. Scale bars: 0.15 mm.

sides of clypeus, giving appearance that face bulges laterally; face and clypeus not visible in profile; antennal bases close together, almost touching; scape light-brown, dorsally infuscate around setulae; pedicel comparatively darker than scape, dorsal surface infuscate; scape and pedicel setulae dark; 1st flagellomere reniform; appearing almost 2× the height of pedicel, same colour as pedicel, basal margins lighter brown; 2nd flagellomere arista-like, dark brown; scape and pedicel setulae similar in size, palpus brown, well-developed, ca 0.5× length of proboscis; proboscis comparatively smaller than ♀ proboscis in relation to head; proboscis dark brown with long dark setulae.

Thorax (Figs 14, 20): Scutum with short dark setulae, postsutural setulae longer than presutural setulae; scutellum with well-developed dark setulae; postpronotal lobe lighter brown than scutum, with fine dark setulae; scutum and scutellum uniformly dark brown when viewed dorsally, scutellum similar in colour as anepimeron, anatergite and katatergite when viewed laterally; pleura generally brown in colour, anepimeron, katatergite and anatergite lighter brown than rest of pleura (more pronounced in ♀); proepimeron with reduced knoblike process near anterior edge; notopleuron with well-developed dark setulae; area surrounding posterior spiracle dark brown, postspiracular scale dark brown almost black; proepisternum brown and pronotum yellow; anterior spiracle bare posteriorly; proepimeron and proepisternum bare, anepisternum with short dark setulae; katatergite with long pale setulae; rest of pleura bare; postmetacoxal bridge narrow.

Legs (Fig. 2): Coxae brown; fore coxa with short black setulae on surface, more pronounced apically, mid coxa with long black setulae on surface, hind coxa with black setulae on anterior and lateral apical edges, with well-developed anterior apical point; trochanters brownish yellow with some scattered

short dark setulae; hind trochanter somewhat lighter than rest; apical margins of trochanters with darker markings; fore and mid femora, and basal ¼ of hind femur yellow, apical ¾ of hind femur dark brown; fore, mid and hind femora with small anterior apical dark mark; fore, mid and hind tibiae yellow; tarsi dark brown; fore tarsal claws symmetrical, empodium at least 2× size of pulvilli on fore legs; fore femur covered with dark setulae on all surfaces, posteroventral setulae forming row, mid femur with row of longer anteroventral setae and hind femur with similar dark setulae on ventral and dorsal surfaces; hind leg stouter than fore and mid legs; fore tarsi densely covered with long setulae along dorsal and ventral surfaces, at least as long as width of segment, similar setulae on fore tibia, although much sparser; hind tarsal segments 0.9–1.1 (♂) as long as hind tibia.

Wing (Fig. 27): Suffused brown on entire surface, with much darker pterostigma over area of veins R_1 and R_{2+3} and cell r_1; veins light brown; cell *br* and discal cell darker suffused apically; costa with distinct downward flexure over pterostigma; cell *cua* closed short distance from wing margin, cell m_3 open, veins M_1, M_2, M_3 present; halter with stalk yellow on basal half, gradually darkening apically with dark brown knob.

***Abdomen*:** Brown, with anterodorsal margins of tergites 2–4 brownish yellow; tergites and sternites similar in colour with short black setulae, except sternite 1 lighter in colour; tergites with longer black setulae laterally; long pale setulae on sternites 1–3; tergite 1 without median suture.

***Terminalia*:** Epandrium and cerci dark brown, hypoproct dark brown; epandrium, hypandrium and cercus with dark setulae; gonostylus finger-like, narrowing slightly towards apex, gonocoxite widening from middle towards base, apically rounded with long setulae; parameral apodeme short, not reaching base of gonocoxite in ventral view; gonocoxal apodeme markedly shorter than gonocoxite.

Female. Similar characters as ♂ except for the following:

Head (Fig. 11): Widely dichoptic, ommatidia of similar size, comparatively larger than in ♂; lateral edge of eye without indentation (present in ♂); ocellar tubercle base with sunken appearance; ocellar tubercle with slightly longer setulae than in ♂; dorsal margin of eye more indented than in ♂; vertex wider than in ♂; dorsal inner edge of eye, surrounding ocellar tubercle with paired dark markings; frons at least 2× width of ocellar tubercle, narrowing only slightly towards antennal base; frons more setulose than ♂; face separated from clypeus by transverse emargination with darkened transverse line; clypeus more prominent than in ♂; face and clypeus visible in profile; 1st flagellomere smaller in comparison to pedicel compared to ♂; 2nd flagellomere ca 0.75× length of proboscis; proboscis comparatively larger than ♂ proboscis in relation to head.

Legs (Fig. 8): Trochanters yellow; apical margins of trochanters with darker markings, less pronounced than in ♂; all femora almost entirely yellow, except for small anterior apical dark marking on each femur; tarsi yellow; fore tarsi without prominent long setulae on dorsal and ventral surfaces; hind tarsal segments 0.8–1.0 (♀) times as long as hind tibia.

Wing (Fig. 28): Some specimens with cell *br* and majority of discal cell surface hyaline.

***Abdomen*:** Darker overall colour, similar anterodorsal margin and setation colour, setulae shorter overall.

***Terminalia*:** Cercus dark brown with dark setulae; genital fork (Fig. 38) with distal apodeme slender, median lobe with shallow apical emargination, paired apical lobes with angular appearance, arms each having angular projection; 3 sclerotised, elongated spermathecae.

Etymology. From the Latin "*intermedius*", meaning in the middle, referring to the species known occurrence being roughly between the distributions (Fig. 40) of *A. adamastor* and *A. stuckenbergi*, but also in reference to *A. intermedius* exhibiting a mixture of characteristics found in both the other two Afrotropical species.

Distribution. South Africa (Mpumalanga).

***Atrichops stuckenbergi* Nagatomi, 1984**
Figs 3, 6, 9, 12, 15, 18, 21, 24, 29, 30, 33, 36, 39

Atrichops stuckenbergi Nagatomi 1984b: 21, figs 3, 10.

Material examined. *T*ype material (based on digital photos): *Holotype*: Kenya • 1♀; Naro Moru, Country Life Lodge; [0°09.3224'S, 37°0.7380'E]; 6300 ft; 25 Dec. 1969; Irwin, M.E. & Ross, E.S. leg.; (CAS). ***Paratype*:** 1♀; Naro Moru, Country Life Lodge; [0°09.3224'S, 37°0.7380'E]; 6300 ft; 25 Dec. 1969; Irwin M.E. & Ross, E.S. leg.; (CAS). Material at time of publication is in the Nagatomi collection of OMNH and will be returned to CAS shortly by the curator in charge at OMNH (pers. comm. B. Sinclair)

Additional material examined. Malawi • 5♂; Southern Region; Mulanje Mountain [Mulanje Massif] at:; 15°56.1667'S, 35°31.1982'E; 1061 m asl; 12–14 Oct. 2016, Kirk-Spriggs, A.H. & Muller, B.S. leg.; Malaise trap, stream bed; Miombo woodland; (5♂ BMSA(D) 97872, 97873, 97874, 97875, 97876). 18♂11♀; Zomba Plateau, Kuchawe Trout Farm; 15°21.2315'S, 35°18.0954'E; 1530 m asl; 8–11 Oct. 2016; Kirk-Spriggs, A.H. & Muller, B.S. leg.; Malaise trap, stream, montane evergreen forest; (11♀ BMSA(D)94146, 94147, 94151, 94153, 94154, 94155, 94158, 94162, 94163, 94165, 94171, 94172, 95851; 18♂ BMSA(D)94142, 94143, 94144, 94145, 94148, 94149, 94150, 94152, 94156, 94157, 94159, 94160, 94161, 94164, 94166, 94167, 94168, 94169, 94170). 5♀8♂; Zomba Plateau, William's Falls; 15°20.8392'S, 35°17.9158'E; 1583 m asl; 15–19 Oct. 2016, Kirk-Spriggs, A.H. & Muller, B.S. leg.; Malaise trap, stream, montane evergreen forest; (8♀ BMSA(D) 95850, 95852, 95853, 95855, 95856, 95857, 95858, 95861). 5♂ BMSA(D)95848, 95849, 95854, 95859, 95860). 1♂; Nyasaland, Cholo [Thyolo], [16°4.2204'S, 35°8.178'E], 14 Sept.1917, Wood, R.C. leg.; (1♂ NMSA-Dip. 158775). Kenya • 1♀; Central Province; Mary Mariuki farm, near Nyamindi River; 00°29.5500'S, 37°23.2932'E; 1457 m asl; 2–16 Jun. 2018, Copeland, R. leg.; Malaise trap; (1♀ ICIPE 3773).

Photographic material. (Scharff et al. 2023): Tanzania • 1♂; Uzungwe [Udzungwa] Mts., Mwanihana Forest above Sanje; [07°46.0000'S, 36°49.0000'E]; 1000 m asl; 1 Aug.1981; Stolze, M. & Scharff, N. leg.; Zool. Museum, Copenhagen; (1♂ ZMUC 100826, GBIF ID: 857827875). 1♀; East Usambara Mts., Amani; [05°05.9833'S, 38°37.9834'E]; 1000 m asl; 10 Jul. 1980, Stoltze, M. & Scharff, N. leg.; Zool. Museum, Copenhagen (1♀ ZMUC 100827, GBIF ID: 857829273).

Note. The Tanzanian specimens are only identified as *Atrichops* sp. on GBIF, but agree with the descriptions and appearance of male and female *A. stuckenbergi*.

Diagnosis. *Atrichops stuckenbergi* can be distinguished from the only other two Afrotropical species, *A. adamastor* and *A. intermedius*, by having its eye sparsely covered in ommatrichia, whereas the others have ommatrichia densely covering the eye. It has the frons shiny black, compared to the frons being dark brown in *A. adamastor*. The wings in *Atrichops stuckenbergi* are brown suffused mainly on the apical half, with the discal cell and cell m_3 less so, much like in *A. intermedius*, compared to *A. adamastor* that has uniformly brown suffused wings. *Atrichops stuckenbergi* differs also from both the other species in the shape of its ♂ and ♀ terminalia (Figs 33, 36, 39).

Remarks. The species was described by Nagatomi (1984b), based on two female specimens. The description is sufficient and no re-description is necessary. The specimens from Malawi were compared to the holotype female (digital photographs) and a non-type female from Kenya and all fit Nagatomi's description of *A. stuckenbergi*. Mention is made of female characters below should they differ from the male. A description of the previously unknown male, based on material from Kenya and Malawi, follows.

Description. ***Measurements*** (♂ n = 5, ♀ n = 5): Wing span: ♂ 5.0–5.7 mm (avg. 5.5 mm); ♀ 5.3–6.5 mm (avg. 5.9 mm); body length: ♂ 5.6–6.2 mm (avg. 5.9 mm); ♀ 3.9–5.2 mm (avg. 4.6 mm); wing span to body length ratio (avg.): ♂ 0.9; ♀ 1.3.

Male. ***Head*** (Fig. 6): Colour dark brown, with silver-white pruinosity on face and clypeus (♀ with similar colour, only with somewhat less pruinosity); eye sparsely covered in ommatrichia, difficult to discern; holoptic, ♀ widely dichoptic (Fig. 6); ommatidia on lower ½ to ⅓ of eye smaller than upper area (♀ ommatidia uniform, comparatively larger than in ♂); lateral edge of eye with prominent indentation, almost absent in ♀; ocellar tubercle with base level with frons; ocellar tubercle with long dark setulae (compared to vertex and occipitals), vertex glossy dark brown, almost black, with shorter setulae than ocellar tubercle; ocelli similar in size; ocellar tubercle in front of dorsal margin of eye, eye margin less indented than in ♀; vertex narrower than in ♀; dorsal inner edge of eye touching ocellar tubercle, without discernible paired dark markings (present in ♀); occiput dark brown with some whitish pruinosity; upper occiput with short dark setulae on dorsal margin and on rest of upper surface, lower occiput with erect, long and pale setulae, continuing ventrally on head to before mouthparts bearing dark ventral setulae; frons shiny black, without any setulae, area just above antennae with silver-white pruinosity; frons widening towards antennal base, ♀ frons at least 2× width of ocellar tubercle, narrowing only slightly towards antennal base; frons bare compared to well-developed setulae on ♀ similar to ocellar tubercle and vertex; face lateral edges bare; gena bare; face separated from clypeus by prominent transverse suture, in ♀ separated by transverse emargination, but never forming suture; clypeus less prominent than in ♀; face much wider than in ♀, with prominent longitudinal emarginations on sides of clypeus, almost giving appearance that face bulges laterally; face and clypeus not visible in lateral view, visible in ♀; antennal bases close together, not touching as in ♀, separated by short, dark longitudinal vitta; scape and pedicel orange-yellow, brown dorsally around setulae; scape and pedicel setulae dark; 1st flagellomere reniform, appearing almost 2× height of pedicel, same colour as pedicel, basal margins lighter brown; 2nd flagellomere arista-like, dark brown; scape and pedicel setulae similar in size, palpus dark brown, basally

orange-yellow, well-developed, ca 0.5× length of proboscis, with long dark setulae; proboscis dark brown with orange-brown base.

Thorax (Figs 15, 21): Scutum with short dark setulae, postsutural setulae longer than presutural setulae; scutellum with well-developed dark setulae; scutum and scutellum uniformly dark brown (♀ holotype with scutellum yellow, darker in ♀ specimens from Malawi); pleura generally yellowish brown in colour, with anepisternum, katepisternum, anatergite and katatergite somewhat darker; proepimeron grey-white pruinose, with somewhat reduced knob-like process near anterior edge, not as apparent as in non-Afrotropical species; notopleuron with well-developed dark setulae; area surrounding posterior spiracle yellow, postspiracular scale dark brown, yellow in ♀; pronotum, postpronotal lobe and postalar callus yellow, all with fine dark setulae; anterior spiracle bare posteriorly; proepimeron, proepisternum bare, anepisternum and katatergite with pale setulae; rest of pleura bare; postmetacoxal bridge narrow.

Legs (Fig. 3): Coxae yellow; fore coxa with short dark setulae on surface, more pronounced apically; mid coxa with long dark setulae on surface, hind coxa with dark setulae on anterior and lateral apical edges, with well-developed anterior apical point; trochanters with some scattered short dark setulae, trochanters yellow, apical margins with darker markings, similar to that of ♀; fore, mid and base and apex of hind femora yellow, hind femur with dark brown middle; fore, mid, and hind femora with small anterior apical dark marking; fore and mid tibiae yellow, hind tibia dark brown; all tarsi dark brown (♀ with yellow mid and hind tarsi); fore tarsal claws symmetrical, empodium at least 2× size of pulvilli on fore legs; fore femur covered with dark setulae on all surfaces, posteroventral setulae forming row, mid femur with row of longer anteroventral setae and hind femur with similar dark setulae on ventral and dorsal surfaces; hind leg stouter than fore and mid legs; fore tibia and tarsi densely covered with long hairs along dorsal and ventral surfaces, at least as long or longer than width of segments (♀ with long hairs only on ventral margins of tarsi); hind tarsal segments 0.94–1.01 (♂) and 0.92–1.00 (♀) times as long as hind tibia.

Wing (Fig. 29): Suffused brown on most of apical half, with much darker pterostigma over area of veins R_1 and R_{2+3} and cell r_1, dark band across area near base of discal cell; majority of discal cell and cell *bm* hyaline; cell *cua* closed short distance from margin, cell m_3 open, veins M_1, M_2, M_3 present; vein R_{4+5} with setulae dorsally, extending to vein R_5; veins dark brown, costa with much weaker downward flexure over pterostigma than in ♀; halter with yellow stalk and dark knob.

Abdomen (Fig. 15): Dark brown, tergites 1–3 with anterodorsal margins yellow (t2 almost appearing triangular) (♀ holotype tergites 1–5 with anterodorsal margins yellow, giving triangular shaped appearance), entirety of sternites 1–5 yellow; tergites with short dark setulae, lateral margins with longer black setulae, more so than ♀; pale setulae on sternites 1–5; tergite 1 without median suture.

Terminalia (Figs 33, 36): Epandrium and cercus dark brown, hypoproct dark brown; epandrium, hypandrium and cercus with dark setulae; gonostylus finger-like, narrowing with prominent sinuous curve towards apex, gonocoxite narrowing from middle towards base, apically rounded with long setulae; parameral apodeme elongate, appearing enlarged, reaching base of gonocoxite in ventral view, aedeagal tine enlarged compared to *A. adamastor* and *A. intermedius*; gonocoxal apodeme similar in length to gonocoxite.

Female. ***Terminalia*** cercus dark brown with dark setulae; genital fork (Fig. 39) with distal apodeme enlarged, median lobe with deep apical emargination, paired apical lobes with rounded appearance, arms each rounding gradually; 3 sclerotised, elongated spermathecae.

Distribution. Kenya, Malawi (new record), Tanzania (new record).

Discussion

The three Afrotropical species are currently known from only four countries in Africa (Fig. 40) and inhabit a minimum of three distinct ecosystems, based on Mucina and Rutherford (2006) and Sayre (2023): *A. adamastor* inhabits

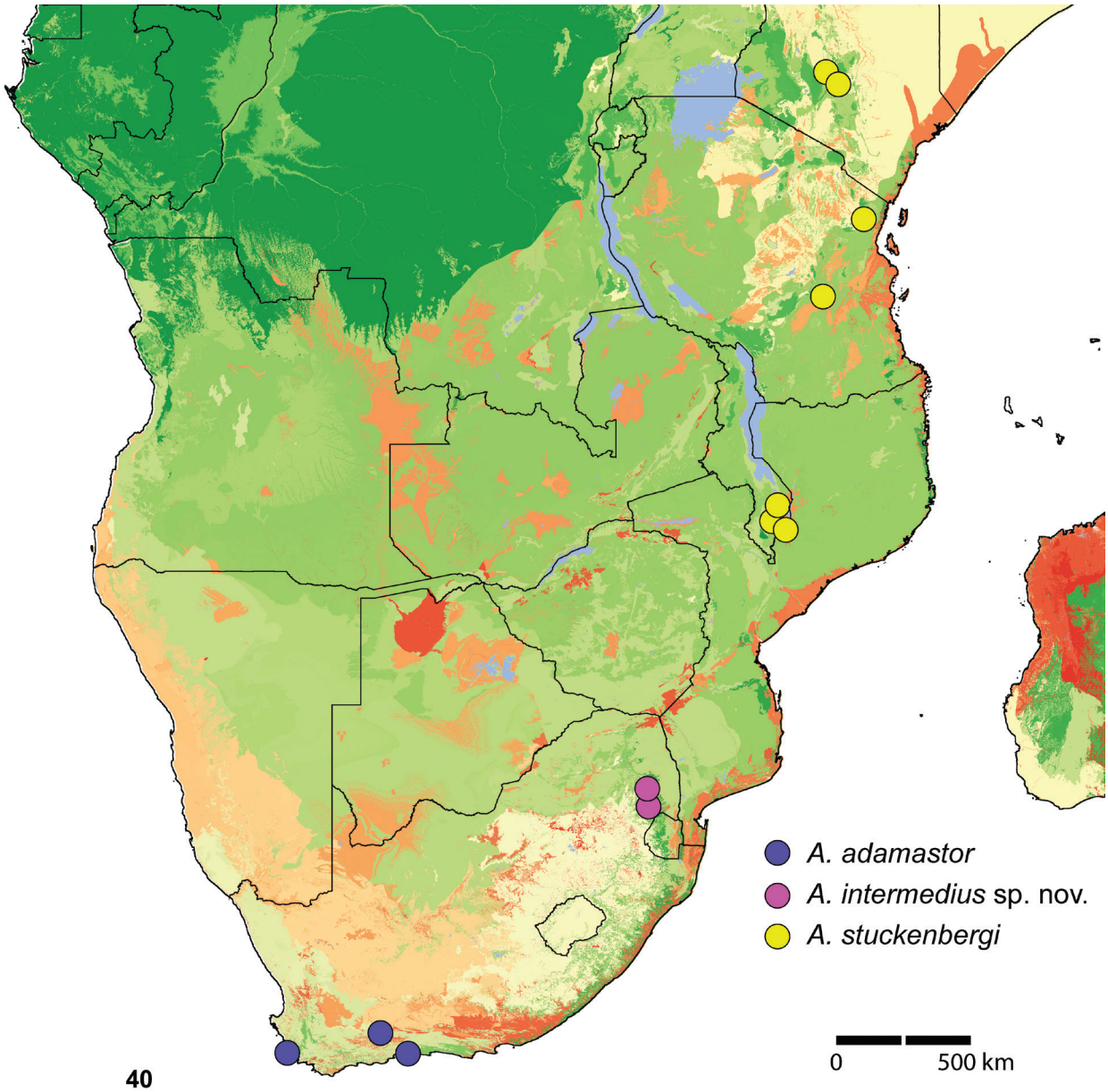


Figure 40. Distribution of *Atrichops* spp. in the Afrotropical Region. Map generated using Africa Terrestrial Ecosystems data (Sayre 2023).

Afromontane forest pockets within the Fynbos biome, but also in adjacent Albany thicket and Succulent Karoo, even in the absence of forest. Brinck and Rudebeck (1955: 77) described the area at 600 ft elevation near the type locality as "dense indigenous wood", with the surroundings as "fairly dry heath with Proteas, and to the east, in a depression with luxuriant vegetation". *Atrichops intermedius* inhabits Savanna and Grassland environments, often featuring similar Afromontane forest pockets. On the other hand, *A. stuckenbergi* has thus far only been found in Afromontane forest of the Eastern Arc Mountains of Kenya and Tanzania, and forests of the Mulanje massif and other mountains in the Southern Region of Malawi. Given the presumed biennial lifecycle of the larvae in the genus, supported by observations of *A. crassipes* by e.g., Gerke and Böttger (2001), it is reasonable to assume that the overall health of water systems and associated environmental conditions in rivers or streams play a crucial role in determining suitable distribution. This includes factors like the availability of prey for larvae and the presence of frog hosts for adult females. Future research into the biology of not only *Atrichops* but also other Athericidae in the Afrotropical Region should consider these aspects. This is especially important in light of the increasing habitat loss in the region, as noted by Chapman et al. (2022). As water snipe flies serve as vital indicators of river health (Dickens and Graham 2002), a better understanding of the group's tolerances is essential. This information can empower relevant stakeholders to make informed decisions in conservation management and planning efforts across Africa.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

BSM confirms ethical clearance, number NMB ECC 2019/1, forming part of project 457 of the National Museum, Bloemfontein, South Africa.

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Author contributions

Muller conceptualised the project with inputs from Swart and Snyman. Muller identified, described and revised the species, and wrote the first draft of the manuscript. Snyman and Swart commented and made additions to the final draft.

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Data availability

All of the data that support the findings of this study are available in the main text.

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